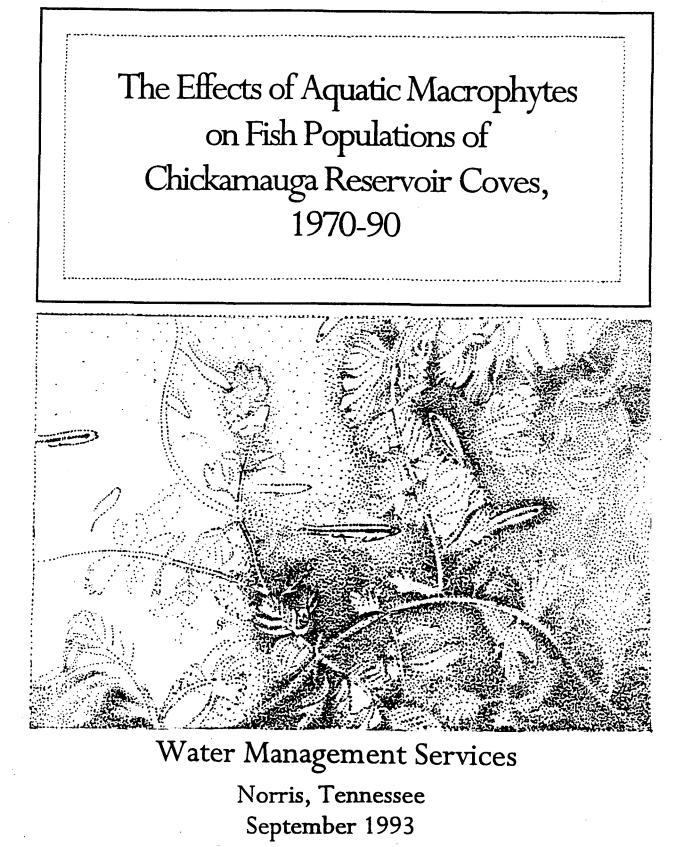
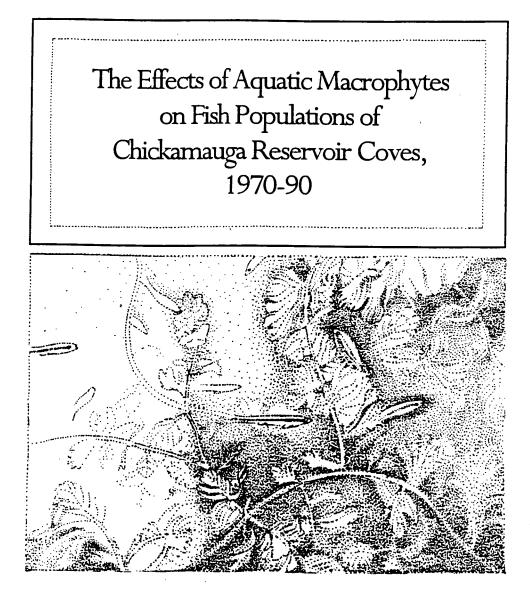
TENNESSEE VALLEY AUTHORITY Water Management



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TENNESSEE VALLEY AUTHORITY Water Management



Prepared by Edwin M. Scott, Jr.

Norris, Tennessee September 1993

TVA/WM--93/24

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ACKNOWLEDGMENTS

Many Tennessee Valley Authority (TVA) employees contributed to the preparation of this report. James Parsly, Hydraulic Engineering Laboratory, Norris, provided seasonal average daily discharges of the Tennessee River from Chickamauga Dam for 1970-83. Tom McDonough, Water Management, Knoxville, provided the computer program for the regression analyses of rotenone data and consultation. Wayne Hamberger, Systems Engineering, Knoxville, provided percentages of possible sunshine measured at the Chattanooga airport, and Ellen Hammond, Chattanooga Engineering Services, retrieved water temperature, secchi depths, and phytoplankton data from STORET. Tom McDonough and Gary Lucas, Systems Engineering. Hydraulic Engineering Laboratory, Norris, provided additional water temperatures. Leon Bates and David Webb, Water Management, Muscle Shoals. provided macrophyte coverages in Chickamauga Reservoir. Earl Burns and Lee Hill, Water Management, Muscle Shoals, supplied recent information about aquatic macrophyte coverage and aquatic plant management on Chickamauga Reservoir. Sportfishing harvest of bluegill and largemouth bass were obtained from Tennessee Wildlife Resources Agency (TWRA) creel surveys, through Todd St. John, Nashville. Neil Woomer, Water Management. Chattanooga, David Webb, and Leon Bates provided extensive review of this manuscript. Julie Madison, Water Management, Norris, supplied the graphics, and with Libby Bailey, also of Norris, prepared the report. Murrie Graser, Water Management, Norris, designed the cover.

EXECUTIVE SUMMARY

The expansion of submersed aquatic macrophytes in Chickamauga Reservoir had a dramatic effect on the resident fish community between 1970 and 1990. Although the original intent of studies upon which this report is based was to monitor the effects of a nuclear power plant on reservoir fish populations, an additional, unforeseen use of data arose when coverages of aquatic macrophytes expanded from less than 2 percent to 21 percent of the surface area of Chickamauga Reservoir during the study period. The primary purpose of this report is to investigate the relationship of submersed aquatic macrophyte coverage to the abundance of selected fish species in cove rotenone surveys. Species whose abundances significantly correlate with aquatic macrophyte coverage are discussed in detail according to the amount and dominant species of macrophyte present during four designated periods. A secondary purpose looks at the possibility of a relationship between the operation of Sequoyah Nuclear Plant (SQN) and the coverage of aquatic macrophytes in Chickamauga Reservoir.

During Period I (1970-75) aquatic macrophyte coverage in Chickamauga Reservoir was less than 100 hectares (ha). Macrophyte coverage increased greatly in Period II (1976-81) to nearly 2200 ha. Coverages of approximately 2800 ha existed during Period III (1982-88) which coincided with several drought years. Coverage of aquatic vegetation was negatively correlated to flows of the Tennessee River, and increased flows and associated factors (such as scouring and turbidity) during Period IV (1989-90) caused vegetation to decrease to approximately 1400

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and 869 ha, respectively. This pattern continued in the 1991 and 1992 seasons when coverage dropped to 275 and 155 ha, respectively.

In the 1960s and 1970s the perennial Eurasian watermilfoil was the dominant species of submersed aquatic vegetation in Chickamauga Reservoir. Then annual species began to increase in abundance, especially spinyleaf naiad which became the most common submersed macrophyte in Chickamauga Reservoir during the 1980s.

Certain fish species became more abundant with increased vegetation coverage. Golden shiner, warmouth, bluegill, redear sunfish, brook silverside, yellow bass, black crappie, and yellow perch are mid-water insectivores which benefited from an increased invertebrate forage base and protective habitat provided by aquatic vegetation. As the forage base for piscivorous species shifted from shad to small sunfish during periods of heavy vegetation coverage, numbers of largemouth bass, an ambush predator, increased.

Other fish species declined in abundance as macrophytes increased. Most of them were open-water, benthic insectivores/omnivores whose feeding habitat in shallow, silted overbanks became colonized with vegetation. Species in this category are carp, smallmouth buffalo, spotted sucker, channel catfish, and freshwater drum. Decline of piscivorous adult white crappie has been attributed to several factors, most of which are directly related to the increase in macrophytes. Declining sauger populations, however, have been attributed to poor spawning success in the headwaters of Chickamauga Reservoir.

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Based on preliminary analysis, milfoil appears more beneficial to several important gamefish species than spinyleaf naiad, possibly because of its different underwater growth form and structure.

The operation of SQN had no measurable effect on the coverage of macrophytes in Chickamauga Reservoir, and thus no indirect effects on resident fish populations.

INTRODUCTION

Macrophyte Effects on Chickamauga Reservoir Fish Populations

Monitoring studies of fish and aquatic macrophyte population trends in Chickamauga Reservoir, with respect to operation of SQN, found an overwhelming relationship of aquatic macrophytes and cove fish populations. This relationship developed over time, as aquatic macrophytes were relatively uncommon in the reservoir in the early years studied, became the purpose of this report. Temporal changes in the fish populations were evident in 1979 (TVA 1980), but were not attributed to aquatic vegetation until 1985 when significant (P>0.05) increases in abundance of certain species, particularly warmouth, redear sunfish, bluegill, and largemouth bass were detected (TVA 1985). At that time smallmouth buffalo, channel catfish, and freshwater drum showed significantly decreasing trends. A seven-fold increase in coverage of rooted aquatic macrophytes between 1976 and 1983 emerged as the overriding influence on the fish community of Chickamauga Reservoir (TVA 1985), and its alteration of the littoral zone habitat has continued to affect rotenone standing stock estimates of several fish species (McDonough and Buchanan 1991, Kerley 1989, 1990, and 1991). This report analyzes 21 consecutive years of fish sampling on Chickamauga Reservoir, spanning a broad range of aquatic macrophyte coverage.

Interpretations of fish community changes are more complex than simple correlations with amount of aquatic vegetation. Fish communities in reservoirs are not balanced (Noble 1986) and are subjected to highly variable water conditions (rate of spring warming, discharges, turbidity, water level fluctuation) which affect planktonic food chains, spawning

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times and success, and early survival of different fish species and interspecific competition between early life stages of fish species. Year class strength of some species depends on high spring discharges following gradual warming of water temperatures. Dissolved oxygen in deeper levels of reservoir such as Chickamauga Reservoir (TVA 1990b) may become limiting for coolwater species during years of low discharges. Aquatic vegetation provides foraging and protective habitat for young of many fish species, according to the size of individual fish.

Influence of Sequoyah Nuclear Plant (SQN) on Aquatic Macrophytes

SQN commenced operation in 1981, the first of eight consecutive years during which Chickamauga Reservoir aquatic macrophyte coverages exceeded 2,000 ha. The existence of SQN as a readily visible factor during these years of macrophyte expansion prompted an examination for any evidence of a connection between SQN operation and macrophyte coverage patterns.

History of Aquatic Macrophytes in Chickamauga Reservoir

Chickamauga Reservoir was created in 1940 with the completion of Chickamauga Dam at Tennessee River mile (TRM) 471. Rooted submersed aquatic macrophytic vegetation was not abundant until the establishment of Eurasian watermilfoil (<u>Myriophyllum spicatum</u>) in 1961 (Smith et al. 1967). Milfoil is an exotic plant that was introduced from an aquarium into Watts Bar Reservoir, the next reservoir upstream, about 1953. From its establishment in Chickamauga Reservoir until the mid-1970s milfoil was the only abundant submersed aquatic macrophyte within the reservoir. While milfoil coverage expanded in most mainstream Tennessee River reservoirs between 1970 and 1985, spinyleaf naiad (<u>Najas minor</u>), another

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exotic species, became the dominant species of vegetation in Chickamauga Reservoir in 1982 (Bates et al. 1985). It continued to be more abundant than milfoil until 1989 when there was a major decline in naiad coverage (Figure 1). Annual coverage of spinyleaf naiad (in combination with southern naiad, <u>N. guadalupensis</u>) and milfoil have fluctuated greatly over time. Hydrilla (<u>Hydrilla verticillata</u>) was found on Chickamauga Reservoir in 1988 (Burns et al. 1989), but has been rarely observed in recent years since treatment with herbicides (Lee Hill, TVA, personal communication).

As milfoil coverage expanded and colonized shallow water habitat along developed shorelines, the Tennessee Valley Authority (TVA) designated priority areas (Figure 2) for herbicide treatment. In addition to the application of herbicides such as 2,4-D to control milfoil and Aquathol K to control naiads, other management techniques currently in use include drawdowns and reservoir surcharge. Milfoil is vulnerable to winter drawdowns because it is a perennial, and its exposed root masses are subjected to lethal winter temperatures. The naiads, which are annuals have seeds which are less affected by dewatering. Winter drawdowns are particularly effective in controlling milfoil within the drawdown zone of Chickamauga Reservoir, relative to other TVA mainstream reservoirs, because the magnitude of the drawdown is greater, over 2.5 m, thereby exposing large areas with milfoil colonies.

Total macrophyte coverage (all species combined) in Chickamauga Reservoir has been dynamic over the past 15 years. Coverage increased from less than 20 ha in 1974 to about 300 ha in 1976. Between 1977 and

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1983, total coverage of aquatic macrophytes increased seven fold (422 to 2,791 ha) and colonized 19 percent of the surface area of Chickamauga Reservoir. During this period, spinyleaf naiad became the dominant submersed species of aquatic vegetation. Submersed aquatic vegetation declined in 1984 (2,161 ha) due to record heavy rains in May, and high flows and increased turbidities, which suppressed growth of deep water colonies (TVA 1985). Aquatic macrophytes in Chickamauga Reservoir reached their greatest abundance during the drought years of 1985-88, averaging over 2,700 ha annually, as clearer water allowed naiads and other macrophytes to grow in deeper water. Peak abundance occurred in 1988, when over 3,000 ha or 21 percent of the total surface area of Chickamauga Reservoir, was colonized. As more normal rainfall patterns returned to the Tennessee Valley in 1989 and 1990, coupled with unusually high flows in June and July 1989, submersed aquatic vegetation dramatically decreased (Burns et al. 1991) to about 1,400 and 900 ha, respectively. High seasonal flows continued in 1991 and 1992, and submersed macrophytes declined to 275 ha in 1991 (Burns et al. 1992) and 155 ha in 1992 (Lee Hill, TVA, personal communication).

Macrophyte Management Plan for Chickamauga Reservoir

The TVA management plan for controlling aquatic macrophytes in Chickamauga Reservoir (Burns et al. 1992) does not attempt the total eradication of aquatic vegetation. Instead the intent is to control vegetation in priority areas: i.e., high-use recreation and public access areas, areas adjacent to residences, resorts, and marinas, small areas of exotic plant colonies in newly colonized reservoirs that could become major problems, and areas of dense vegetation that harbor mosquito

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populations. Approximately 3 percent of the surface area of Chickamauga Reservoir is designated for herbicide treatment if vegetation is excessive. During recent drought years (1985-88), when vegetation coverages exceeded 3,000 ha, about 650 ha-treatments were were made with herbicides annually. Although some of these areas during good growth periods received 2 or 3 treatments during the growing season, the physical areas treated did not exceed 3 percent of the total reservoir surface. Treatment has been reduced along with a natural decline of vegetation in 1989 and 1990, and only 130 ha (cumulative) were treated in 1990 (Burns et al. 1991). The downward trend in vegetation continued in 1991 and 1992, and treatment was reduced to 26 ha in 1992 (David Webb, TVA, personal communication).

Role of Macrophytes in the Aquatic Ecosystem

The literature is rich with information regarding the role of aquatic macrophytes in aquatic ecosystems. To quote John Muir, "When one tugs at a single thing in nature, he finds it attached to everything else in the world." So it seems with aquatic macrophytes. The complex morphology of aquatic macrophytes creates a diversity of microhabitats for colonizing organisms (from epiphytic algae and invertebrates to zooplankton to benthic macroinvertebrates to young and/or small fishes, as well as adult fish of some species) and provides refuge from predators (Miller et al. 1989). Macrophytes also modify the physical environment by directly or indirectly affecting the chemical composition, nutrient cycles, and biological features of the ecosystem. Published literature reporting importance and impacts of macrophytes have been reviewed by Gregg and Rose (1982), McDermid and Naiman (1983), Pandit (1984), Engel (1985,

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1988), and Carpenter and Lodge (1986).

Aquatic macrophytes alter the velocity of waves and currents, modify sedimentation patterns and substrates, stabilize habitats, reduce erosion, affect temperature regimes, and influence available light. Macrophytes store nutrients during spring and early summer, thereby delaying blooms of blue-green algae until the macrophytes senesce in midsummer (Engel 1985). Macrophyte beds, by intercepting runoff, storing nutrients, and retarding algal blooms, can improve water quality (Goulder 1969, Modlin 1970, according to Engel 1985).

Diel variation in photosynthetic and respiration rates within macrophyte beds can cause depletion of oxygen and dissolved inorganic carbon and change pH. Oxygen depletion may be severe in morning hours following night respiration (Losee and Wetzel 1988). During summer when aquatic macrophyte biomass is greatest, diel ranges of dissolved oxygen and pH in a Missouri reservoir were as wide as 14 mg/l and 3 pH units, respectively (Wiley and Jones 1987). Daytime DO concentrations were 230 percent greater near macrophyte beds than no-plant zones (Sculthorpe 1967, according to Miller et al. 1989). Submersed macrophytes are important in maintaining chemical equilibria of the carbon dioxidebicarbonate buffering system of the surrounding water mass.

Sediment-water interactions of aquatic macrophytes greatly influence the hydrochemistry and nutrient cycles in aquatic ecosystems. Nutrients are absorbed from the sediments by rooted species of plants and released back into the water during senescence, providing a nutrient resource for

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phytoplankton (Miller et al. 1989).

Aquatic macrophytes greatly increase habitat available to aquatic invertebrates. Submersed macrophytes provided nearly 10 times more surface area for invertebrate colonization than did the entire benthic area of Lawrence Lake, MI (Losee and Wetzel 1988). Macrophytes support complex interactions of invertebrate behavior, life cycles, and predator-prey relationships. The most common invertebrate organisms in macrophyte beds are crustaceans, chironomids, oligochaetes, and gastropods, with the majority being herbivores such as scrapers, shredders, filterers, and collectors. Less common are predators represented by Coleopterans (beetles), Odonates (dragonflies and damselflies), and Hemipterans (giant water bugs, backswimmers, water scorpions, and water boatmen) (Miller et al. 1989). Core samples indicated more benthic invertebrates below macrophytes than areas devoid of macrophytes.

Significant differences have been documented in the macroinvertebrate communities associated with Eurasian watermilfoil and open littoral habitats of the mainstream Tennessee River (Pardue and Webb 1985). Greater numbers of taxa and individual organisms occurred in dense beds of watermilfoil. Immature insects, amphipods, naidid worms, and leeches were more abundant in the milfoil than in open water areas. A non-burrowing mayfly, <u>Caenis</u>, an important fish food organism, was more abundant in the milfoil, probably due to the increased surface area provided by the highly dissected leaves of watermilfoil as well as the protection from predation afforded by the vegetation. However, a

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burrowing mayfly, <u>Hexagenia bilineata</u>, a highly important fish food organism, preferred the open littoral zone, presumably because the milfoil root system interferes with the ability of the mayfly nymphs to establish burrows.

Larval fishes are typically more abundant in submersed aquatic macrophyte beds than open water areas (Paller 1987), using them as a refuge from predators and feeding on associated zooplankton (Healey 1984). In Orange Lake, Florida, the greatest number of larval fish species was captured in floating-emergent vegetation, which was also important as a nursery area for juveniles of many species (Conrow et al. 1990). In Spirit Lake, Iowa, 17 of 21 YOY species studied were more abundant along vegetated shorelines than adjacent areas devoid of vegetation (Bryan 1989).

Macrophyte beds support higher densities of fish than areas without macrophytes (Borawa et al. 1979, Killgore 1979, Savitz 1981). Largemouth bass and bluegill behavior differs in lakes with and without aquatic macrophytes (Savitz et al. 1983). Small fishes (less than 120 mm long) find shelter in macrophyte beds denser than 200 g dry weight/m², while movement of larger fish is restricted (Hall and Werner 1977, Engel 1985, Mittlebach 1988). Small bass and bluegills avoid no-plant zones along shore, while larger individuals inhabit lower density macrophyte beds offshore (Engel 1985). Juvenile bluegill and longear sunfish numbers were positively correlated with the height of vegetation, and largemouth bass and other game species were more abundant in beds of hydrilla in shallow water than open water areas (Killgore 1979).

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METHODS

Aquatic macrophyte coverage has been quantified annually since 1978 using aerial photography. Overflights were conducted in September or October. Aquatic macrophyte colonies were identified to species or dominant species combinations, delineated during photointerpretation, and area of coverage determined by planimetry (TVA 1985). Macrophyte coverages before 1977 were estimated from herbicide treatment records and by visual inspection (Leon Bates and David Webb, TVA, Water Management, Muscle Shoals, Alabama).

Relationships of vegetation coverage with average daily discharge from Chickamauga Reservoir, mean water temperature, percentage of possible sunshine, and the previous year's vegetation coverage were analyzed by linear regression (SAS Institute, 1985) for the period 1971-92. Daily discharges at Chickamauga Dam, water temperatures (1.5 m depth) at the SQN intake (TRM 484.7), phytoplankton densities and secchi readings near SQN, and sunlight availability at the Chattanooga airport (National Oceanic and Atmospheric Administration, Climatic Data Center, Asheville, North Carolina) were averaged for each year during the first four months of the growing season (March - June), which is the critical period for growth of submersed aquatic vegetation (David Webb and Leon Bates, TVA, personal communication).

Aquatic macrophyte coverage for Chickamauga Reservoir and management units 1, 3, 5, and 8 (Figure 3) was used to evaluate impacts of SQN operation on submersed aquatic vegetation. Coverage data were obtained

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from seasonal workplans of TVA's aquatic plant management program. Trends upstream of SQN were from units 5 and 8, and those downstream from units 1 and 3.

Fish community trend analysis was based on twenty-one consecutive years (1970-90) of cove rotenone surveys conducted in the lower and middle areas of Chickamauga Reservoir (Figure 2), following standardized methods (TVA 1980). The four coves sampled were Gold Point (TRM 476.2R), Chigger Point (TRM 478.0L), Sale Creek (TRM 495.0R/TRM 495.1R), and Gillespie Bend (TRM 508.0R). Log transformed standing stock estimates (number +1/ha) were analyzed by linear regression to determine correlations of submersed aquatic vegetation coverage with total and individual fish species abundances for three size groups. Significant trends, determined at the 0.05 alpha level, were discussed in the text. Trends in total fish biomass were also evaluated by regression analysis. Although the rotenone coves were sprayed with herbicides beginning in 1982 to facilitate the retrieval of fish, vegetation in areas adjacent to the coves was not treated.

Cove rotenone surveys are biased toward fish species inhabiting shallow shoreline areas. Common shoreline species in Chickamauga Reservoir (carp, golden shiner, several sunfish species, largemouth bass, yellow perch, brook silverside, and adult gizzard shad) are adequately sampled by cove rotenone surveys. However, during summer, when rotenone surveys are conducted, species such as black crappie, white crappie, sauger, freshwater drum, channel catfish, yellow bass, smallmouth buffalo, and spotted sucker prefer open-water and/or deeper, cooler water

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than that usually found in coves. Although open-water and deepwater species are not collected in their true abundance by cove rotenone methods, trends in their abundance may reasonably be discerned in TVA's long-standing database.

The twenty-one year time span was divided into four periods according to amount and dominant species of submersed aquatic macrophyte coverage (Figure 4). Standing stocks of fish species significantly correlated with macrophyte abundance were discussed for each period. During Period I, 1970-75, aquatic vegetation was sparse, covering less than 100 ha annually (ranged from 4 to 77 ha), and was primarily milfoil. Macrophyte coverage increased from 293 to 2,188 ha during Period II, 1976-81, and milfoil continued to be dominant. In Period III, 1982-88, vegetation on Chickamauga Reservoir reached its greatest coverage, ranging between 2,161 and 3,175 ha. Spinyleaf naiad, and to a lesser extent southern naiad, were the dominant species of aquatic vegetation during this time period, which included several drought years, especially 1985-88. During Period IV, 1989-90, vegetation decreased to 1,388 and 861 ha, respectively, and was primarily milfoil. The recent trend in macrophyte coverage in Chickamauga Reservoir is similar to trends observed in three other Tennessee River mainstream reservoirs (Figure 5).

Annual reservior-wide estimates of bluegill and largemouth bass harvest, 1977-91, were provided by TWRA creel surveys (TWRA 1992). Because creel surveys span the entire reservoir and year, their results are not directly comparable to shallow cove rotenone surveys conducted in midsummer. Data collected in 1986 were disregarded due to questionable validity (Todd St. John, TWRA, Nashville, personal communication).

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Overall Fish Community

Total fish standing stocks of each size class showed significant positive correlations with aquatic macrophyte coverage in Chickamauga Reservoir (Figure 6A). The trend was most evident for young-of-year (YOY) fish, whose average abundance increased from 8,097/ha during Period I to 26,000/ha and 37,150/ha during Periods II and III, respectively. But when vegetation sharply declined in Period IV, average standing stock of young fish fell to 20,000 fish/ha. Biomass of young fish was also significantly related to aquatic vegetation, ranging from 10.7 kg/ha in 1970 to 112 kg/ha in 1985 (Figure 6B). Average densities of intermediate fish increased from 571 fish/ha in Period I, to 1,212 and 1,206 for the next two time periods. Unlike young fish, intermediate sizes increased when vegetation plummeted in Period IV, averaging nearly 2,000 fish/ha. Biomass of intermediate fish was not significantly correlated with vegetation, and ranged from 12.6 kg/ha in 1980 to 91.2 kg/ha in 1971. Standing stocks of adults averaged 962 individuals/ha in Period I, and grew to averages of 1,407 and 2,035/ha during Periods II and III, respectively. Adult abundance dropped slightly in Period IV. The expansion began with a low of 537 fish/ha in 1970, peaked at 2,818 in 1983 during Period III, after which there has been a gradual, general decline. Biomass of adults was also significantly related to the amount of vegetation coverage, and was greatest during Period III when the average biomass was 269 kg/ha.

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A hydroacoustic survey of Chickamauga Reservoir embayments in August, 1990, indicated fish numbers and biomass were positively related to surface coverage of aquatic macrophytes (Meinert et al. 1992). That study agrees with findings of the present study described above.

Species Increasing with Aquatic Vegetation

Abundance of several fish species increased with macrophyte abundance, most notably golden shiner, warmouth, bluegill, redear sunfish, largemouth bass, black crappie, and brook silverside. Yellow bass and yellow perch, relatively new inhabitants of Chickamauga Reservoir, also increased in abundance along with vegetation. All of these species are midwater insectivores with the exception of largemouth bass, which is an ambush predator on many of the other species. Gizzard shad also appeared to increase with vegetation; however, quantification of their abundance in Chickamauga Reservoir based on cove rotenone data following the initiation of herbicide treatment of rotenone coves in 1982 is somewhat unreliable. Increased abundance of YOY bluegill, redear sunfish, and warmouth shifted the forage base from shad to small sunfish, as also observed at Guntersville Reservoir during heavy macrophyte infestations (TVA 1989).

Golden Shiner

Golden shiner (<u>Notemigonus crysoleucas</u>) abundance (all sizes combined as YOY) was positively correlated with the amount of aquatic macrophyte coverage (Table 1). Average standing stock during Period I was only 3 fish/ha, but increased over a hundred-fold to 369/ha during Period II

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(Figure 7). The peak year of abundance during Period II, 1980, occurred at 794/ha. Stocks of golden shiners declined to an average of 270/ha during Period III. Golden shiners reached their greatest abundance to date in Chickamauga Reservoir during Period IV. The two-year average was 547/ha, and the all-time annual peak abundance was 1,000/ha in 1990. The decline in golden shiner abundance during Period III implies that milfoil provides more favorable habitat for this species than spinyleaf naiad.

Preferred golden shiner habitat is moderate to very dense vegetation. Golden shiners consume a wide variety of food organisms, including protozoans, copepods, cladocerans, aquatic insects, and often filamentous algae and higher aquatic plants (Becker 1983). Being an omnivorous, phytophilic species, golden shiners benefit from aquatic vegetation by deriving habitat, invertebrate food sources, and even macrophytic food sources, explaining their recent upward population trends in Chickamauga Reservoir.

Warmouth

Warmouth (Lepomis gulosus) YOY standing stocks appeared to be highly correlated with aquatic macrophyte abundance (Figure 8). Averaging only 14/ha in Period I, their abundance increased dramatically to 818/ha (maximum 3,235/ha in 1981) in Period II. Average stock density continued high, 1,303/ha, during Period III, but declined with declining vegetation in Period IV, averaging 237/ha.

Standing stocks of intermediate warmouth significantly increased with vegetation, and their average abundances rose steadily during the four

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designated periods. Densities increased from 6/ha to 22/ha to 47/ha and 73/ha during Periods I-IV, respectively. Apparently intermediate warmouth were more able to survive; i.e., escape predation, in less vegetated cover than YOY because of their larger size.

Warmouth are considered food generalists due to the large variation in prey chosen by individual warmouth (Savitz 1981). They are in considerable competition for food with the other fish species present (bluegill, largemouth bass, and yellow perch). Due to their affinity for vegetation, warmouth are considered habitat specialists.

<u>Bluegill</u>

Standing stocks of young and intermediate bluegill (Lepomis macrochirus) significantly increased with aquatic macrophytes in Chickamauga Reservoir, 1970-90 (Table 1). In Period I standing stocks of young and intermediate bluegill averaged 1,484 and 261 fish/ha, respectively (Figure 9). Dramatic increases in abundance occurred during Period II when average standing stocks of young bluegill expanded over 10 times (to 16,620/ha), peaking in 1980 at nearly 27,000/ha, and numbers of intermediate sizes almost tripled (to 717/ha). The second-highest abundance of intermediate bluegill occurred in 1980 at 977/ha. But in Period III bluegill abundance fell. Young bluegill declined to about 70 percent (11,913/ha) of their density during Period II, while intermediate bluegill were less affected, dropping to 575/ha. During Period IV intermediate bluegill rebounded to an average of 973/ha, almost double their density during Period III. Peak abundance of intermediate bluegill, 1,096/ha, occurred in 1990. However, YOY bluegill continued to

-15-

decline in Period IV, dropping to 6,071/ha. As was the case for warmouth, intermediate size bluegill are better able to escape predation in less dense vegetation than the smaller sizes.

Adults followed the same trend as intermediate bluegills, although their abundance did not significantly increase (P>0.05) with total vegetation coverage during the 21-year timespan. During Period III numbers of adults declined in rotenone samples, but rebounded in Period IV following the demise of spinyleaf naiad. Adult bluegill abundance in 1990 was similar to that of Period II, when milfoil was the dominant form of aquatic vegetation.

Annual harvest of bluegill fluctuated between 15,000 and 80,000 (Figure 10A), according to creel surveys conducted 1977-91 (TWRA 1992). Over 50,000 were taken in 1979 (Period II), 1987 and 1988 (Period III), and 1990 and 1991 (Period IV). The drop in 1989 harvest may have resulted from the four year drought which lasted until 1989. Increasing harvest in 1990 agrees with cove rotenone data, and the 1991 harvest is the largest recorded in the 15 year period.

Usual catch rates ranged from 1-2 fish/hr, but were highest in 1979 (Period II) when nearly 5 fish/hr were caught (Figure 10B). Lowest catch rates were in 1984 (Period III). Catch rates improved in 1990 and 1991 following the demise of spinyleaf naiad.

Certain aspects of bluegill life history favor their success, measured in increased abundance, in the presence of aquatic vegetation.

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Hatched from nests in littoral areas, bluegill larvae migrate into open-water habitat before returning to littoral zones at a length of 12 mm (Conrow et al. 1990). Higher larval densities are found in dense, floating-emergent vegetation than littoral zones devoid of vegetation. Young bluegills find both shelter from predation and an abundant food supply in vegetated areas. Food consists of small invertebrates, including chironomids, mayflies, water mites, amphipods, cladocerans, caddisflies, and odonate larvae, supplemented with bryozoans, snails, and vegetation (Savitz 1981, Engel 1985). Bluegills frequently consume filamentous algae, pondweed leaves, and sprouting macrophytes, presumably for clumps of attached invertebrates (Savitz 1981). Lack of invertebrate food due to depletion of Daphnia and the emergence of chironomid larvae force bluegill to consume vegetation in July and August (Engel 1985). As bluegill grow to approximately 180 mm, their diet changes from microcrustaceans to larger insects, and competition with young largemouth bass sometimes occurs (see discussion below). Bluegills less than 100 mm standard length restrict their habitat to vegetated areas for protection from predation at the expense of higher foraging return available in open waters. Larger bluegills switch to open-water zooplankton as their risk of predation diminishes (Mittelbach 1981).

Bluegill in Lake Conroe, Texas, declined following removal of submersed aquatic vegetation by introduced grass carp (<u>Ctenopharyngodon</u> <u>idella</u>) (Bettoli et al. in press). This is the mirror effect of observations made during Period II in Chickamauga Reservoir when bluegill abundance increased dramatically with increases in aquatic macrophytes.

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<u>Redear sunfish</u>

Standing stocks of redear sunfish (Lepomis microlophus) have followed a trend similar to that of bluegill in relation to aquatic macrophyte coverage (Figure 11). YOY redear have shown the strongest relationship (Table 1). Average standing stocks of YOY redear of 84/ha in Period I increased fifty-fold in Period II to 4,435/ha, peaking in 1981 at nearly 22,000/ha. Average densities rose slightly during Period III to 4,623/ha, reaching almost 8,000/ha in 1988 which was roughly a third of the peak abundance of Period II. When macrophytes declined in Period IV, standing stocks of YOY redear fell to an average of only 130/ha.

Intermediate redear sunfish abundance also corresponded positively to increased macrophyte coverage. There was a two-fold increase in average standing stocks between Period I and Period II, 21/ha vs. 41/ha. During Period III, standing stocks averaged 128/ha, nearly triple the previous average. Still greater abundances occurred in spite of vegetation declines in Period IV, peaking at over 700/ha in 1989, and averaged 413/ha. Increasing abundances of intermediate redear sunfish during Periods III and IV, while YOY abundances were declining, indicated the advantage of a larger size in being more able to escape predation in sparse vegetation than YOY.

Although overall correlation of adult redear sunfish and aquatic vegetation was not significant at the 0.05 level, useful information in the average standing stocks during the four designated periods was found. Adults were more abundant during the periods when milfoil was the predominant form of aquatic vegetation (Periods II and IV). Standing

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stocks averaged 74/ha and 101/ha, respectively for these periods, compared to averages of 42/ha during Period I and 54/ha during Period III. This is consistent with the pattern observed in adult bluegill standing stocks.

Largemouth Bass

Young and intermediate largemouth bass (<u>Micropterus salmoides</u>) standing stocks were significantly correlated with acreages of aquatic macrophytes (Table 1). Densities of young ranged from 18/ha in 1972 to 912 in 1980. Average densities in Period I, 66/ha, increased nearly nine-fold to 526/ha during Period II (Figure 12). In Period III average densities of young largemouth bass fell to 206/ha. Average abundance increased somewhat in Period IV to 285/ha.

Standing stocks of intermediate largemouth bass ranged between 3/ha in 1970 to 145/ha in 1981. Average densities were lowest, 28/ha, in Period I, and increased to 87 and 85/ha during Periods II and III, respectively. During Period IV average standing stocks of intermediate largemouth bass fell slightly to 58/ha.

Although the relationship between standing stocks of adult largemouth bass and aquatic vegetation was not statistically significant at the 0.05 level, their numbers generally increased over the 21-year period (Figure 12). The lowest abundance was 3/ha in 1970 (Period I), and the highest was 48/ha in 1985 (Period III). Cove populations of adult largemouth bass varied little since 1987 in spite of declining vegetation after 1988.

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An increasing trend in largemouth bass abundance (all sizes combined) relative to aquatic macrophyte coverage is evident in Chickamauga Reservoir coves (Figure 12). The positive relationship between largemouth bass and aquatic vegetation was also evident in Lake Conroe, Texas, when numbers of age-1 and older bass declined after grass carp removed aquatic macrophytes (Bettoli et al. in press).

Fishing effort for largemouth bass in Chickamauga Reservoir increased tremendously between 1977 and 1991 (TWRA 1992). Creel surveys conducted by the State of Tennessee indicate black bass, predominantly largemouth bass, have been the most sought sportfish species in the reservoir since 1982. Estimated angler hours of fishermen seeking black bass increased from 35,000 in 1977 to nearly 600,000 in 1991 (Table 3). Expressed in trips, these figures represent about 8,250 in 1977 compared to nearly 190,000 in 1991. Harvest of largemouth bass increased nearly thirteen-fold between 1977 and 1991, as annual estimates increased from about 5,000 fish in 1977 to nearly 64,000 in 1991, peaking at over 125,000 fish in 1988 (Table 3, Figure 13A). Harvest estimates of largemouth bass numbers and weights were directly (r=0.64, P=0.0100 and r=0.89, P=0.0001, respectively) correlated to fishing pressure.

Annual catch rates for harvested fish over the past 16 years averaged about 0.20 fish/hr, ranging from 0.09 to 0.37 fish/hr (Figure 13B). Highest harvest rates occurred in 1983, 1984, and 1988, while lowest rates occurred in 1990 and 1991. Actual catch rates were higher than harvest rates, since many bass fishermen practiced catch and release fishing. Data from 1988 to 1991 indicate roughly 3 times more largemouth

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bass were caught than harvested in Chickamauga Reservoir (TWRA 1992). Declining harvest rates in recent years may be partly explained by catch and release fishing.

Harvest of largemouth bass by number and weight and fishing pressure were not significantly correlated (P>0.05) with the amount of aquatic macrophyte coverage during the study period. However, there was a positive relationship between catch rates for harvested fish and aquatic macrophytes (r=0.58, P=0.0248). Largemouth bass were harvested at higher rates during years of heavy aquatic vegetation, perhaps because they were more vulnerable to capture around patches of vegetation. Fishing pressure (r=0.86, P=0.0001) and harvest of largemouth bass numbers (r=0.61, P=0.0158) and weights (r=0.83, P=0.0001) were significantly correlated with year, indicating that all three parameters increased over time. However, harvest rates and average size of largemouth bass did not show a significant trend relative to year.

Comparison of data from 1988 and 1991, years of highly contrasting vegetation coverage, illustrates the effect of aquatic macrophytes on a largemouth bass sport fishery. During 1988, when vegetation was heavy, the number of bass harvested was greatest of the study period, but mean size was smallest, 0.56 pounds (Table 3). Total weight harvested was about 70,000 pounds. Bass were larger when vegetation coverage was sparse in 1991, averaging 1.32 pounds each. Total weight harvested in 1991 was highest of the study period, nearly 85,000 pounds, although the total number harvested was only half that of 1988 (TWRA 1992). Other studies have shown that usually there is an inverse relationship between

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mean size of largemouth bass and aquatic macrophyte coverage (Mike Maceina, Auburn University, personal communication).

From 1987 to 1991 over 50,000 pounds of largemouth bass were taken annually from Chickamauga Reservoir (TWRA 1992). The question arises as to the amount of fishing pressure and harvest Chickamauga Reservoir can sustain and still provide a quality sport fishery. Since 1987 the annual harvest rate averaged 1.71 pounds/acre. The greatest harvest rate, 2.39 pounds/acre, occurred in 1991, and was followed in 1992 by the lowest rate in recent years, 1.19 (Table 3). However, all harvest rates, are less than the national average of 2.67 pounds/acre (Jenkins 1982). At Guntersville Reservoir, another Tennessee River mainstem reservoir, 46 miles downstream, harvest rates were higher, but have decreased from 4.70 pounds/acre in 1990 to 3.66 and 1.59 in 1991 and 1992, respectively (Donny Lowery, TVA, personal communication). Fishing pressure and harvest at Chickamauga Reservoir declined in 1992 to 282,000 hours and 30,846 fish, respectively, but harvest rate, catch rate, and mean size improved from 1991 estimates (TWRA 1993).

Small bass and bluegill (39 to 119 mm) seek shelter in aquatic vegetation, but their access into extremely dense vegetation (greater than 300 g dry wt/m²) is restricted. Medium-sized largemouth bass and bluegill (120 to 179 mm) select loose vegetation or disperse offshore. Bass larger than 180 mm stay offshore or in channels or open areas of the foliage, although during summer they may seek cooler water temperatures provided by overhanging plant canopies (Engel 1988). They frequently occupy small "holes" devoid of plants within dense vegetation, and become

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ambush predators, whereas in sparse vegetation they actively pursue prey (Killgore et al. 1989).

Principal food items of small largemouth bass inhabiting vegetation are chironomids, odonates, mayflies, fishes, and cladocerans (Engel 1985). The onset of piscivory by young largemouth in Lake Conroe, Texas, was delayed by abundant aquatic vegetation, as largemouth bass smaller than 100 mm rarely consumed fish (Bettoli et al. 1992). Most did not become piscivorous until reaching lengths of 140 mm. But when vegetation was very sparse, piscivory of young largemouth bass was common at smaller lengths, down to about 60 mm. Earlier piscivory translates into faster growth rates of young largemouth. In Wisconsin fish in the diet outnumber insects when bass lengths exceed 240 mm. (Engel 1985). In Oklahoma crayfish are the dominant food of bass in dense vegetation, followed by bluegill and YOY bass. In heavier vegetation YOY bass are more vulnerable to predation than bluegill, until they grow large enough to escape predation, about October (Summers 1980).

Largemouth bass production is enhanced by the presence of aquatic macrophytes; however, there is a point where increasing aquatic vegetation leads to decreased bass production, presumably due to foraging inefficiencies brought about by excessive plant growth. That point appears to be above 20 percent coverage in studies of 30 Texas reservoirs (Durocher et al. 1984). Optimal largemouth bass production in Illinois ponds dominated by <u>Potamogeton crispus</u> and <u>Najas flexilis</u> occurs at an aquatic macrophyte standing crop of 52 grams dry weight/m³ (Wiley et al. 1984). Bass growth is also reduced in waters having dense vegetation (Engel 1985).

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In experiments of various densities of simulated vegetation, largemouth bass switch foraging behavior from active searching to ambush tactics as plant density increases. Fathead minnows and golden shiners are more vulnerable to predation than bluegills (Savino and Stein 1982a). As simulated vegetation becomes extremely dense, bass predatory activity declines in response to reduced visual contact with bluegills and the ability of the bluegills to "hide" (Savino and Stein 1982b). Other laboratory experiments indicate that in sparsely vegetated environs largemouth bass are food specialists, whereas in dense vegetation they are food generalists (Anderson 1984).

Largemouth bass and bluegill are commonly found together in aquatic macrophyte beds, and the initial relationship observed is that of predator-prey. However, this is a function of size; i.e., the larger bass is the predator and the bluegill is the prey. Given the differences in size, both species have been designated as food specialists since there is very little or no competition for food (Savitz 1981). But when size is taken into account, and both species are smaller than 120 mm, diet overlap can be as high as 70 percent (Engel 1988). Diet overlap decreases to only 11 percent as the two species grow beyond 180 mm with bass consuming insects and small fish while bluegills graze zooplankton. Young of year bass and YOY yellow perch (25-49 mm) have a slightly overlapping diet for invertebrates in West Point Lake, Alabama-Georgia until midsummer at which time bass switch to small fishes (Timmons 1984).

<u>Black crappie</u>

Although black crappie (Pomoxis nigromaculatus) were present in cove

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rotenone samples as far back as 1972, they have only recently become common in Chickamauga Reservoir (Figure 14). Recent increases in YOY black crappie standing stocks were significantly correlated with increases in aquatic macrophyte coverages (Table 1). Increased water clarity during the drought years of 1985-88 may have favored black crappie over white crappie, which tend to do better in turbid water. Since increased water clarity also encourages aquatic macrophyte growth, it is not possible to say which had the larger effect on black crappie abundance. YOY black crappie were very rare in cove rotenone samples in Periods I and II, occurring in only three years and averaging only 1 fish/ha. During Period III average standing stocks improved greatly, to 16/ha, and the peak abundance occurred in 1988 when 102/ha were found. Following the decrease of naiads in Period IV, YOY black crappie abundance remained high, as standing stocks were 18 and 72/ha, respectively.

Increases in intermediate black crappie abundance were also correlated with aquatic macrophytes, but their numbers have not expanded greatly. None were found in rotenone surveys in Period I, and the average during Period II was only 1 fish/ha. Three fish/ha was the average density during Periods III and IV. Peak abundance of intermediate black crappie occurred in 1988 when standing stocks were 12/ha.

The shift in dominance to black crappie from white crappie in Chickamauga Reservoir has been recently attributed to increased aquatic macrophyte coverage (McDonough and Buchanan 1991). Young black crappie

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are thought to benefit more from vegetated areas for invertebrate feeding and shelter than white crappie which prefer deeper, more open-water habitats. This is particularly important in Chickamauga Reservoir where juvenile crappie survival is critical to harvestable-size recruitment. At lengths less than 200 mm both crappie species are zooplanktivorous and insectivorous (Ellison 1984). Adults of both species are mainly piscivorous, but black crappie are somewhat slower to convert to a fish diet, and are less adapted to capturing prey in turbid water than white crappie.

In a natural Wisconsin lake, crappie (presumably black crappie) occupied the pelagic region adjacent to macrophyte beds during summer and were spatially segregated from largemouth bass and bluegill (Engel 1985). They ate mainly zooplankton (<u>Daphnia</u>, <u>Mesocyclops</u>, and <u>Leptodora</u>) and relatively few insects, making their diet more specialized than largemouth bass or bluegill, and remained zooplanktivorous until about 200 mm. Small crappie consumed food under 2 mm long, while large crappie food measured 5 mm or more in length. The diets of medium bluegills and crappies overlapped by 90 percent, as both species shared <u>Daphnia</u> and <u>Leptodora</u>. This interspecific competition intensified with the collapse of <u>Daphnia</u> in July. Diets diverged as they grew to larger sizes, with overlaps of only 25 percent (Engel 1985). Fish are of primary importance to adult black crappie on a volume basis, although invertebrates are numerically more important (Becker 1983).

Brook silverside

Brook silverside (Labidesthes sicculus) also have benefited by

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increases in aquatic macrophytes (Table 1). Their average standing stocks (all sizes considered YOY) increased from 18/ha during Period I to 100/ha in Period II (Figure 15). Their greatest average abundance, 212/ha, occurred in Period III, peaking in 1986 at 490/ha. Average standing stocks of brook silverside fell to 92/ha in Period IV. Apparently this surface-dwelling species is more suited to abundant naiad growth than abundant milfoil growth.

Affinity of brook silverside for aquatic macrophytes was also reported from Lake Conroe, Texas (Bettoli et al. 1991, Bettoli et al. in press). Stocks of brook silverside collapsed following the removal of aquatic macrophytes by grass carp. However another atherinid species, the inland silverside (<u>Menidia beryllina</u>), increased in abundance, presumably because it was better adapted to open water conditions than brook silverside.

Young brook silverside are pelagic most of their first summer, feeding on cladocerans, copepods, and other small crustaceans. They migrate to shallow shoreline areas as summer ends, joining adult brook silverside. Diet shifts to aquatic and terrestrial insects along with move to shoreline habitat (Pflieger 1975). Their entire lifespan is 17 months (Becker 1983).

<u>Yellow Bass</u>

Standing stocks of all three sizes of yellow bass (<u>Morone</u> <u>mississippiensis</u>), a species not found in Chickamauga Reservoir before 1969 (Starnes et al. 1982), have shown significant positive correlations

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with aquatic macrophyte coverage in Chickamauga Reservoir (Table 1). Average YOY abundances have steadily increased over the four periods, although there were marked declines in 1978 and 1979 (Figure 16). Standing stocks increased from an average of 10/ha in Period I to 16/ha in Period II. During Period III, YOY yellow bass abundance averaged 28/ha, but the greatest abundance occurred in Period IV when average stocks were 190/ha. The all-time peak abundance of YOY occurred in 1990 (372/ha).

Intermediate and adult yellow bass also rose in average abundance in the first three periods, but, in contrast to YOY, fell in Period IV. Standing stocks of intermediate yellow bass were 3, 13, 13, and 5/ha, respectively, while adult stocks averaged 1, 4, 5, and 3/ha, respectively, over the four periods.

Young yellow bass feed primarily on small crustaceans (copepods and cladocerans), switching to aquatic insects as they grow (Pfleiger 1975). Based on the positive correlation of YOY yellow bass and aquatic macrophytes in Chickamauga Reservoir, and the enlarged insect fauna associated with aquatic macrophytes, it appears the macrophyte beds provide a desirable invertebrate food supply for YOY yellow bass.

Larger yellow bass also benefited from increased macrophyte coverage, presumably due to invertebrate fauna inhabiting submersed vegetation (Starnes et al. 1982), although fish become more important in their diet as they grow. Adults are mostly piscivorous, feeding on small fish, even young of their own species (Pfleiger 1975).

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Yellow Perch

The young of yellow perch (Perca flavescens), a non-native species that invaded the Tennessee River system following introductions into headwater reservoirs of the Hiwassee River in North Carolina in the 1950's (Etnier and Starnes, in press), have increased in abundance along with aquatic vegetation in Chickamauga Reservoir (Table 1). Average standing stocks were 1/ha in Period I, 21/ha during Period II, 12/ha during Period III, and finally 32/ha in Period IV. Relatively high abundances were found in three years of Period II (1978 [36/ha], 1980 [41/ha], and 1981 [39/ha]) and one year of Period IV (1990 [48/ha]), when milfoil was the dominant macrophyte (Figure 17). YOY yellow perch abundance was also high in one year of Period III, 1988 (48/ha), but was less than 10/ha during the other six years of Period III.

Although trends in intermediate and adult abundance were not significantly correlated with coverage of total aquatic macrophytes, annual standing stock estimates imply yellow perch abundance is related to the type of aquatic vegetation Chickamauga Reservoir. Stocks rose from averages of 8 and 4/ha for intermediate and adult sizes, respectively, during Period I to 54 and 20/ha during Period II. But in Period III, average intermediate and adult stocks fell to 28 and 14/ha, respectively. When naiads declined and milfoil was once again the dominant macrophyte in Period IV, average standing stocks of intermediate and adult yellow perch rebounded to all-time highs of 118 and 83/ha, respectively. Thus, standing stocks of yellow perch appear to be more related to the species of aquatic macrophyte present than the amount of aquatic macrophyte present; i.e., they are more numerous during years of

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greater milfoil coverage than years when naiads are more abundant.

Yellow perch are termed food generalists due to large trophic diversity (Savitz 1981). They feed mostly on small invertebrates, along with some minnows and darters, putting them in competition with other fish species inhabiting littoral vegetation. They prefer habitats with cooler water than bluegill, warmouth, and largemouth bass (Timmons 1984), thus segregating them somewhat spatially.

Gizzard Shad

Standing stocks of YOY gizzard shad (<u>Dorosoma cepedianum</u>) have significantly increased with aquatic macrophytes in Chickamauga Reservoir (Table 1), but have been highly erratic over time, according to cove rotenone surveys (Figure 19). The average density in Period I was only 38 fish/ha, although they were considerably more abundant during 1970 and 1971. In Period II the average YOY gizzard shad density increased to 338/ha due to high densities of 1,905 and 115/ha during 1977 and 1978, respectively. Highest average standing stocks, 850/ha, occurred during Period III, and ranged from about 60/ha in 1984 to 2,800 in 1985. YOY gizzard shad dropped to about 8/ha but rebounded to 741/ha during Period IV.

There are several possible explanations for the variable abundance of gizzard shad in rotenone samples. YOY gizzard shad form pelagic, actively foraging schools in open-water areas during daylight hours, creating patchy distributions. When the block net is placed across the mouth of the rotenone cove late in the afternoon, schools of YOY shad may

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or may not be trapped in the cove. Increasing milfoil colonies between 1978 and 1981 reduced feeding zones inside the rotenone coves, resulting in apparent declining stocks of YOY shad. Beginning in 1982, coves were treated with herbicides to remove vegetation and facilitate collection of fish during rotenone surveys. However, this activity created open areas in the vegetation and may have attracted YOY shad into the "new" feeding zones. Standing stocks jumped in 1982 and remained high during the years of naiad dominance, but this is not believed to be a result of the increased vegetation, rather an attraction to the open areas following herbicide treatment.

Another factor complicating conclusions about YOY gizzard shad abundance drawn from rotenone surveys is species interactions with YOY threadfin shad. Evidence of competition between the two shad species followed the severe winters of 1976-77 and 1977-78, which caused high mortalities of temperature-sensitive threadfin shad. Coincident with greatly reduced stocks of YOY threadfin shad in cove rotenone samples, the following summers were very high stocks of YOY gizzard shad (Figure 19). Both species have extended, overlapping spawning seasons of approximately three months in the spring and early summer (Wallus and Kay 1990), and resultant young shad may be in frequent competition for planktonic food organisms during their first summer. (Inability to identify larval shad prevents research on interspecific competition in areas where they coexist.)

An obvious shortcoming of cove rotenone surveys to measure YOY gizzard shad abundance is the fact that standing stocks of adults are

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usually one or two orders of magnitude greater than the abundance of young (Figure 18). This amplifies the pelagic nature of YOY shad; i.e., the young are not sampled by cove rotenone in their "true" reservoir abundance. Hydroacoustic surveys provide better measurement of fish in the pelagic zone, which are primarily YOY shad during the summer.

Regression analysis indicated that adult gizzard shad standing stocks are significantly correlated to aquatic macrophyte coverage (Table 1). However, for the same reasons as YOY gizzard shad, adult shad should not be expected to increase in abundance with vegetation because their preferred habitat is open water (Wallus and Kay 1990). Declining abundance in 1979 and 1980 samples, before coves were treated with herbicides, may just indicate foraging habitat of adult shad was reduced by increased macrophyte coverage (Figure 18). There was little difference in average standing stocks between Periods I and II, 622 vs. 773 fish/ha, respectively. However, a major increase occurred during Period III, when average abundance doubled to 1,547 fish/ha. But since coves were treated with herbicides during this period, their observed abundance is not considered to be directly associated with the amount of aquatic vegetation, but instead due to the attraction of open-water foraging habitat within the macrophyte beds created by herbicide treatments. As the abundance of macrophytes plummeted in Period IV, gizzard shad abundance "fell" to an average of 933 fish/ha, which may be a more accurate measurement of their true abundance.

Aspects of gizzard shad early life history have a direct bearing on year classes of other species. Due to their sheer numbers and extended

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period of occurrence, larval and juvenile gizzard shad have an enormous effect on the planktonic food supply necessary to support young fish of spatially and temporally coincident species. Gizzard shad measuring between 6-25 mm in length feed exclusively on zooplankton (Kissick 1988) and later switch to phytoplankton. Selective predation on zooplankton populations by young gizzard shad is substantial (Cramer and Marzolf 1970), and can lead to zooplankton depletion (Kissick 1988). In one study gizzard shad less than 20 mm in length ate zooplankton (Cyclops, Bosmina, and Daphnia), but gradually switched to phytoplankton when they grew beyond 22 mm (Cramer and Marzolf 1970). Another study showed shad zooplanktivory continued to 30 mm, with small shad larvae less than 25 mm selecting larger zooplankton (Cyclops and Calanoida) and larger larvae feeding on smaller zooplankton (Bosmina and Keratella), while a 35 mm specimen had fed exclusively on the phytoplankter Pediastrum (Barger and Kilambi 1980). Young gizzard shad virtually cease feeding at night (Kissick 1988). In pond experiments gizzard shad suppressed bluegill growth and recruitment by removing planktonic food needed by small bluegill which indirectly affected growth of largemouth bass that preyed upon bluegill (Kirk and Davies 1985). In Ohio reservoirs peak densities of larval gizzard shad can drive zooplankton to temporal extinction, thereby limiting recruitment of other fish species and controlling fish community composition (DeVries and Stein 1992). Competition for planktonic food during larval stages to the detriment of other species has also been demonstrated with threadfin shad (Ziebell, et al. 1986, DeVries et al. 1991).

The timing of shad spawning, determined by water temperature, is a

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major factor in largemouth bass year class strength. Gradually rising temperatures in the spring delay shad spawning, separating them more temporally from the earlier-spawned largemouth bass. This favors YOY bass survival by allowing them to grow larger before the young shad become available as prey (Adams and DeAngeles 1987). If water temperature rises rapidly, young shad will grow too quickly to be utilized by YOY bass.

Species Decreasing with Aquatic Vegetation

Most of the fish species that have decreased over time are open-water benthic insectivores or omnivores, such as carp, smallmouth buffalo, spotted sucker, channel catfish, and freshwater drum. Their feeding habitat is predominantly broad mud flats. Piscivorous species that have declined, such as sauger and white crappie are also associated with open-water habitat.

Carp

Standing stocks of adult carp (<u>Cyprinus carpio</u>) have decreased relative to aquatic macrophyte abundance (Table 1). They were most numerous in Period I when average stocks were 10/ha (Figure 20). Peak abundance was measured in 1971 at 17/ha. As milfoil expanded during Period II average adult carp abundance dropped to 4/ha. During Period III average stocks were 2/ha, and remained at that level through Period IV.

A curious contrast to declining adult carp abundance relative to aquatic vegetation is increasing YOY abundance (Table 1). YOY carp were

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virtually absent in cove rotenone samples during Period I, averaging 0.06/ha. As milfoil invaded, average densities jumped to 5/ha, peaking at 24/ha in 1981. Average density fell to 3/ha in Period III. No YOY carp were found in Period IV.

Carp is a highly adaptive species, and tolerates considerable pollution. They are omnivorous, and consume a wide variety of animal and plant material (Pflieger 1975). Aquatic insects are the most important food category. Their decline in abundance in Chickamauga Reservoir cannot be explained solely in terms of vegetation dynamics.

Smallmouth Buffalo

Intermediate and adult smallmouth buffalo (<u>Ictiobus bubalus</u>) standing stocks were inversely related to aquatic macrophyte abundance (Table 1). Intermediate smallmouth buffalo abundance averaged highest (7/ha) during Period I (Figure 21). Peak abundance was 36/ha in 1971. This size has been virtually absent from cove rotenone samples since 1975, appearing during only three years.

Adult smallmouth buffalo also were most abundant during Period I, averaging 10/ha during 1970-75. Peak abundance of 25/ha occurred in 1972. As milfoil proliferated in Period II, adult smallmouth buffalo stocks fell to average 2/ha, and remained at that level during Period III. Stocks fell to 1/ha during Period III, following the collapse of naiads.

Smallmouth buffalo are opportunistic bottom-feeders, consuming

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aquatic insects, attached algae, crustaceans, and zooplankton (Tomelleri and Eberle 1990). Its feeding behavior is adapted for open, soft substrates, and is apparently hindered by rooted aquatic vegetation.

Spotted sucker

Another benthic invertivore significantly decreasing in abundance relative to aquatic macrophytes is spotted sucker (<u>Minytrema melanops</u>), specifically YOY and adults. Average YOY standing stocks were 19/ha in Period I (Figure 22). Peak YOY occurrence was in 1973 when 93/ha were observed. Average density fell to 4/ha during Period II, and further declined in Period III to 2/ha. Their abundance did not change in Period IV.

Standing stocks of adults averaged 13/ha in Period I, and increased slightly to 15/ha during Period II. Peak abundance was 31/ha in 1974. However, as total vegetation reached its greatest coverage in the 1980's, average abundance of adult spotted suckers dropped to 4/ha, and remained low (3/ha) in Period IV.

Food habits of spotted sucker have been little studied, but probable food organisms are molluscs and insect larvae (Forbes and Richardson 1920). A 150-mm specimen had ingested mostly ostracods and chironomid larvae, along with lesser amounts of mayfly larvae, amphipods, copepods, filamentous algae, and higher plant material (Becker 1983).

Channel Catfish

YOY and intermediate channel catfish (Ictalurus punctatus) abundance

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in Chickamauga Reservoir has decreased in relation to aquatic macrophytes (Table 1). Although YOY have never been particularly abundant in cove rotenone surveys in Chickamauga (all-time peak annual abundance was 9/ha), average standing stocks fell from 3/ha to 2/ha between Period I and Period II (Figure 23). YOY abundance further declined during Period III to an average of 0.4/ha, as none were sampled in five of the seven years of that period. Their numbers remained below 1/ha in Period IV.

Stocks of intermediate channel catfish were highest in Period I, averaging 8/ha. Their greatest abundance, 12/ha, occurred in 1971 and 1972. When milfoil coverage increased in Period II, intermediate stocks decreased to an average of only 2/ha. During Period III this size of channel catfish continued to decline in abundance, falling to an average of 1/ha. No rebound in abundance occurred during Period IV, and the average density of intermediate channel catfish remained at 1/ha.

Standing stocks of adult channel catfish were not significantly correlated at the 0.05 alpha level to aquatic macrophyte abundance, but were highly erratic over time. The peak abundance, 50/ha, occurred in 1981 when milfoil was very abundant; and were it not for this unusual observation, a significant negative relationship would have been found. Average standing stocks were 14/ha and 17/ha during the first two periods, respectively, and dropped to 6/ha in Period III. In Period IV adult channel catfish abundance still averaged only 5/ha.

Channel catfish feed on the bottom and detect food by touch and smell. Food is highly varied, both living and non-living (Becker 1983).

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Insects, freshwater clams, snails, fish, and algae are common food items, with fish becoming more important numerically for adult catfish (Tomelleri and Eberle 1990).

White Crappie

The recent decline of white crappie (Pomoxis annularis) in Chickamauga Reservoir has been of great concern (Buchanan and McDonough 1990, McDonough and Buchanan 1991). Declining adult stocks have been attributed at least in part to increased aquatic vegetation. Other factors were obviously involved because average density during Period I. 8/ha, increased to 11/ha in Period II (Figure 24). The all-time annual peak density, 30/ha, occurred in 1979 following three years of rapidly climbing milfoil coverage (from 293 to 802 ha). Major declines in adult stocks occurred during Period III, as average abundance fell to 1/ha. In 1987 and 1988 no adult white crappie at all were found in any of the four Chickamauga coves sampled. White crappie did not recover following the naiad decline in Period IV, as average density was less than 1/ha. The peak abundance of YOY in 1987, 135/ha, did not survive to intermediate and/or adult size in succeeding years.

Although spawning success and survival through the first summer were found in Chickamauga Reservoir, year class strength declined during the second summer when mortality was correlated with increased aquatic vegetation, YOY and yearling sunfish and largemouth bass, and adult largemouth bass and gizzard shad (Buchanan and McDonough 1990). Larval sunfish abundance greatly increased during the years of heaviest macrophyte colonization, and the resultant competition for planktonic

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food may have reduced larval white crappie abundance in some years. Pelagic crappie also compete with preponderant numbers of larval gizzard and threadfin shad, which are capable of causing zooplankton depletion. Circumstantial evidence from Douglas Reservoir, Tennessee, indicates young, pelagic white crappie do very well in the absence of threadfin shad (Wilson 1991).

In recent years, white crappie have been replaced in dominance by black crappie in Chickamauga Reservoir due to differences in food habits and preferred habitats, especially during the drought period of 1985-88. YOY white crappie abundance was positively correlated with abundance of YOY gizzard shad, and inversely correlated with the abundance of adult threadfin shad. Adult white crappie abundance was positively correlated with dissolved oxygen, and negatively correlated with aquatic vegetation; fishing pressure; density of YOY black crappie, sunfish, gizzard shad, and threadfin shad; yearling stocks of sunfish; and adult density of gizzard shad (Buchanan and McDonough 1990).

Zooplankton food of young white crappie (11-100 mm) included <u>Daphnia</u>, <u>Cyclops</u>, <u>Diaptomus</u>, and <u>Leptodora</u>, with <u>Cyclops</u> being most strongly selected in crappie less than 30 mm. Bottom fauna organisms were unimportant food items. As the fish grew, the order of selection was <u>Daphnia</u>, <u>Diaptomus</u>, and <u>Leptodora</u> (Nelson et al. 1967). Small white crappie (8-17 cm) are pelagic in reservoirs (O'Brien et al. 1984). In Chickamauga Reservoir, juveniles of both crappie species (50-200 mm) had very similar feeding habits, consuming primarily copepods, with lesser amounts of cladocerans and chironomid larvae (Buchanan and McDonough

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1990). As previously mentioned, competition with bluegills and other sunfish appears to have seriously impaired year class strengths of white crappie in Chickamauga Reservoir during the 1980's, coincident with heavy colonization of aquatic macrophytes.

In pond experiments the presence of gizzard and threadfin shad decreased the total number and biomass of YOY white crappie (Guest et al. 1990), due to overlapping zooplankton diets as larvae and juveniles. Threadfin shad continue to feed on zooplankton as adults by visual feeding, and suppressed densities of copepods and <u>Daphnia</u>. Adult threadfin shad compete with white crappie for zooplanktonic food, while adult gizzard shad do not. This is consistent with previous studies on Chickamauga Reservoir (Buchanan and McDonough 1990).

<u>Sauger</u>

All three sizes of sauger (<u>Stizostedion canadense</u>) declined coincident with increasing vegetation (Table 1). However the major factor for their decline is believed to be poor spawning success at Hunter Shoals (TRM 521) due to low discharges from Watts Bar Dam during April, especially during the drought years of the mid to late 1980s (Hickman et al. 1990).

YOY sauger abundance averaged 1.2/ha in Period I before aquatic macrophytes were common (Figure 25). Average abundance fell 0.6/ha during Period II, and sauger were only found one year of that 6-year period, 1977, which was also the all-time peak occurrence year, when 4/ha were measured. No YOY sauger were collected in cove rotenone surveys

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during Period III, but small numbers reappeared in 1990. These could have been naturally reproduced in Chickamauga Reservoir, but it is more likely they were from 191,000 fingerlings released by the Tennessee Wildlife Resources Agency (TWRA) into Chickamauga Reservoir during 1990 (Hevel and Hickman 1991).

Intermediate sauger declined from 1.0/ha in Period I 0.8/ha in Period II. No intermediate sauger were collected in cove rotenone surveys after 1979. Adult sauger average stocks fell successively during the four designated periods from 1.2/ha to 1.0, 0.4, and finally 0.0/ha, respectively.

Food of young saugers in the 12-50 mm size range includes <u>Daphnia</u>, <u>Cyclops</u>, chironomids, and YOY of other fishes, such as white bass and freshwater drum (Priegel 1969). Fish become more important in sauger diet as the young grow, but microcrustaceans are eaten when smaller fish are unavailable. Juvenile and adult sauger are highly piscivorous in Tennessee River reservoirs, preying heavily upon gizzard and threadfin shad, although <u>Hexagenia</u> nymphs occasionally occur in the diet.

Due to preference for coolwater habitats, sauger are not commonly found in the relatively shallow coves chosen for rotenone samples. Furthermore they are not directly affected by aquatic macrophytes due to their cooler, deeper habitat. But even though rotenone surveys do not provide accurate estimates of sauger abundance, the virtual absence of all three sizes from 1980 to 1990 indicates the species suffered considerable decline in Chickamauga Reservoir during that time period.

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Estimates of the sauger spawning population fell from 18,000+ in 1986 to approximately 2,000 in 1991 (Hevel and Hickman 1991)

Sauger larvae do not compete for zooplankton with larval shad and sunfish due to temporal separation; i.e., sauger larvae hatch earlier in the spring than most other species. Instead of competition for food during the larval period, their decline in recent years is attributed to unfavorable conditions for spawning and/or egg survival on the spawning grounds at Hunter Shoals (TRM 521), primarily due to low river flows during drought years (Hickman et al. 1990). Increasing flows since 1989 coupled with a newly-instituted April minimum discharge of 4000 cfs in "dry" years or 8000 cfs during "wet" years for a three year test period at Watts Bar Dam (Yeager and Shiao 1992) should combine with the fingerling stocks by TWRA to improve the sauger fishery in Chickamauga Reservoir in the near future.

Freshwater drum

Decreases in YOY and intermediate freshwater drum (<u>Aplodinotus</u> <u>grunniens</u>) were significantly correlated with increasing aquatic macrophytes (Table 1). YOY drum, whose annual standing stocks were as high as 200/ha and 224/ha in 1972 and 1973, respectively, averaged 116/ha during Period I (Figure 26). In Period II standing stocks of YOY drum sharply decreased to only 4/ha. Their average abundance remained at 4/ha during Period III. In Period IV, following the decline of naiads, average density of YOY climbed to 69/ha, mostly due to a large increase in 1990 when 135/ha were found.

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Intermediate drum also declined relative to aquatic plant abundance, with average standing stocks dropping from 135/ha in Period I to 39/ha in Period II. Their peak abundance was 275/ha in 1974 while vegetation was still relatively sparse. During Period III average intermediate drum abundance (surprisingly) rose to 64/ha, roughly half the Period I average. In Period IV stocks of intermediate drum rose to an average of 103/ha, and like YOY stocks, had the greater abundance in 1990, 138/ha.

The pelagic young of freshwater drum consume cladocerans and copepods, as do other larval fishes. Aquatic insects, such as chironomids and <u>Hexagenia</u>, become more important as the young grow and become bottom feeders (Becker 1983). Fish, crayfish, and immature aquatic insects provide the bulk of adult drum diet (Pflieger 1975), although Asiatic clams (<u>Corbicula</u> spp.) are frequently consumed by Tennessee River drum.

Milfoil and Spinyleaf Naiad Growth Forms

Many fish species whose numbers increased with aquatic macrophytes reached their greatest abundances during the period when milfoil was the dominant form of aquatic vegetation, 1976-81. Their numbers declined as spinyleaf naiad replaced milfoil as the dominant macrophyte, 1982-88. Abundance of these phytophilic species was not just related to the total amount of vegetation present, but also to the dominant species of macrophyte present.

Differences in leaf morphology and underwater growth form of Eurasian watermilfoil and spinyleaf naiad provide different habitats for

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invertebrates and fish. Broad, feathery leaves of milfoil provide abundant surface area for aquatic invertebrates and epiphytic algae (Figure 27A). Bunched stems of milfoil grow from the substrate to the water surface. Near the surface the stems branch to form dense canopies. From substrate level milfoil colonies have the appearance of a forest with distinct trunks (i.e. bunched stems) and an overhead canopy. Since the water column below the canopy remains somewhat open, movement of fish is not entirely restricted. Milfoil colonies usually are most abundant at depths between 1.5 and 3 m. Strands of milfoil persist into the winter, providing habitat for aquatic life well beyond the seasonal growth period.

In contrast, the growth form of the annual macrophyte spinyleaf naiad is dense, bushy, and brittle (Figure 27B). It usually grows in shallower areas than milfoil, often in backs of coves and along shorelines, and has a high density of slender, wire-like stems. During the drought years of the 1980s when the water was unusually clear, this plant expanded its coverage to deeper water (1-2 m) to areas once colonized by milfoil. In shallow water spinyleaf naiad may not form a distinct canopy near the surface. Instead its "Brillo pad" growth form may occupy a large portion of the water column. Fish are less able to move within this growth form than within milfoil. Although some foraging and protection benefit is afforded, spinyleaf naiad may actually decrease shoreline habitat for small fish due to its dense, bushy growth. Furthermore, spinyleaf naiad plants generally "break-up" by September, leaving shoreline areas devoid of vegetated cover for fish and aquatic invertebrates in the mid-fall, winter, and early spring months.

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Factors Affecting Aquatic Macrophytes

Univariate regression analysis found that of the variables tested the amount of aquatic macrophyte coverage during the previous year had the greatest influence (r=0.90, P=0.0001) on coverage during the next year (Figure 28). In other words, propagules (milfoil root masses, fragments of milfoil stems, and seed banks of annual species) from the previous season are of prime importance to the magnitude of macrophyte coverage during the following season. Paucity of aquatic vegetation upstream from Chickamauga Reservoir also lessens the likelihood of recolonization by floating fragments. Favorable abiotic growing conditions alone will not produce abundant vegetation.

Sunlight availability was also highly correlated (r=0.68, P=0.0005) with aquatic macrophyte coverage (Figure 29). Sunlight availability in this model was the percentage of possible sunshine reaching the earth's surface. (Actual amounts of sunlight penetrating the reservoir's water surface in vegetated areas, measured with a secchi disc, would have been more appropriate, but were not available for the study period.) As with terrestrial plants, sunlight, the energy source for photosynthesis, has been documented as the most important environmental factor affecting submersed aquatic macrophyte abundance (Barko et al. 1986, Johnstone and Robinson 1987).

During the period 1971-92 aquatic macrophyte coverage was inversely related (r=0.54, P=0.0100) to discharges of the Tennessee River at Chickamauga Dam during the critical part of the growing season (March to June) (Figure 30). Factors associated with high discharges that

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negatively affect growth of aquatic macrophytes are increased scouring, siltation, and turbidity caused by suspended solids. Scouring action during high flows may uproot plants or cause fragmentation that results in reduced abundance of aquatic vegetation. Siltation can smother root masses, small plants, and bury seed banks with sediment. Turbidities due to suspended solids absorb sunlight penetrating the water's surface, thereby depriving submersed macrophytes of their photosynthetic energy source. A recent study of Chickamuga embayments also found aquatic vegetation negatively correlated to turbidity (Meinert et al. 1992).

A biogenic source of turbidity that may be important during some years is excessive growth of phytoplankton. Phytoplankton absorb sunlight, and like suspended solids may limit the amount of sunlight available for macrophytes. Although insufficient chlorophyll <u>a</u> and secchi depth data precluded regression analysis, it would be expected that reduced light availability caused by phytoplankton blooms would negatively affect aquatic macrophyte growth and coverage. An unusually large phytoplankton bloom in Chickamauga Reservoir during spring 1990 (Wayne Poppe, TVA, personal communication) may have suppressed aquatic macrophyte growth that year. Excessive phytoplankton blooms and epiphyton growth, associated with eutrophication, have led to reductions of submersed aquatic macrophytes in Chesapeake Bay (Orth and Moore 1983) and Europe (De Nie 1987).

As in other biological processes, temperature affects the growth rate of aquatic vegetation. However, when mean March-June water temperatures for years 1975-92 were regressed against macrophyte coverage, the

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relationship was not significant (P=0.4875). This was because two of the lowest mean water temperatures occurred during two of the highest vegetation years, 1983 and 1984 (Figure 31); and the effect of cooler water was overridden by the amount of previous years' vegetation coverage. Removing these two years from the regression yielded a significant relationship (r=0.55, P=0.0290).

Two multiple regression models explained large percentages of the variance in aquatic macrophyte coverage in Chickamauga Reservoir, 1975-92. The better model was:

Sumbersed aquatic macrophytes, hectares =

- 8347.71 + 0.67(Previous season's coverage, hectares)

+ 153.15(Sunlight availability, percentage)

where: $R^2 = 0.85 F = 43.02 Prob > F = 0.0001; n = 18.$

The second model was:

Sumbersed aquatic macrophytes, hectares =

2814.26 + 0.77(Previous season's coverage, hectares)

- 0.07(Average March-June daily discharge, cfs)

where: $R^2 = 0.83 F = 36.6$; Prob>F = 0.0001; n = 18.

Because sunlight and discharge were highly correlated (r = -0.84, P = 0.0001), it was inappropriate to include both in the same model.

Herbivorous animals, such as certain species of turtles, can affect the establishment and regrowth of some aquatic macrophytes. This may be especially important when vegetation is sparse. Turtles are trapped and removed from experimental vegetation plantings on Guntersville Reservoir to allow establishment of desirable plant species (Doug Murphy, TVA, personal communication).

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Effects of SQN Operation on Aquatic Macrophytes

Because this report has identified populational levels of submersed aquatic macrophytes as having a dramatic effect on the resident fish community in Chickamauga Reservoir, a brief discussion and evaluation of the operational impacts of SQN on aquatic macrophyte communities is included to assess potential indirect impacts to the resident fish community. Macrophyte coverage data used in this analysis would detect only major changes rather than restricted or localized impacts associated with SQN operation.

Since initial operation in 1981, SQN has had no documented effect on growth or distribution of aquatic vegetation in Chickamauga Reservoir. After four years of operation, similarities in trends of vegetation coverage in littoral habitat along mainchannel areas upstream and downstream of SQN showed no significant effect on aquatic macrophyte communities (TVA 1985). During most years between 1977 and 1992, vegetation coverage fluctuated similarly in mainchannel habitat downstream of SQN and upstream of SQN, regardless of SQN operational status (Figure 32). During drought years from 1984 to 1988 aquatic macrophyte coverage remained relatively stable or increased in mainchannel habitat both upstream and downstream of SQN. Beginning in 1989 aquatic macrophyte coverage has declined dramatically on Chickamauga Reservoir (Figure 4) in mainchannel areas both upstream and downstream of SQN. In 1992 there was only about 155 ha of aquatic macrophytes in Chickamauga Reservoir (Table 2).

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Significant changes that have occurred in aquatic macrophyte populations on Chickamauga Reservoir are considered to be primarily related to environmental factors associated with natural climatic events such as sunlight availability, water clarity, and flow and not the operational status of SQN. The increase in aquatic macrophyte coverage during the drought years from 1984 to 1988, followed by a dramatic decline in 1989 and 1990 is similar to that of other mainstream reservoirs such as Kentucky, Wheeler, and Guntersville (Figure 5). Because changes in aquatic macrophyte communities are related to factors other than the operational status of SQN, indirect effects of SQN to fish populations are minimal.

CONCLUSIONS

Aquatic macrophytes in the littoral zones of Chickamauga Reservoir have a profound effect on the fish community. Total numbers and biomass of all fish species combined increased with submersed aquatic vegetation coverage. Certain species (golden shiner, yellow bass, warmouth, bluegill, redbreast sunfish, largemouth bass, black crappie, yellow perch, and brook silverside) are favored by increasing vegetation, while others (carp, smallmouth buffalo, spotted sucker, channel catfish, white crappie, and freshwater drum) are hindered. The forage base for piscivorous predators is primarily shad when aquatic vegetation is sparse and small sunfish when vegetation is dense.

The dominant species of vegetation, i.e., milfoil and naiads, differ in leaf morphology and growth form and provide different habitats for

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invertebrates and fish. This is believed to affect the abundance of many fish species in Chickamauga Reservoir. Based on preliminary analysis, milfoil appears more beneficial to most of the important gamefish species than spinyleaf naiad because of its growth form.

Black crappie and white crappie abundances are oppositely affected by dense growth of aquatic macrophytes. Young black crappie benefit from an increased invertebrate food source and protective habitat within macrophyte colonies, while white crappie prefer deeper, open-water habitats and do not benefit from shoreline vegetation. Both species are in competition for zooplanktonic food with enormous numbers of young gizzard and threadfin shad and, in seasons of abundant aquatic macrophytes, young of three sunfish species. Clear water in Chickamauga Reservoir during the drought years of the 1980s also contributed to the shift in dominance from white crappie to black crappie.

Decreases in aquatic macrophyte coverage since 1989 should result in a restructuring of the fish community that would reverse some of the trends presented in this report. Species that increased with heavy infestations of aquatic macrophytes are expected to decrease in abundance, while open-water species should increase. White crappie are expected to return as the dominant crappie species in Chickamauga Reservoir.

Factors identified as affecting submersed aquatic macrophyte coverage were macrophyte coverages from the previous year, sunlight availability, and discharges of the Tennessee River. Water temperature

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was an important factor in most years. Two factors affecting water clarity, suspended solids and phytoplankton, could not be identified as important factors given the limited amount of available data.

There is no evidence that SQN operation affected the coverage of submersed aquatic macrophytes in Chickamauga Reservoir; thus, there were no indirect effects to the resident fish community.

RECOMMENDATIONS

Further studies of fish communities within milfoil and spinyleaf naiad colonies are needed to verify that differences in community structure of fishes are related to the growth forms of the dominant vegetation species. The present study compared fish densities on an annual basis, and in many years the habitat of the coves was altered by herbicide treatment to facilitate rotenone sampling. Recommended future studies would involve taking rotenone samples in two milfoil-dominated coves and in two similar coves where spinyleaf naiad is the dominant macrophyte. Herbicides would not be used in any of the four coves. All four coves would be sampled during the same summer, and would not be the coves routinely sampled for SQN monitoring.

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Species	Size Group	Slope	F-Value	PR > F
ncreasing species				
Total species	YNG_NO	0.00009	52.953	0.0001
Fotal species	YNG_WT	0.00005	16.842	0.0001
Fotal species	INT_NO	0.00004	12.612	0.0006
fotal species	ADT_NO	0.00004	9.701	0.0025
Total species	ADT_WT	0.00002	5.533	0.0211
Gizzard shad	YNG_NO	0.00021	15.052	0.0002
Gizzard shad	ADT_NO	0.00005	5.495	0.0215
Golden shiner	YNG_NO	0.00020	31.766	0.0001
Carp	YNG_NO	0.00007	17.213	0.0001
Brook silverside	YNG_NO	0.00003	18.379	0.0001
Cellow bass	YNG_NO	0.00008	5.608	0.0203
Cellow bass	INT_NO	0.00010	15.958	0.0001
Cellow bass	ADT_NO	0.00007	17.929	0.0001
Narmouth	YNG_NO	0.00029	101.323	0.0001
larmouth	INT_NO	0.00012	43.691	0.0001
Marmouth	ADT_NO	0.00008	16.719	0.0001
Bluegill	YNG_NO	0.00011	28.029	0.0001
Bluegill	INT_NO	0.00004	8.434	0.0047
ledear sunfish	YNG_NO	0.00029	90.765	0.0001
ledear sunfish	INT_NO	0.00009	16.591	0.0001
argemouth bass	YNG_NO	0.00008	13.553	0.0004
argemouth bass	INT_NO	0.00007	16.721	0.0001
Black crappie	YNG_NO	0.00007	7.933	0.0061
Black crappie	INT_NO	0.00005	12.125	0.0008
Yellow perch	YNG_NO	0.00011	10.140	0.0021
ecreasing species				
arp	ADT_NO	-0.00007	13.856	0.0004
mallmouth buffalo	INT_NO	-0.00004	8.922	0.0037
mallmouth buffalo	ADT_NO	-0.00005	6.777	0.0110
potted sucker	YNG_NO	-0.00009	10.875	0.0014
potted sucker	ADT_NO	-0.00008	13.301	0.0005
hannel catfish	YNG_NO	-0.00004	8.474	0.0047
hannel catfish	INT_NO	-0.00009	28.297	0.0001
hite crappie	ADT_NO	-0.00011	30.338	0.0001
auger	YNG_NO	-0.00003	8.439	0.0047
auger	INT_NO	-0.00002	6.717	0.0113
auger	ADT_NO	-0.00002	5.928	0.0171
reshwater drum	YNG_NO	-0.00017	26.305	0.0001
reshwater drum	INT_NO	-0.00005	5.398	0.022

Table 1. Fish species showing significant correlation in numerical abundance (numbers/hectare) or weight (kg/hectare) and aquatic macrophyte coverage in Chickamauga Reservoir, 1970-90 by size group*.

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* YNG_NO = YOY/ha, INT_NO = intermediates/ha, ADT_NO = adults/ha, YNG_WT + YOY kg/ha, ADT_WT = adult kg/ha. Table 2. Aquatic macrophytes (hectares) in Chickamauga Reservoir, 1970-92.*

Year	Total	Milfoil	Naiads	Pondweeds	Mixed	Algae
1970	11	-	-	-		-
1971	13	-	-	-	-	-
1972	20	-	-	-	-	-
1973	4	-	-	-	-	_
1974	13	-	-	-	-	-
1975	77	_	-	-	-	-
1976	293	-	-	-	-	-
1977	422	-	-	-	-	-
1978	802	190	612	0	tr	0
1979	635	-	-	-	-	-
1980	1,328	482	844	1	0	0
1981	2,188	1,225	879	79	4	0
1982	2,626	701	1,827	85	13	0
1983	2,791	759	1,921	2	48	60
1984	2,161	292	1,788	0	116	3
1985	2,275	341	1,799	13	114	8
1986	2,778	477	1,853	3	438	7
1987	2,770	465	1,994	23	267	21
1988	3,017	849	1,901	8	251	8
1989	1,388	869	312	11	221	tr
1990	861	638	69	17	134	4
1991	275	100	8	11	149	tr
1992	155	71	27	9	24	6

tr--less than 0.5 * Coverages for 1970-77 and 1979 are estimates from surveys and herbicide treatment records; coverages for 1978 and 1980-92 were determined by aerial photography.

Table 3. Angling summary for black bass in Chickamauga Reservoir, 1977-92.*

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Year	Estimated Hours	Estimated Trips	Estimated Number Harvested	Estimated Weight Harvested	Harvest Rate (No./Hr)	Mean Weight (Lbs.)	Estimated Weight Per Acre
1977	35,331	8,251	5,014	6,736	0.141	1.34	0.19
1978	28,418	6,675	5,931	6,792	0.208	1.15	0.19
1979	44,171	9,830	11,930	14,010	0.270	1.17	0.39
1980	123,020	28,829	17,760	25,989	0.144	1.46	0.73
1981	123,304	26,909	22,730	28,027	0.184	1.23	0.79
1982	163,736	36,088	31,332	41,000	0.191	1.31	1.15
1983	132,551	27,022	39,983	48,765	0.301	1.22	1.37
1984	113,106	23,310	36,654	41,720	0.324	1.14	1.17
1985	135,017	29,840	23,006	27,943	0.170	1.21	0.78
1986	-	-	-	· -	-	-	-
1987	291,517	58,733	46,355	57,996	0.159	1.25	1.63
1988	336,764	114,336	125,686	70,233	0.373	0.56	1.98
1989	273,732	53,399	43,477	54,795	0.158	1.26	1.54
1990	449,698	86,198	42,030	54,314	0.093	1.29	1.53
1991	598,039	189,486	63,971	84,542	0.106	1.32	2.39
1992	281,895	104,793	30,846	41,951	0.120	1.36	1.19

* TWRA 1992, 1993.

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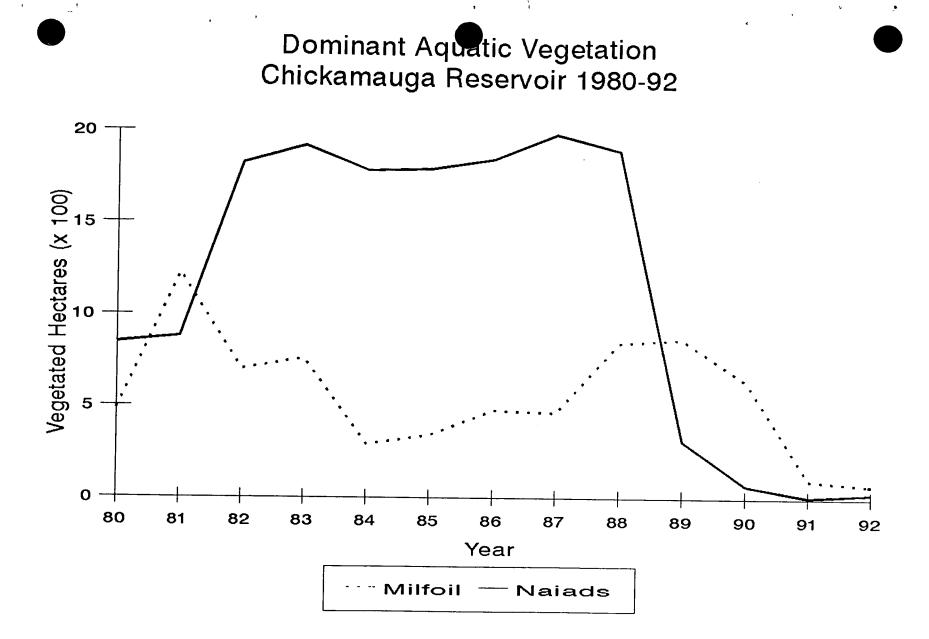


Figure 1. Coverages (hectares) of Eurasian watermilfoil and spinyleaf naiad in Chickamauga Reservoir, 1980-92.

-63-

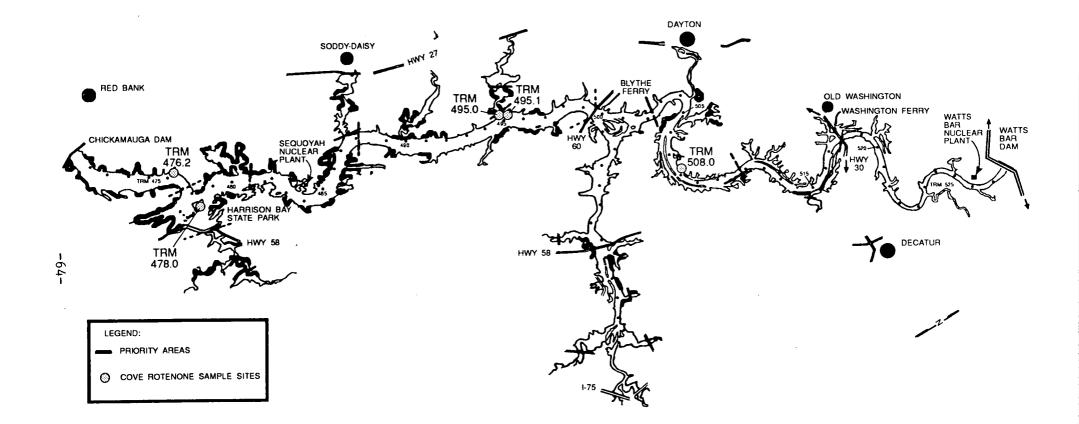


Figure 2. Locations of cove rotenone sample sites and priority areas treated with herbicides in Chickamauga Reservoir, 1970-90.

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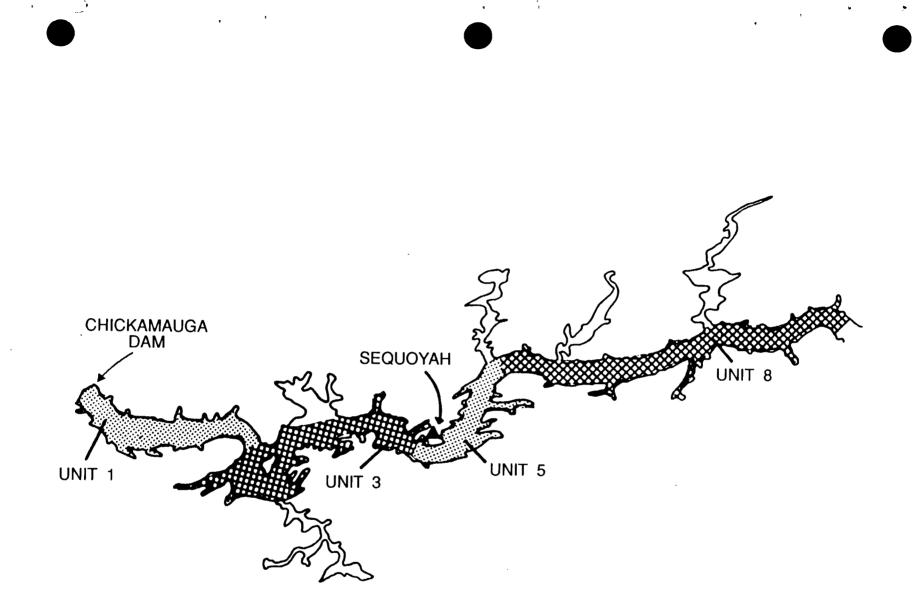


Figure 3. Mainchannel aquatic plant management units on Chickamauga Reservoir in the vicinity of SQN.

-65-

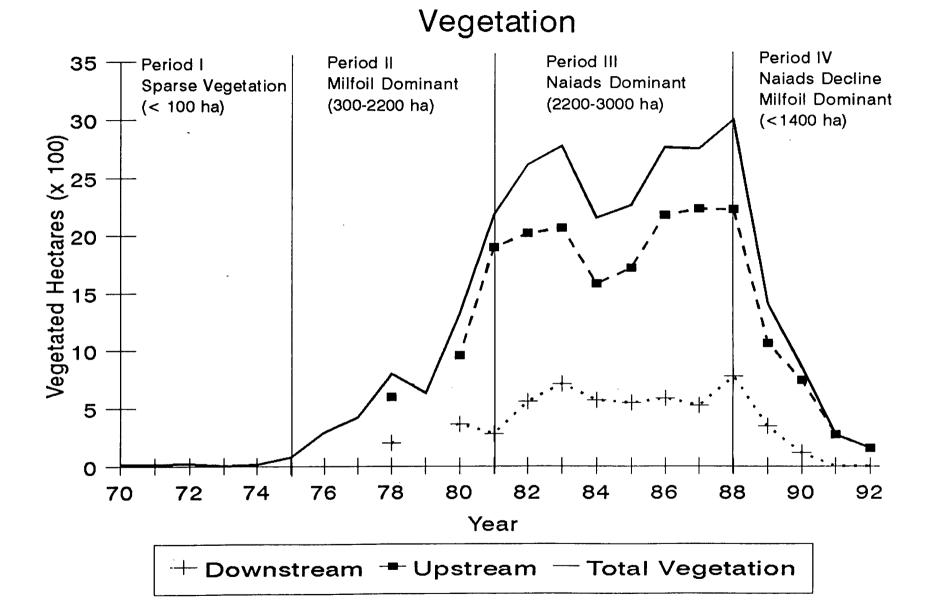


Figure 4. Total coverage of aquatic macrophytes in Chickamauga Reservoir (solid line), upstream and downstream from SQN (dashed lines), and four time periods based on extent of macrophyte coverage and dominant species present, 1970-92.

-66-



Aquatic Macrophyte Coverage

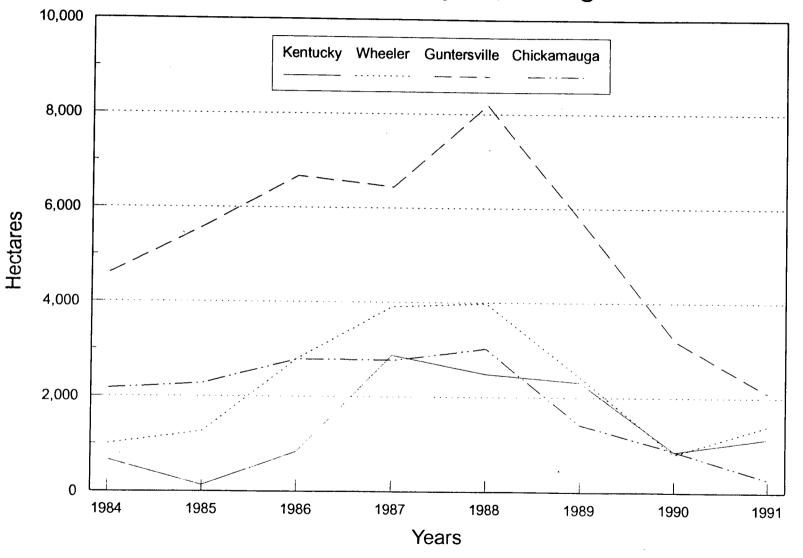


Figure 5. Total macrophyte coverages of four TVA mainstream reservoirs, 1984-91.

-67-



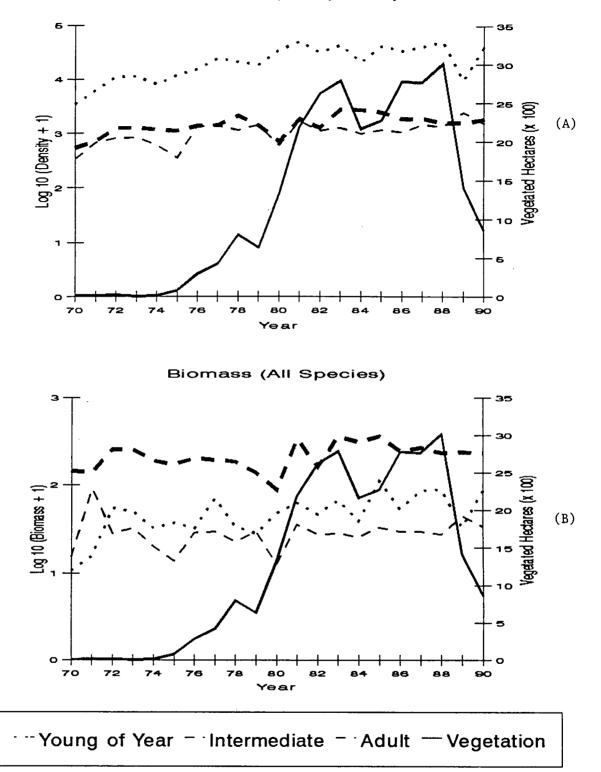


Figure 6. Annual standing stock densities (A, numbers/hectare) of all species combined and biomass (B, kg/hectare) as determined by cove rotenone sampling, 1970-90.

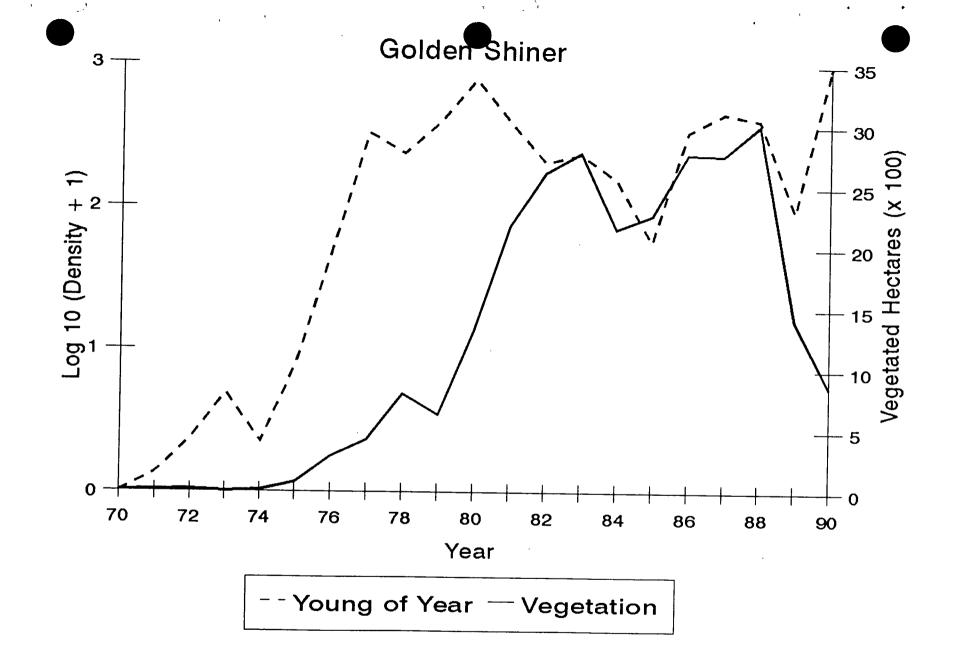


Figure 7. Annual standing stock densities (numbers/hectare) of golden shiner, as determined by cove rotenone sampling, 1970-90.

-69-

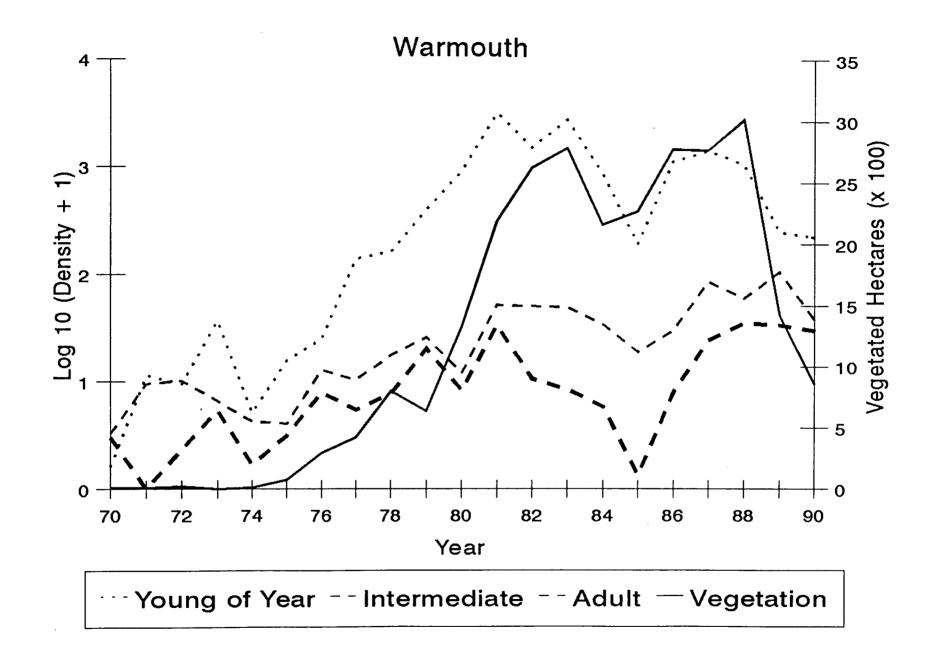


Figure 8. Annual standing stock densities (numbers/hectare) of warmouth, as determined by cove rotenone sampling, 1970-90.

-70-

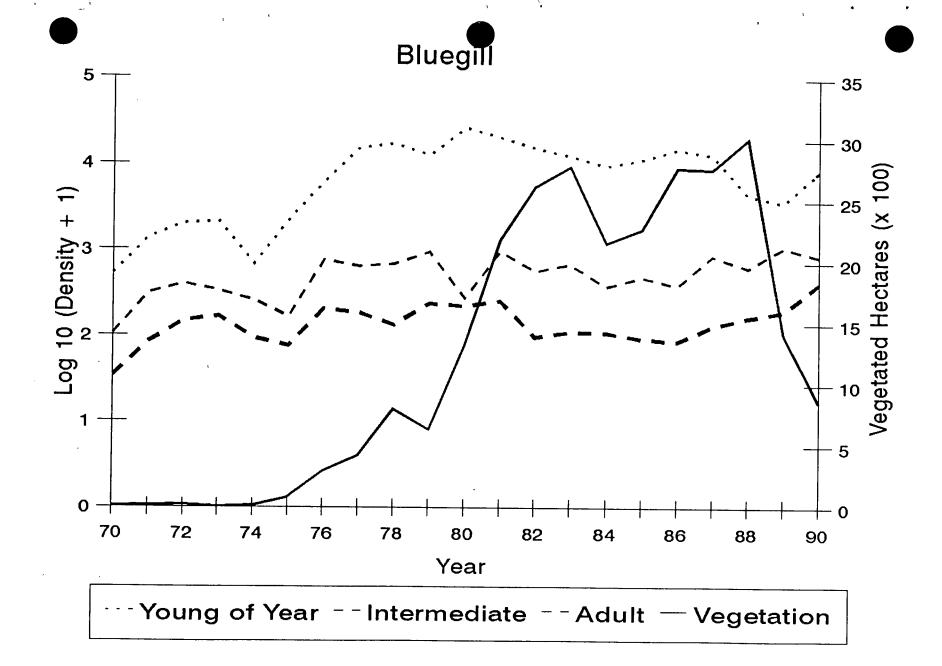
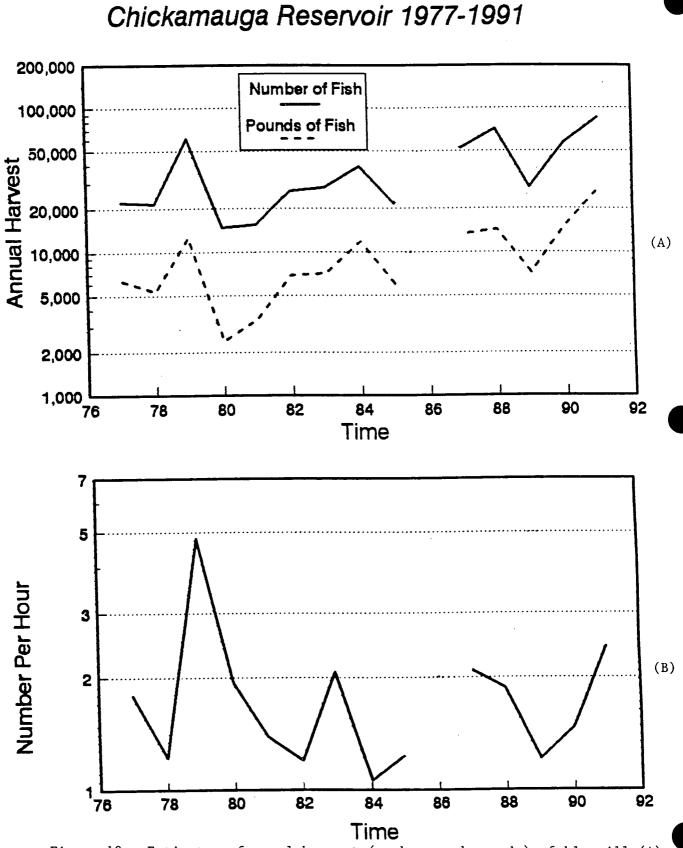


Figure 9. Annual standing stock densities (numbers/hectare) of bluegill, as determined by cove rotenone sampling, 1970-90.

-71-



Bluegill Creel Data

Figure 10. Estimates of creel harvest (numbers and pounds) of bluegill (A) and catch per hour, (B) 1977-91 (TWRA data).

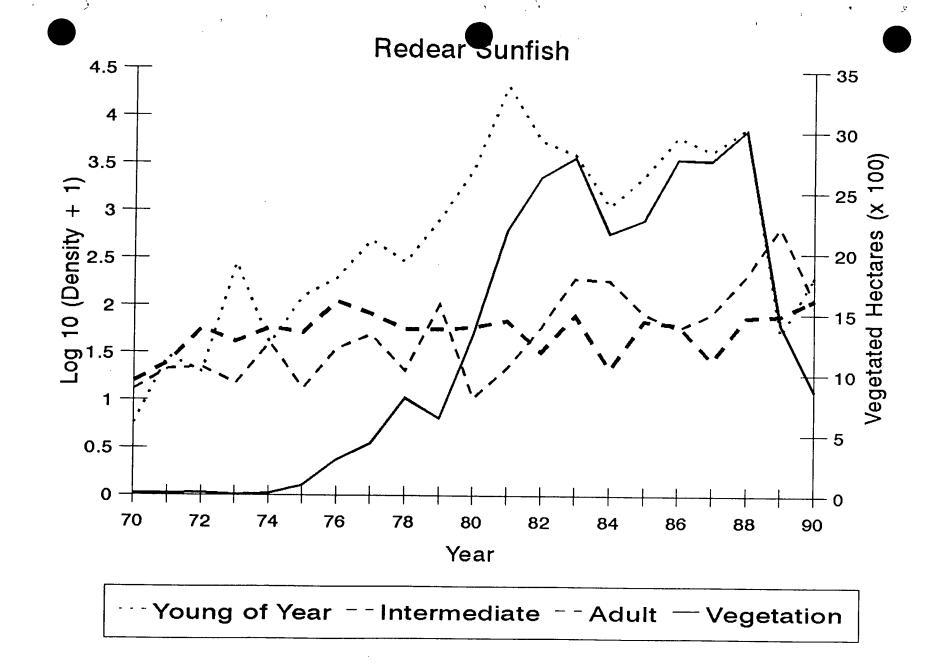


Figure 11. Annual standing stock densities (numbers/hectare) of redear sunfish, as determined by cove rotenone sampling, 1970-90.

-73-

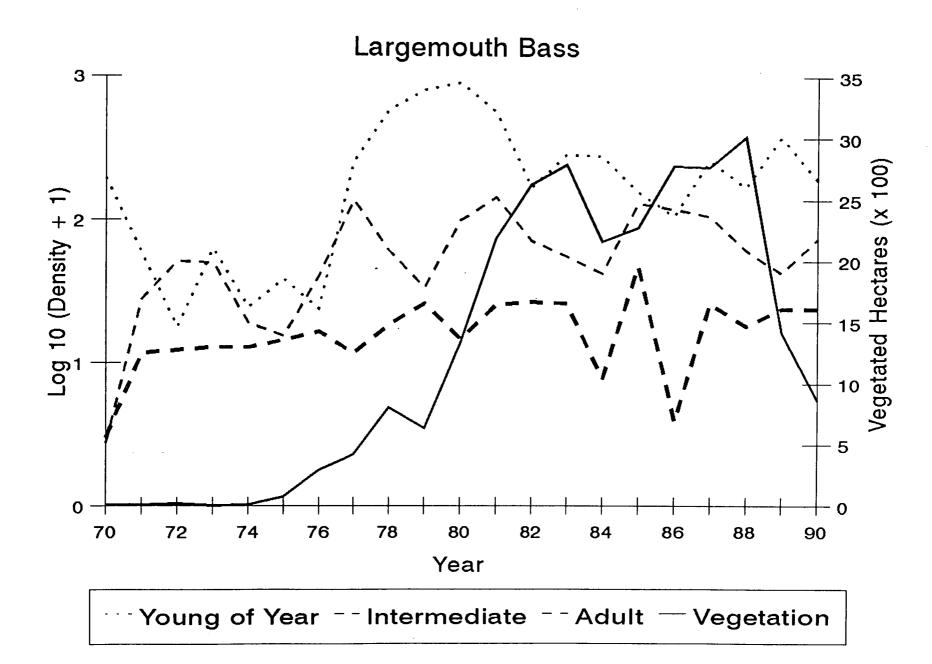
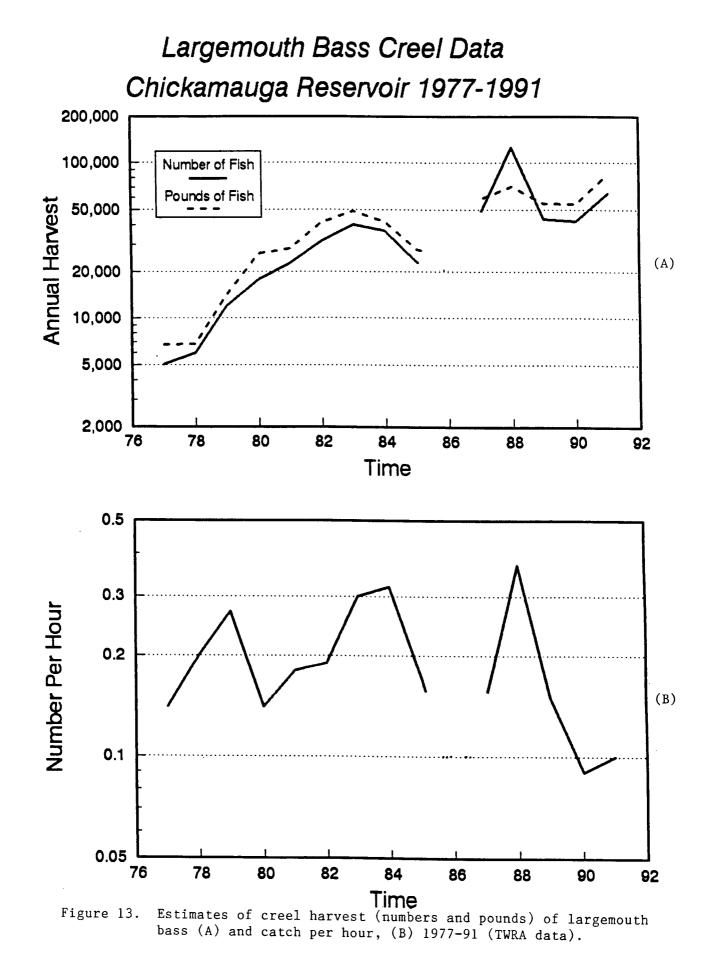


Figure 12. Annual standing stock densities (numbers/hectare) of largemouth bass, as determined by cove rotenone sampling, 1970-90.

-74-



-75-

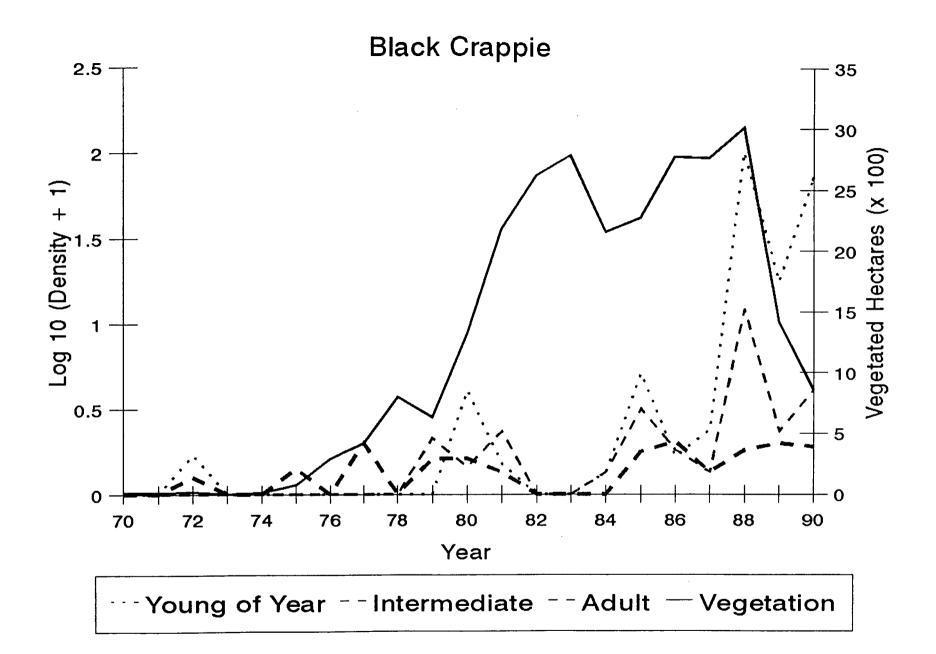


Figure 14. Annual standing stock densities (numbers/hectare) of black crappie, as determined by cove rotenone sampling, 1970-90.

-76-

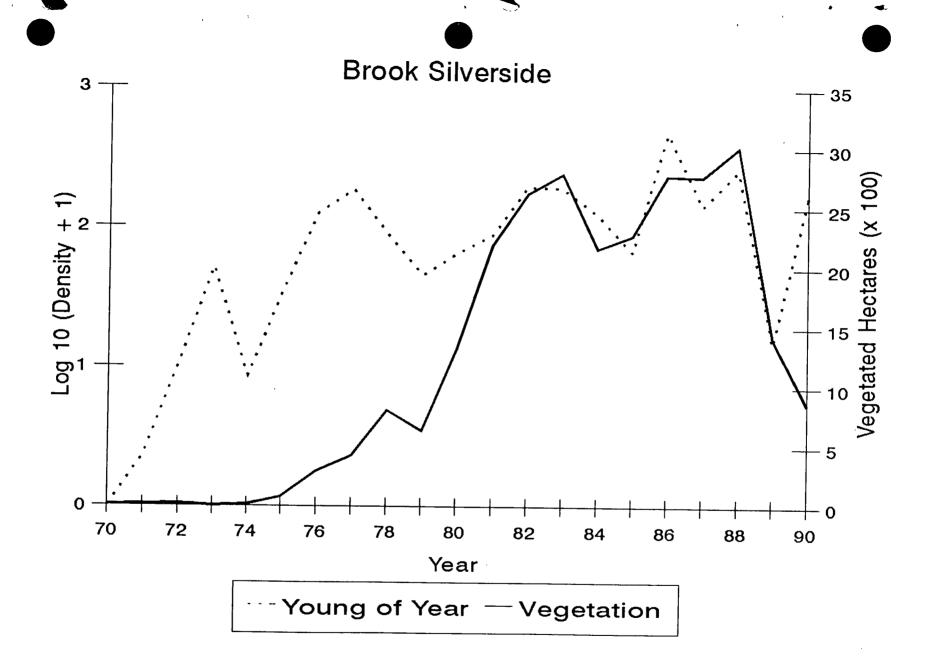


Figure 15. Annual standing stock densities (numbers/hectare) of black crappie, as determined by cove rotenone sampling, 1970-90.

-77-

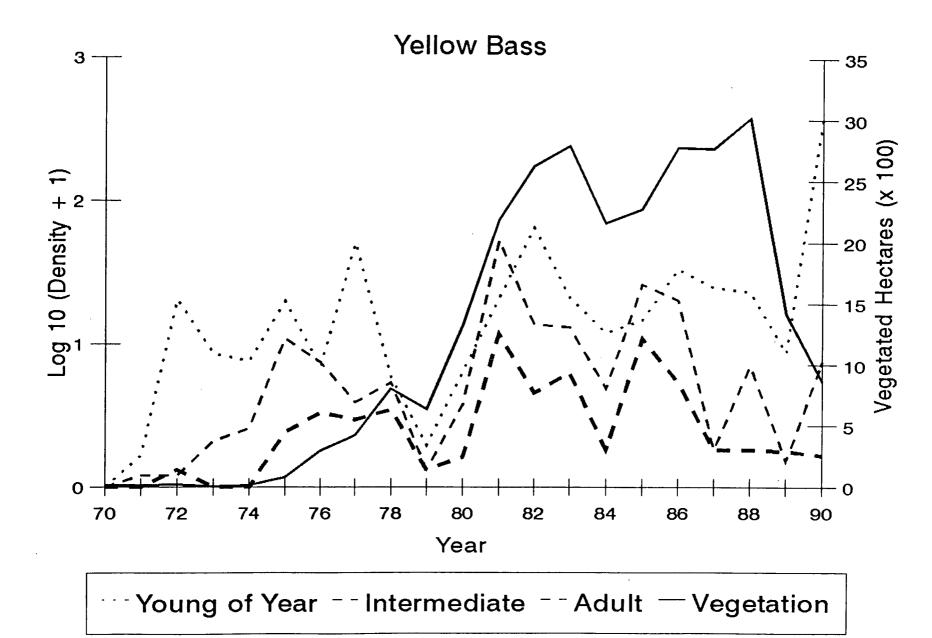


Figure 16. Annual standing stock densities (numbers/hectare) of yellow bass, as determined by cove rotenone sampling, 1970-90.

-78-

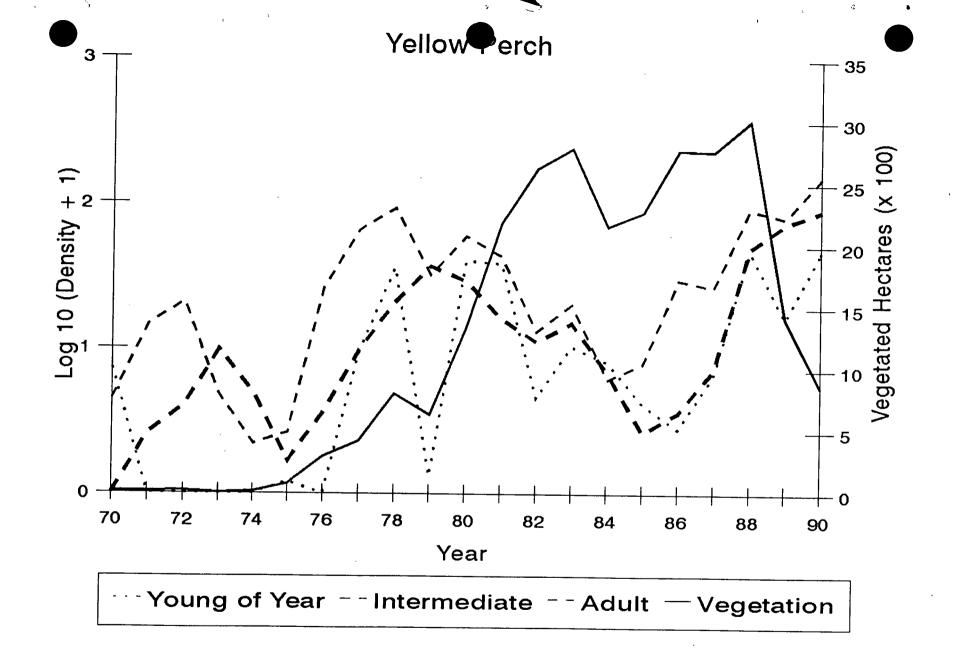


Figure 17. Annual standing stock densities (numbers/hectare) of yellow perch, as determined by cove rotenone sampling, 1970-90.

-79-

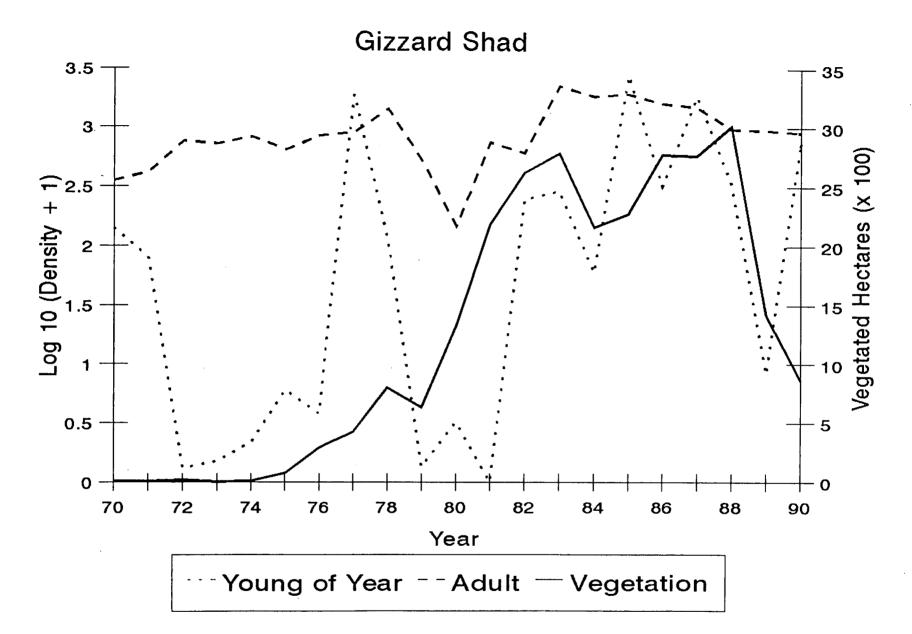


Figure 18. Annual standing stock densities (numbers/hectare) of gizzard shad, as determined by cove rotenone sampling, 1970-90.

-80-

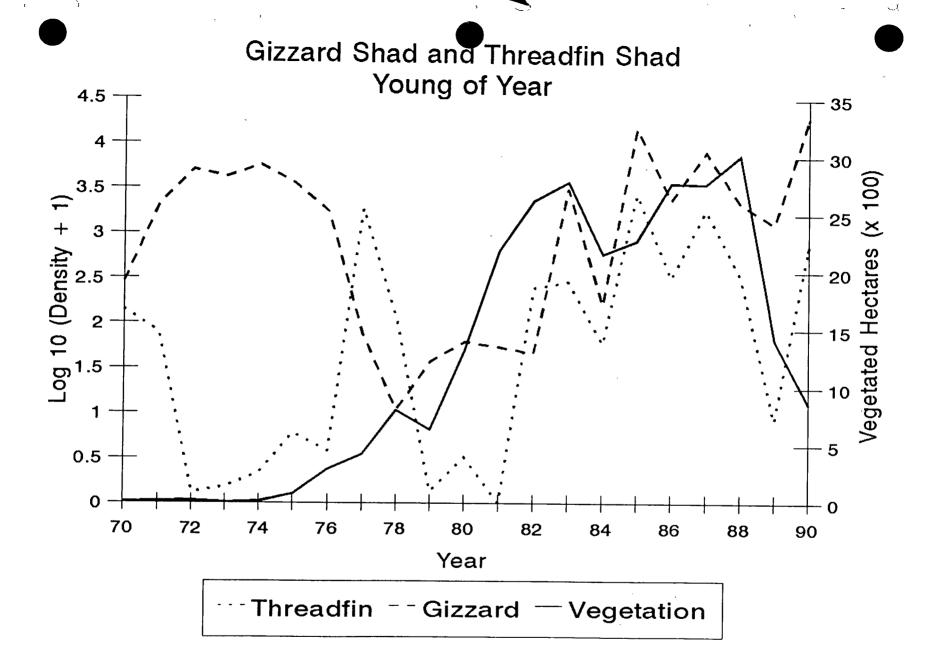


Figure 19. Annual standing stock densities (numbers/hectare) of YOY gizzard shad and threadfin shad, as determined by cove rotenone sampling, 1970-90.

-81-

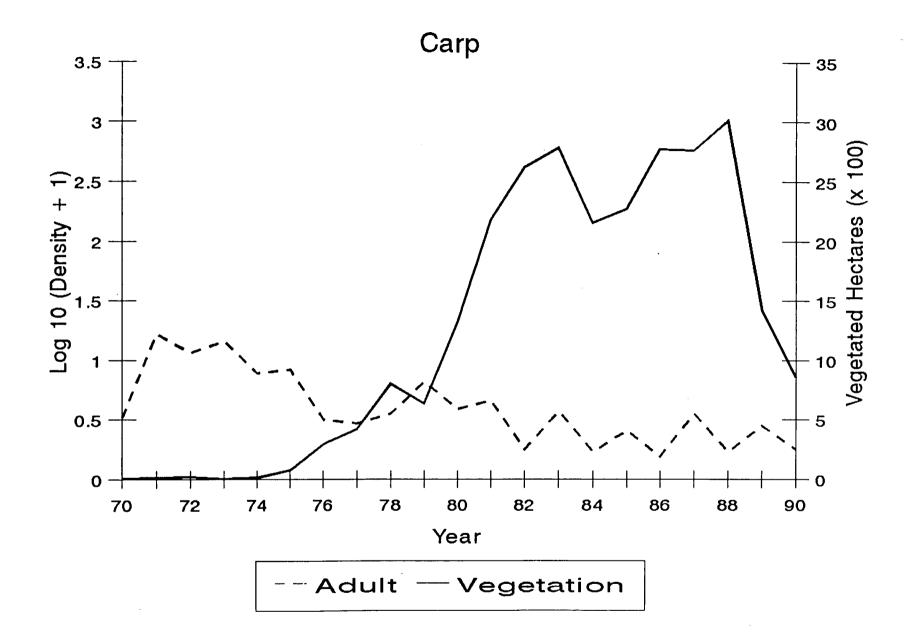


Figure 20. Annual standing stock densities (numbers/hectare) of carp, as determined by cove rotenone sampling, 1970-90.

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Smallmouth Buffalo

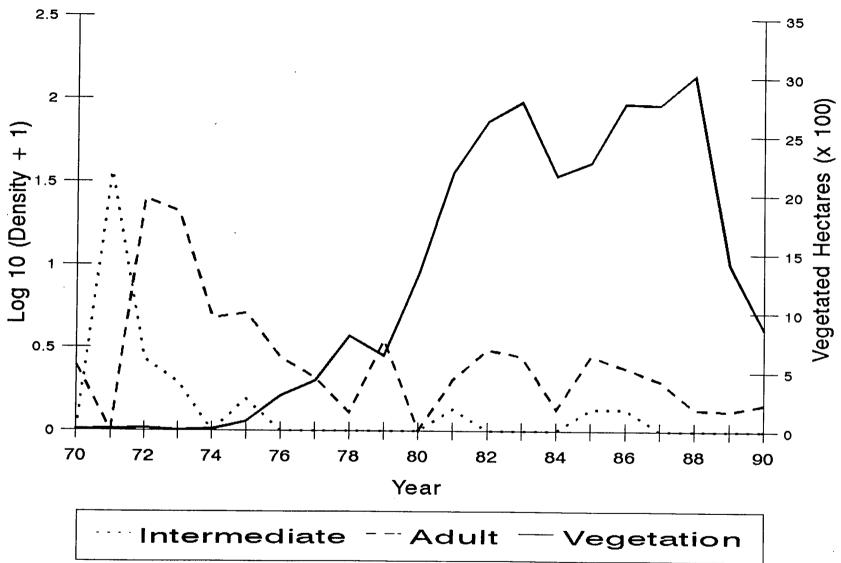
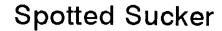


Figure 21. Annual standing stock densities (numbers/hectare) of smallmouth buffalo, as determined by cove rotenone sampling, 1970-90.

-83-



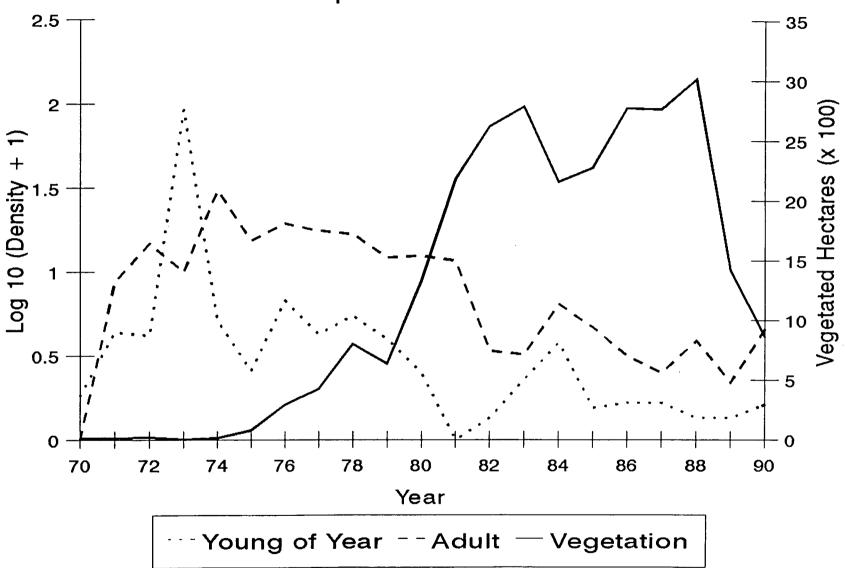


Figure 22. Annual standing stock densities (numbers/hectare) of spotted sucker, as determined by cove rotenone sampling, 1970-90.

-84-

Channel Catfish

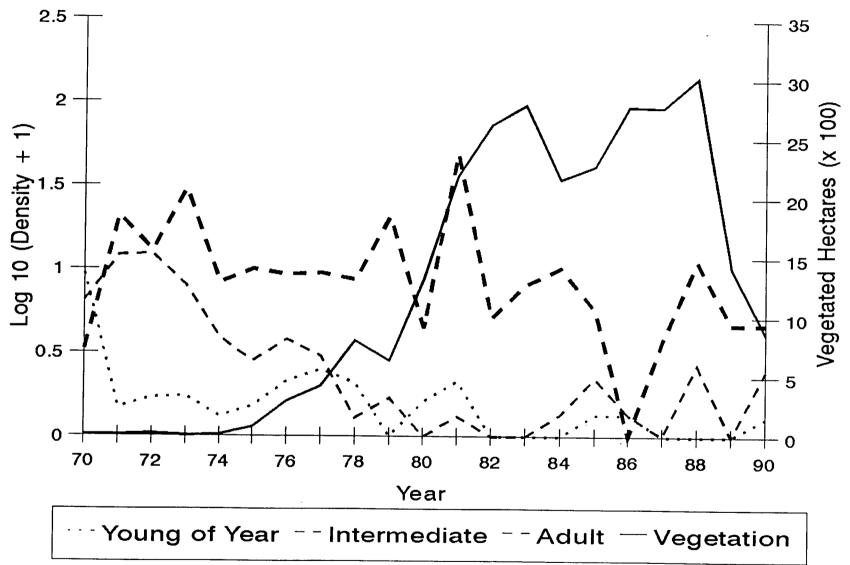


Figure 23. Annual standing stock densities (numbers/hectare) of channel catfish, as determined by cove rotenone sampling, 1970-90.

-85-

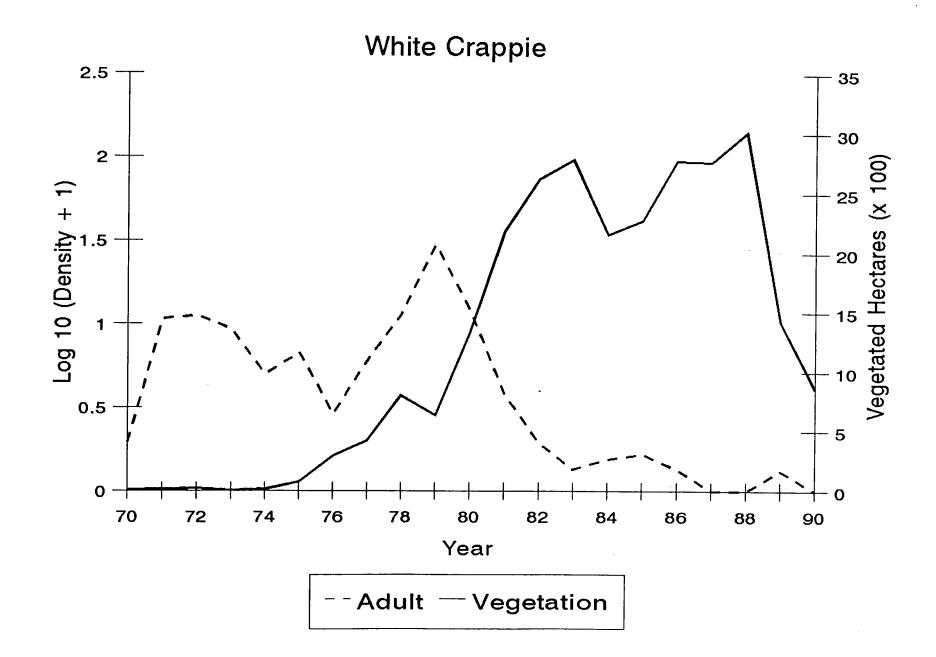


Figure 24. Annual standing stock densities (numbers/hectare) of white crappie, as determined by cove rotenone sampling, 1970-90.

-96-

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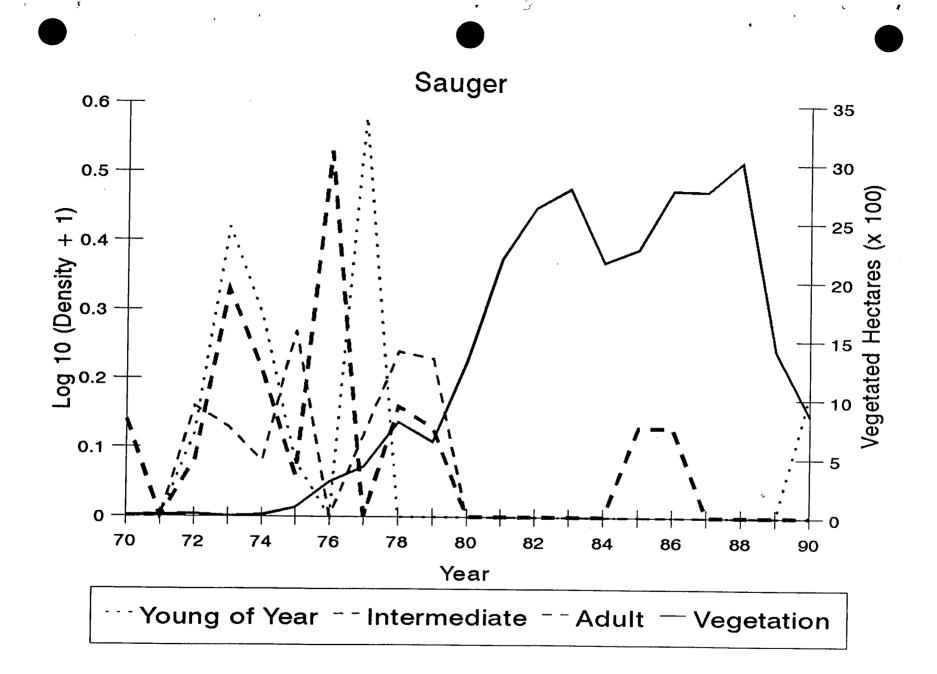


Figure 25. Annual standing stock densities (numbers/hectare) of sauger, as determined by cove rotenone sampling, 1970-90.

-87-

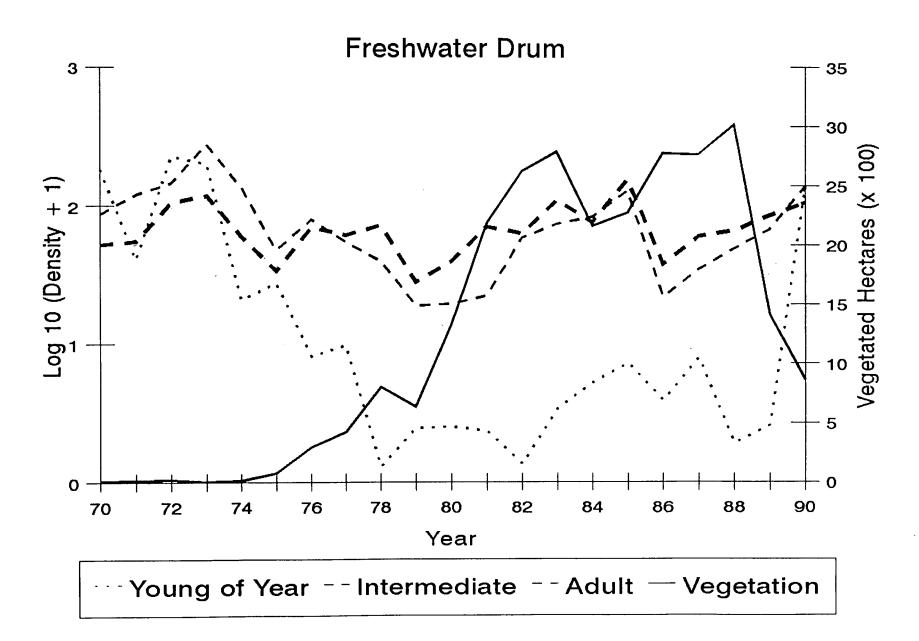
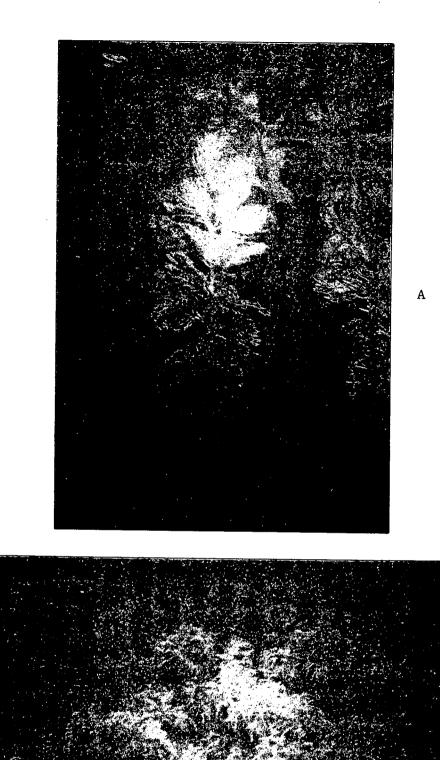


Figure 26. Annual standing stock densities (numbers/hectare) of freshwater drum, as determined by cove rotenone sampling, 1970-90.



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Figure 27. Underwater photographs of Eurasian watermilfoil (A) and spinyleaf naiad (B) illustrating differences in growth forms.

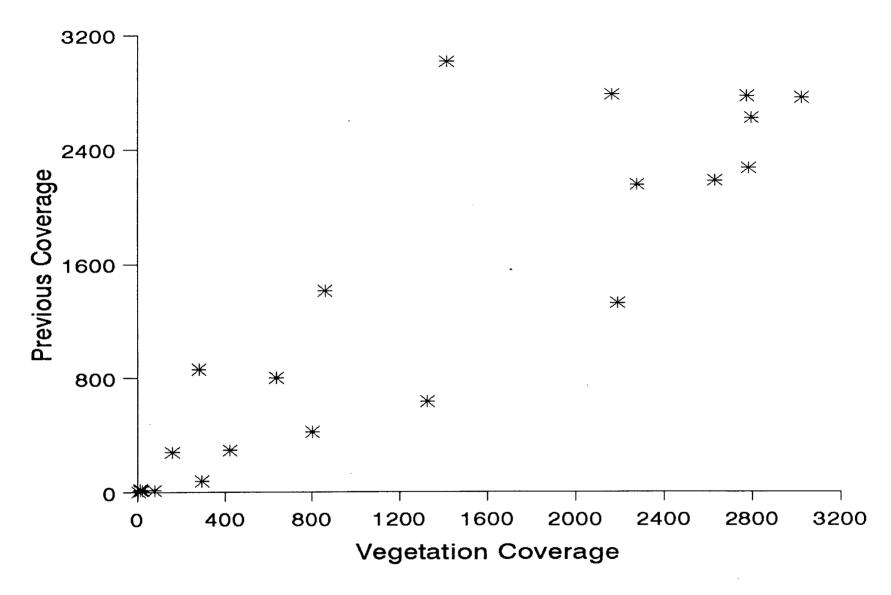
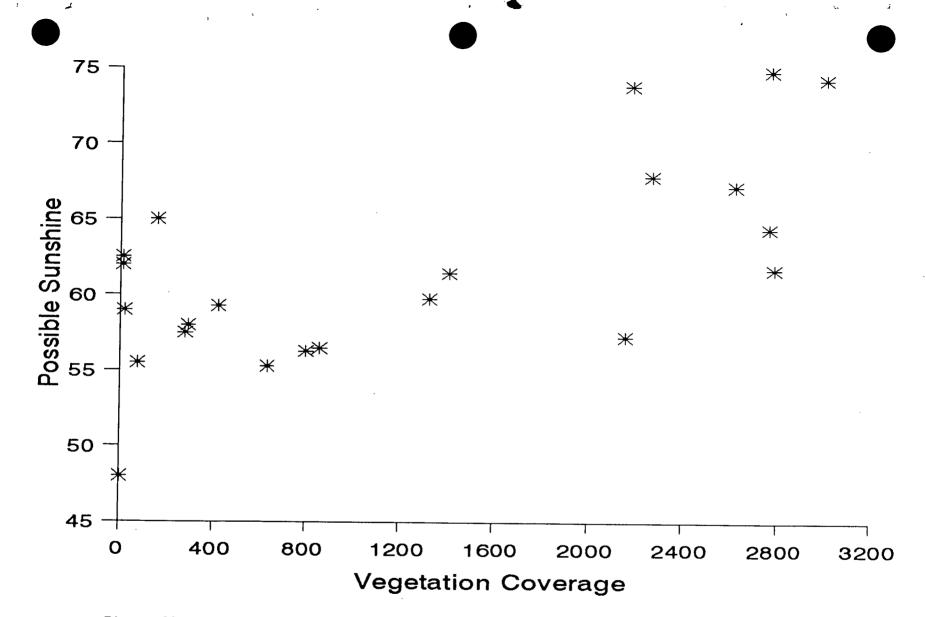
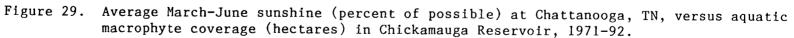


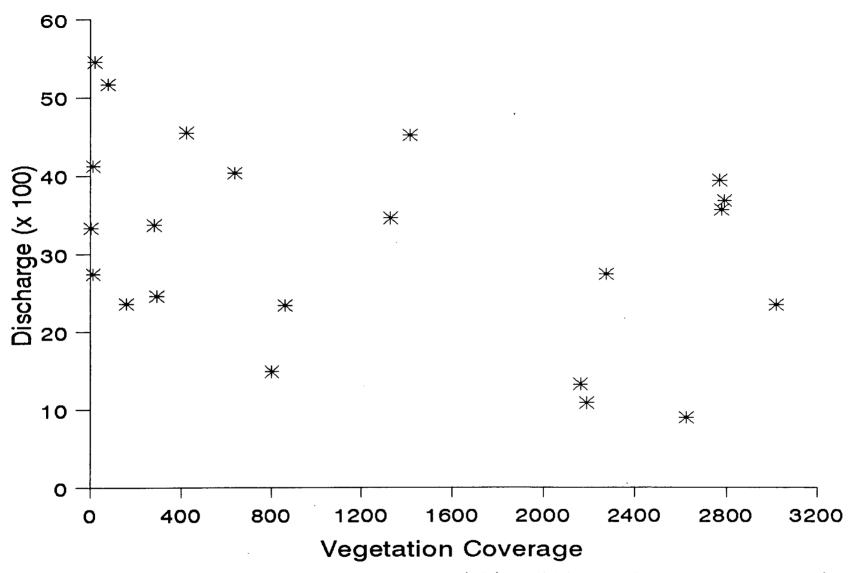
Figure 28. Previous year's aquatic macrophyte coverage (hectares) versus current year's coverage in Chickamauga Reservoir, 1971-92.

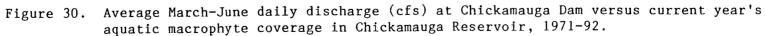
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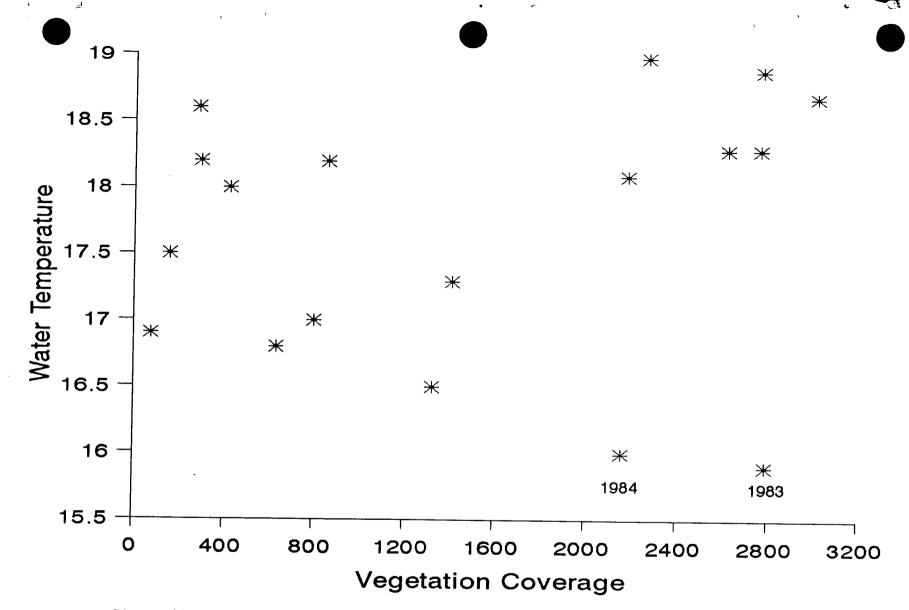


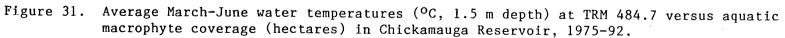
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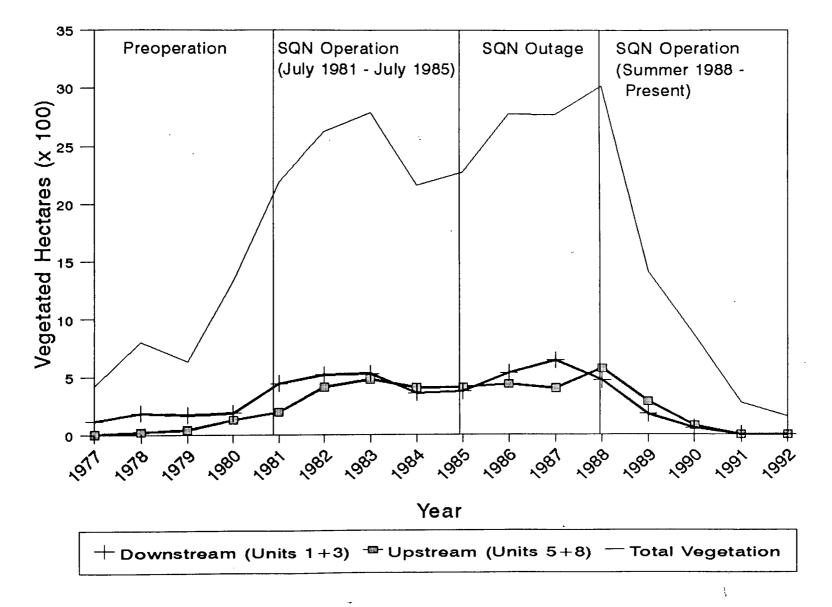
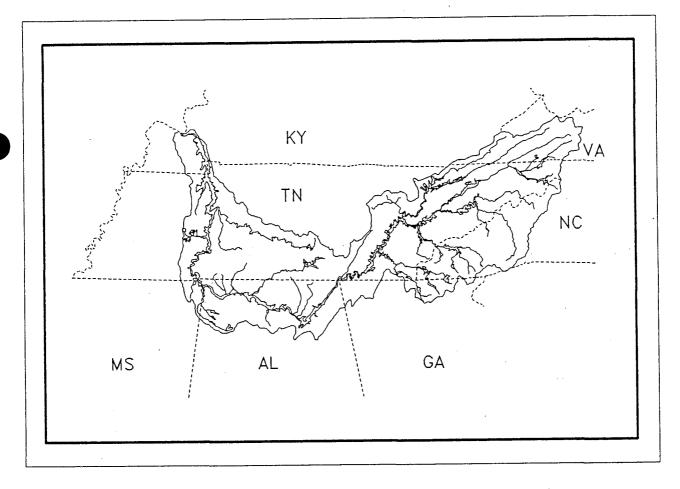


Figure 32. Aquatic macrophyte coverage (hectares) in mainchannel areas downstream and upstream from SQN and total coverage in Chickamauga Reservoir, 1977-92.

Tennessee Valley Authority Water Resources Division Chattanooga, Tennessee TVA/WR/WQ--91/10 May 1991

RESERVOIR VITAL SIGNS MONITORING - 1990 PHYSICAL AND CHEMICAL CHARACTERISTICS OF WATER AND SEDIMENT



WATER RESOURCES & ECOLOGICAL MONITORING

INTRODUCTION

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In FY 1990, the Tennessee Valley Authority (TVA) initiated a Reservoir "Vital Signs" Monitoring program on 12 TVA reservoirs (the nine main stem Tennessee river reservoirs - Kentucky through Fort Loudoun and three major tributary reservoirs - Cherokee, Douglas and Norris) as part of its Water Resources and Ecological Monitoring Program (TVA, 1989). The objective of the Vital Signs program is to provide basic information on the "health" or integrity of the aquatic ecosystem in each TVA reservoir and to provide screening level information for describing how well each reservoir meets the "fishable and swimmable" goals of the Clean Water Act. This is the first time in the history of the Agency that a long term, systematic sampling of all major TVA reservoirs has been conducted. The basis of the Vital Signs monitoring is examination of appropriate physical, chemical and biological indicators in three areas of each reservoir. These three areas are the forebay (the lacustrine region of the reservoir, immediately upstream of the dam), the transition zone (the midreservoir region where the water changes from free flowing to more quiescent, impounded water), and the inflow or headwater region of the reservoir. The information gathered is used to make seasonal and spatial assessments of each reservoir's health and the overall health of the reservoir system, and to implement more detailed short-term studies where problems seem to exist. In addition, this information establishes a baseline for comparing with future water quality conditions and monitoring water quality trends for TVA reservoirs.

The Vital Signs program employs several activities to assess reservoir health. They include physical and chemical characteristics of water and sediment, acute toxicity screening of water and sediment, benthic macroinvertebrate population assessments, and fish abundance, composition and health.

This report presents information on the physical and chemical characteristics of water and sediment for water year

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(WY) 1990. Water samples were collected and water quality measurements made at two locations (forebay and transition zone) on each of twelve TVA reservoirs (the nine main stem Tennessee River Reservoirs and three tributary reservoirs - Norris, Cherokee, and Douglas). Water samples were not collected at inflow locations for two reasons: (1) the ambient stream monitoring program includes inflow locations for Norris, Douglas and Cherokee; (2) "inflow" water samples at the other inflow locations would be dominated by the effects of the impoundment immediately upstream, rather than the process occurring in the reservoir being sampled. Sediment samples were also collected at these same locations. The twenty four Reservoir Vital Signs Monitoring locations sampled in WY 1990 are shown on Figure 1 and listed in Table 1.

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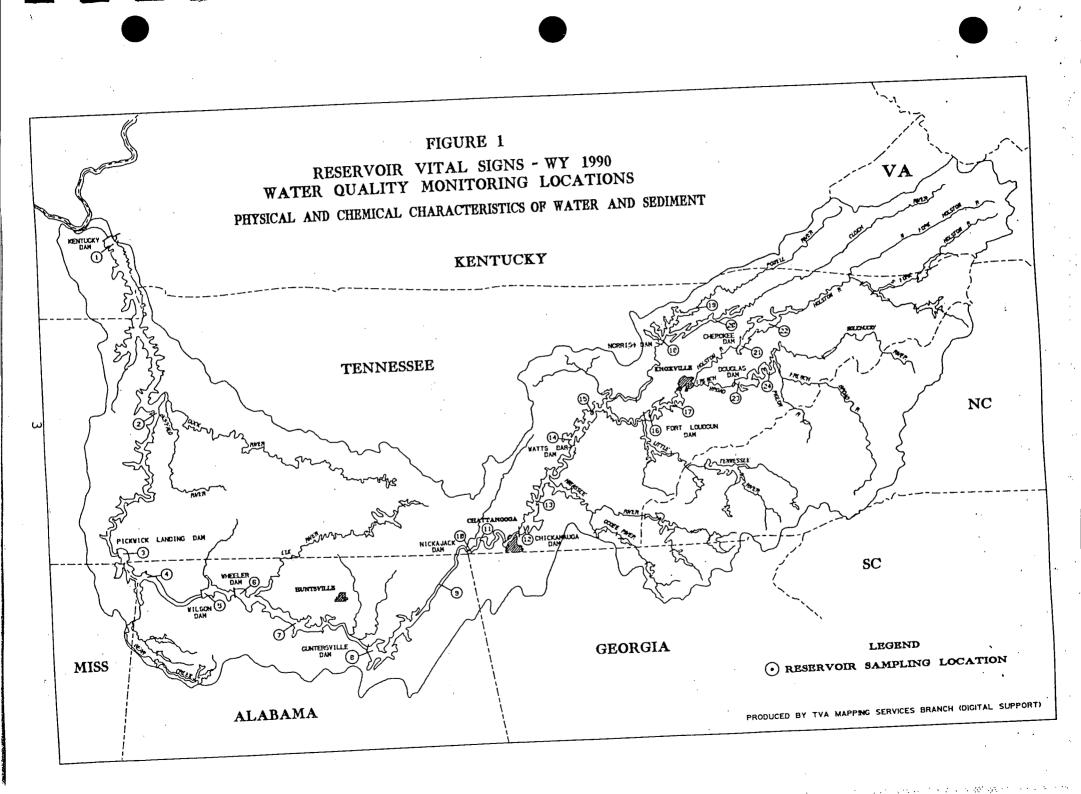


Table 1

	Fore	bay Location	1S	Transition Zone Locations							
		Map ID	Storet		Map ID	Storet					
Reservoir	River Mile	Number	<u>Station #</u>	<u>River Mile</u>	Number	Station #					
	(Tennessee)			(Tennessee)							
		Main	n Stream								
Kentucky	23.0	1.	202832	112.0	2.	475015					
-		2	476799	230.0	4.	016923					
Pickwick	207.3	3.	4/0/33	250.0							
Wilson	260.8	5.	016912								
Wheeler	277.0	6.	016900	307.5	7.	017012					
Guntersville	350.0	8.	017261	396.8	9.	017101					
Nickajack	425.5	10.	476344	433.0	11.	476239					
Chickamauga	472.3	12.	475358	490.5	13.	475265					
Watts Bar	531.0	14.	475317	560.8	15.	476041					
Fort Loudoun	603.2	16.	475602	624.6	17.	475603					

WATER QUALITY MONITORING LOCATIONS RESERVOIR VITAL SIGNS MONITORING, WY 1990

	For	Transition Zone Locations						
Reservoir	River Mile	Map ID Number	Storet Station #	River Mile	Map ID <u>Number</u>	Storet <u>Station #</u>		
		Tr	ibutary					
Cherokee	HRM 53.0	21.	475025	HRM 76.0	22.	475028		
Douglas	FBRM 33.0	23.	475081	FBRM 60.7	24.	475993		
Norris	CRM 80.0	18.	476009	PRM 30.0	19.	477187		
 ,				CRM 125.0	20.	477186		

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WRC 0336M-1

	<u></u>	Transition Zone						
Variable	<u>N</u>	Mean	Min	Max	N	Mean	<u>Min</u>	Max
Temperature (° C)	85	22.1	8.3	28.0	51	20.4	7.2	26.5
Dissolved Oxygen, (mg/l)	85	7.3	3.1	11.4	51	7.4	4.2	11.9
pH (s.u.)	85	.7.5	6.8	8.7	51	7.6	7.1	8.5
Conductivity, (µmhos/cm)	85	163.1	123.0	192.0	51	163.3	129.0	190.0
	18	0.24	0.12	0.48	18	0.23	0.10	0.38
Organic - N (mg/l)	18	0.03	0.01	0.08	18	0.04	0.01	0.11
Ammonia – N (mg/1)	18	0.23	0.13	0.41	18	0.26	0.18	0.43
Nitrate+Nitrite N (mg/l)	18	0.5	0.4	0.7	18	0.5	0.4	0.7
Total Nitrogen (mg/l)	18	0.03	0.02	0.04	18	0.03	0.02	0.04
Total Phosphorous (mg/l) TN/TP Ratio	18	20.1	14.8	27.5	18	19.9	11.0	28.5
	18	0.007	0.003	0.010	18	0.009	0.004	0.020
Dissolved Ortho - P (mg/l) Total Organic Carbon (mg/l)	18	1.9	0.9	2.4	18	2.0	0.8	2.4
-	18	1.7	0.6	2.1	18	1.8	0.7	2.2
Soluble Organic Carbon (mg/l) Chlorophyll-a (µg/l)	14	8.9	2.0	24.0	13	7.0	2.0	17.0
Basabi dopth (m)	7	1.2	0.9	1.6	7	1.2	1.1	1.4
Secchi depth (m)	18	6.2	3.0	16.0	18	7.1	3.0	18.0
Turbidity (NTU)	18	4.9	3.0	8.0	18	5.8	3.0	10.0
Suspended Solids (mg/l)	18	18.2	2.0	30.0	18	17.4	2.0	35.0
True Color (PCU)	18	23.6	5.0	35.0	18	24.4	5.0	43.0
Apparent Color (PCU) Fecal Coliform (#/100 ml)	9.	57.8	10.0	440.0	9	25.6	10.0	150.0

Table 13												
Chickamauga	Reservoir -	Water Quality Summary										
Reservoir	Vital Signs	Monitoring, WY 1990										

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depth in April were at super saturation levels of 116% and 120%, respectively, at the forebay and transition zones. Contour plots of temperature and DO versus depth for the period January through the end of September for both the forebay (figure 23) and the transition zone (figure 24) depict the seasonal variation and rather weak stratification of Chickamauga reservoir in WY1990.

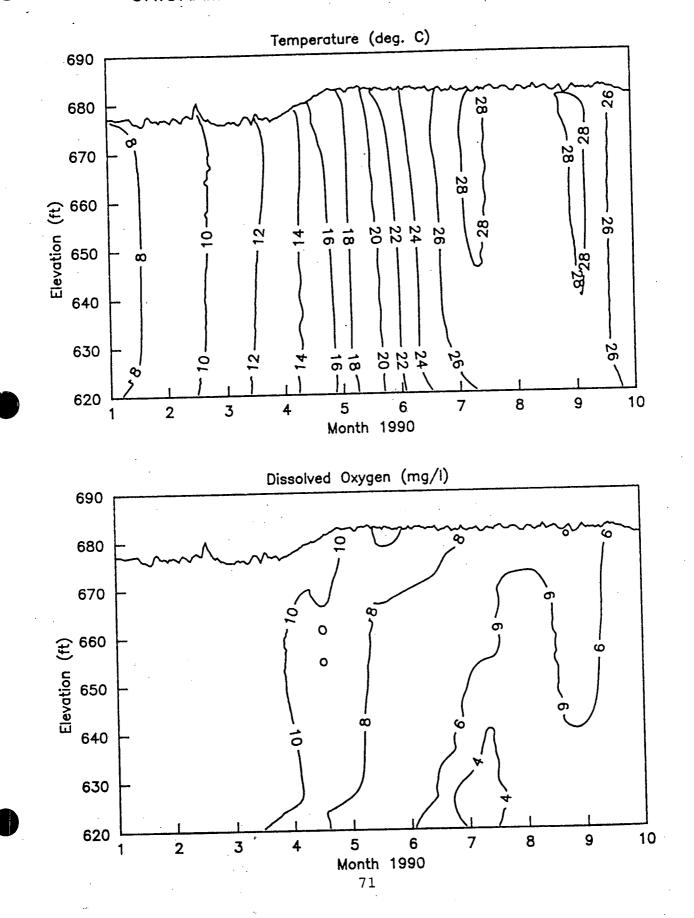
Values of pH ranged from 6.8 to 8.7 on Chickamauga reservoir. Values of pH approaching 8.5 and higher and supersaturation DO levels were observed during algal blooms in April and May at both the forebay and transition zones.

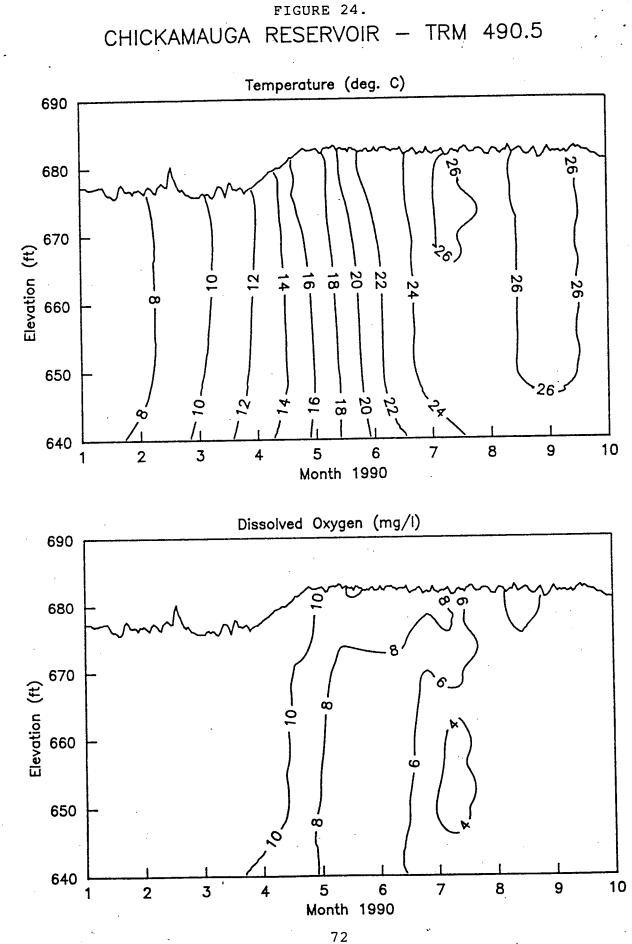
Conductivities ranged from 123 to 192 umhos/cm, with highest conductivities coinciding with low streamflows in July, and averaged about 165 umhos/cm.

<u>Biochemical Measurements</u> - In Chickamauga reservoir, during WY 1990, concentrations of organic nitrogen, ammonia nitrogen, nitrite plus nitrate nitrogen, and total phosphorus averaged 0.24, 0.03, 0.23 and 0.026 mg/l, respectively at the forebay; and 0.22, 0.04, 0.26 and 0.028 mg/l, respectively, at the transition zone. The total phosphorus concentrations measured in Chickamauga reservoir were among the lowest observed at any of the Vital Signs monitoring locations, with the exception of those sampling locations on Norris reservoir.

TN/TP ratios ranged from 11 to 28 in Chickamauga reservoir, indicating periods of phosphorus limiting conditions. The highest chlorophyll-a concentrations were measured in May (12-19 ug/l and 13-17 ug/l, respectively, at the forebay and transition zones) and in June (24 ug/l at the forebay). Values of pH up to 8.7 were measured during this May bloom. Surface concentrations of chlorophyll-a averaged 12 ug/l at the Chickamauga reservoir forebay, and 9.5 ug/l at the transition zone. Organic carbon concentrations (both total and soluble) in Chickamauga reservoir are quite low, averaging 1.9 and 1.7 mg/l, respectively, at the forebay; and averaging 2.0 and 1.8 mg/l, respectively, at the transition location.

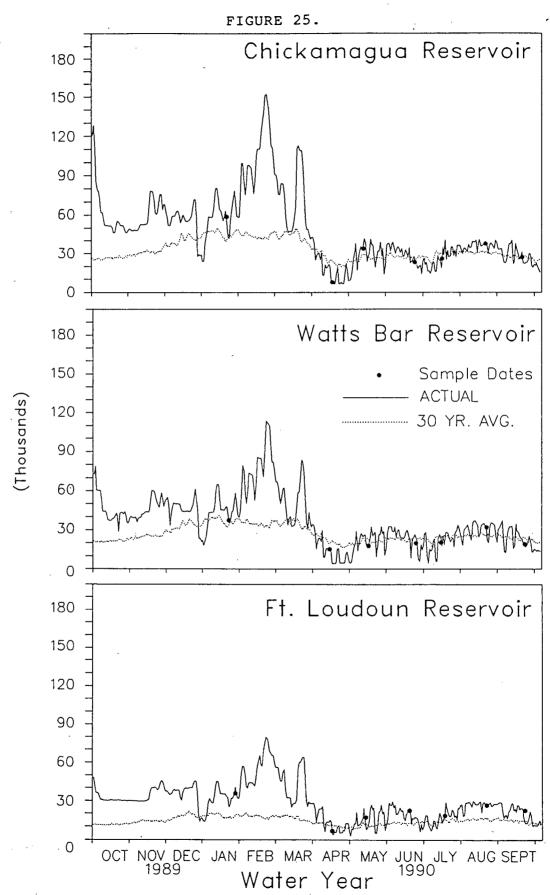
CHICKAMAUGA RESERVOIR - TRM 472.3



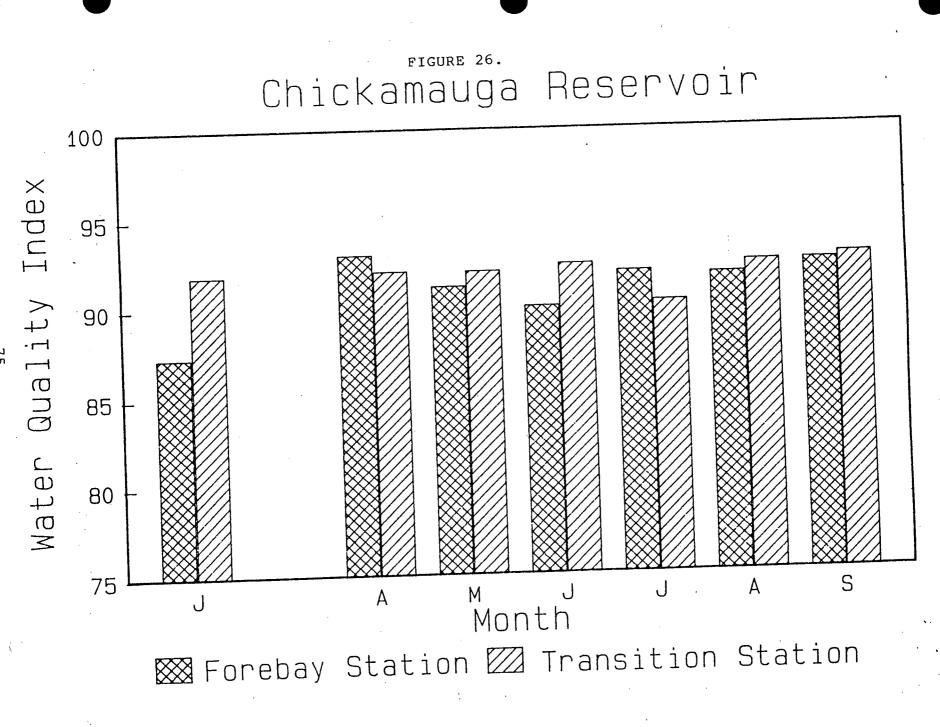


Physical/Bacteriological Measurements - Figure 25 illustrates the average daily discharge from Chickamauga reservoir in WY 1990 with water quality sampling dates superimposed. Periods of high flow in January and February are reflected in the water quality data with low Secchi depth measurements and high concentrations of turbidity and suspended Secchi depth measurements of 0.88 meters, turbidity solids. values in excess of 15 NTU's and suspended solids concentrations of 8 mg/l at the forebay were observed in January. Forebay Secchi depth, turbidity, and suspended solids measurements averaged 1.19 meters, 6.2 NTU's, and 4.9 mg/l, respectively. Transition zone Secchi depths, turbidity, and suspended solids averaged 1.22 meters, 7.1 NTU's, and 5.8 mg/l, respectively. These values indicate the light transparency of Chickamauga reservoir to be high when compared with the other mainstem Tennessee River reservoirs. True color values, averaging 18 and 17 PCU's, at the forebay and transition zones, respectively, were igher than the next upstream monitoring location (Watts Bar forebay), indicating the effects of a colored waste outfall upstream of the transition zone. A large paper company discharges to the Hiwassee River which joins the Tennessee River at mile 499.4, about nine miles upstream of the transition zone monitoring location. Two of eighteen samples collected for fecal coliform organisms were positive; with one of these positive occurrences (440/100 ml at the forebay in January) exceeding a water-contact recreation guideline of 200/100 ml.

Water Quality Indices -The results for the TVA RWQI (Figure 26) show that except for samples collected in January, both the forebay and transition zone evidenced similar water quality index values. In January, a high fecal coliform concentration of 440/100 ml at the forebay resulted in the lower RWQI value when compared with the transition zone. Overall, RWQI values were high, averaging over 91 at the forebay and 92 at the transition zone and indicating very good reservoir water quality.



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<u>Watts Bar Reservoir</u>

In situ Measurements -In situ measurements of temperature, dissolved oxygen (DO), pH and conductivity show the reservoir to be well mixed early in the year but developing a moderate degree of thermal stratification in July and August (Appendix A-8, Table 14). Surface water temperatures ranged from 7.0 °C in January to 28.3 °C in July in the forebay, and from 7.8 °C in January to 26.2 °C in June at the transition zone. Temperatures at the transition zone are influenced by the inflow of cool water from the Clinch River which joins the Tennessee River about seven miles upstream of the transition zone at river mile 567.7. In July, a warming in surface temperature of almost 3 °C was noted between the transition zone (25.4 °C) and the forebay (28.3 °C).

Values for DO at the 1.5 meter depth ranged from 11.5 mg/l in June to 6.1 mg/l in September at the forebay, and from 11.5 mg/l to 6.2 mg/l for the same months at the transition zone. Contour plots of temperature and DO versus depth for the period January through the end of September for both the forebay (figure 27) and the transition zone (figure 28) depict the seasonal variation with a weak summer time thermal stratification and a rather strong oxycline in the forebay of Watts Bar reservoir. In late June, at the forebay, the data show a 6 °C decrease in temperature and nearly an 11 mg/l decrease in DO from the surface to the bottom of the reservoir. This stratified condition persisted through mid-August. In June and July DO concentrations were less than 1 mg/l in the hypolimnion of Watts Bar forebay.

Values of pH ranged form 6.6 to 9.2 on Watts Bar reservoir. In late June, values of pH exceeding 9.0 and DO saturation values approaching 150% were measured, giving evidence of a large algal bloom which existed at both the forebay and transition zone locations. In addition, at the forebay, pH's greater than 8.5 and supersaturation DO values were also observed in May, July and August.

•		Fo	rebay			Transition Zone					
Variable	<u>N</u>	Mean	Min	Max	·	<u></u>	Mean	Min	Max		
- (1.0)	112	20.8	7.0	28.3		55	21.2	7.8	26.2		
Temperature (° C)	112	7.0	0.2	11.5		55	8.2	5.8	12.0		
Dissolved Oxygen, (mg/l)	112	7.8	6.8	9.2		55	7.8	7.3	9.1		
pH (s.u.) Conductivity, (µmhos/cm)	112	170.6	114.0	208.0		55	174.7	126.0	209.0		
$\Omega_{\rm max} = N_{\rm max} (m_{\rm max} / 1)$	18	0.28	0.15	0.50		17	0.24	0.11	0.52		
Organic - N (mg/l)	18	0.04	0.01	0.09		17	0.03	0.01	0.07		
Ammonia – N $(mg/1)$	18	0.23	0.01	0.47		17	0.27	0.19	0.52		
Nitrate+Nitrite - N (mg/1)	18	0.6	0.4	0.8		17	0.5	0.4	0.8		
Total Nitrogen (mg/l)	18	0.03	0.01	0.05		17	0.03	0.02	0.04		
Total Phosphorous (mg/1)	18	22.9	9.0	51.0		17	19.1	14.0	31.5		
TN/TP Ratio	18	0.007	0.002	0.010		17	0.006	0.003	0.020		
Dissolved Ortho - P (mg/1)	18	1.8	0.9	2.7		17	1.8	1.0	2.4		
Total Organic Carbon (mg/1)	18	1.7	0.7	2.5		17	1.6	0.9	2.2		
Soluble Organic Carbon (mg/l) Chlorophyll-a (µg/l)	14	9.6	2.0	20.0		13	9.5	5.0	14.0		
Secchi depth (m)	5	1.4	1.3	1.8		7	1.1	0.8	1.3		
-	17	6.5	2.0	11.0		17	9.3	3.0	20.0		
Turbidity (NTU)	17	5.7	1.0	18.0		17	8.6	4.0	21.0		
Suspended Solids (mg/l)	18	15.5	5.0	30.0		17	14.5	5.0	25.0		
True Color (PCU)	18	19.6	10.0	33.0		17	18.7	7.0	30.0		
Apparent Color (PCU) Fecal Coliform (#/100 ml)	7	11.4	10.0	20.0		7	18.6	10.0	70.0		

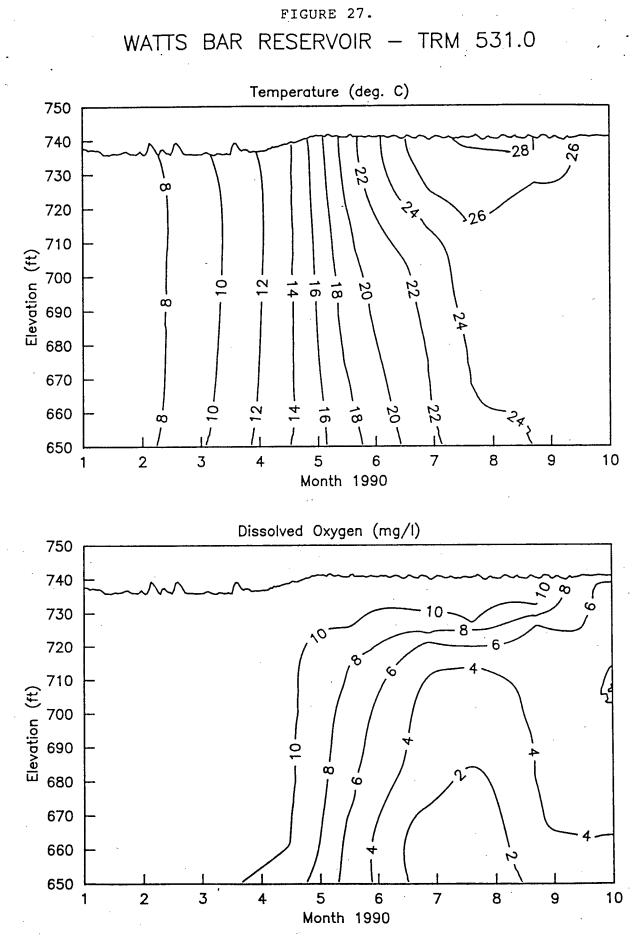
Watts Ba	Table 14 ar Reservoir - Water Quality Summar
Reservo	oir Vital Signs Monitoring, WY 1990

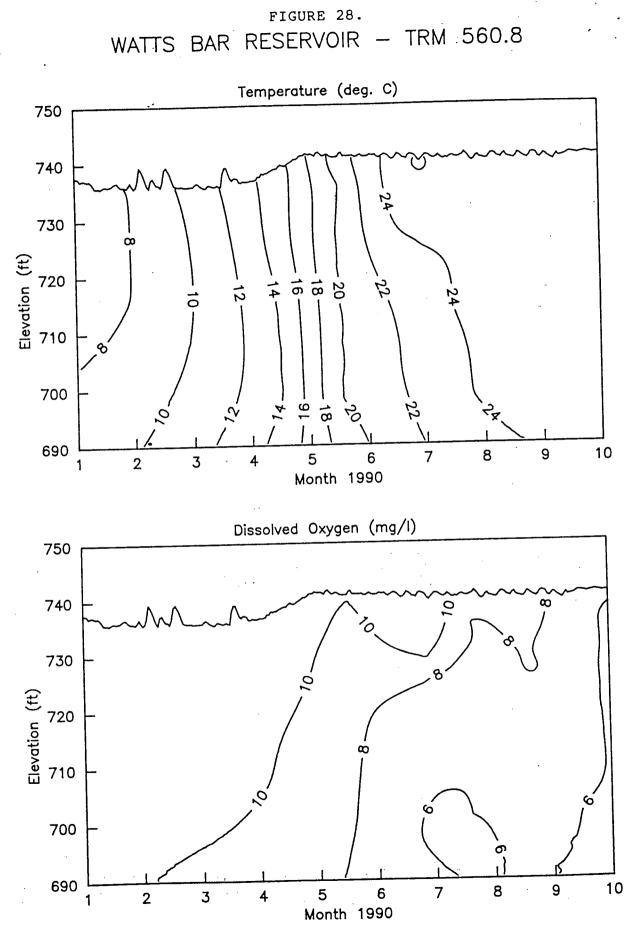
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Conductivities ranged from 114 to 209 umhos/cm and averaged about 175 umhos/cm, with highest conductivities coinciding with low streamflows in June and July.

<u>Biochemical Measurements</u> - In Watts Bar reservoir during WY 1990, concentrations of organic nitrogen, ammonia nitrogen, nitrite plus nitrate nitrogen, and total phosphorus averaged 0.28, 0.04, 0.23 and 0.028 mg/l, respectively at the forebay; and 0.24, 0.03, 0.27 and 0.029 mg/l, respectively, at the transition zone. The total phosphorus concentrations observed at the forebay and transition zones on Watts Bar reservoir were quite low. The dissolved orthophosphorus concentrations of 0.007 and 0.006 mg/l, respectively, at the forebay and transition zones were the lowest observed at any of the twenty four Vital Signs monitoring locations.

TN/TP ratios ranged from 9 to 51 in Watts Bar reservoir, indicating periods of phosphorus limiting conditions, particularly at Watts Bar forebay. The highest chlorophyll-a concentrations were measured in May, 20 ug/l and 14 ug/l, respectively, at the forebay and transition zones. Surface concentrations of chlorophyll-a pigment averaged about 13 ug/l at the Watts Bar reservoir forebay, and about 11 ug/l at the transition zone. Organic carbon concentrations (both total and soluble) in Watts Bar reservoir were low, averaging 1.8 and 1.6 mg/l, respectively, at both the forebay and transition zones. These organic carbon concentrations were the lowest measured at any of the mainstem Tennessee river Vital Signs monitoring locations.

Physical/Bacteriological Measurements - Figure 25 illustrates the average daily discharge from Watts Bar reservoir in WY 1990 with sampling dates superimposed. Forebay Secchi depth, turbidity, and suspended solids measurements averaged 1.44 meters, 6.5 NTU's, and 5.7 mg/l, respectively. Transition zone Secchi depths, turbidity, and suspended solids averaged 1.05 meters, 9.3 NTU's, and 8.6 mg/l, respectively. These values indicate the light transparency of Watts Bar reservoir forebay to

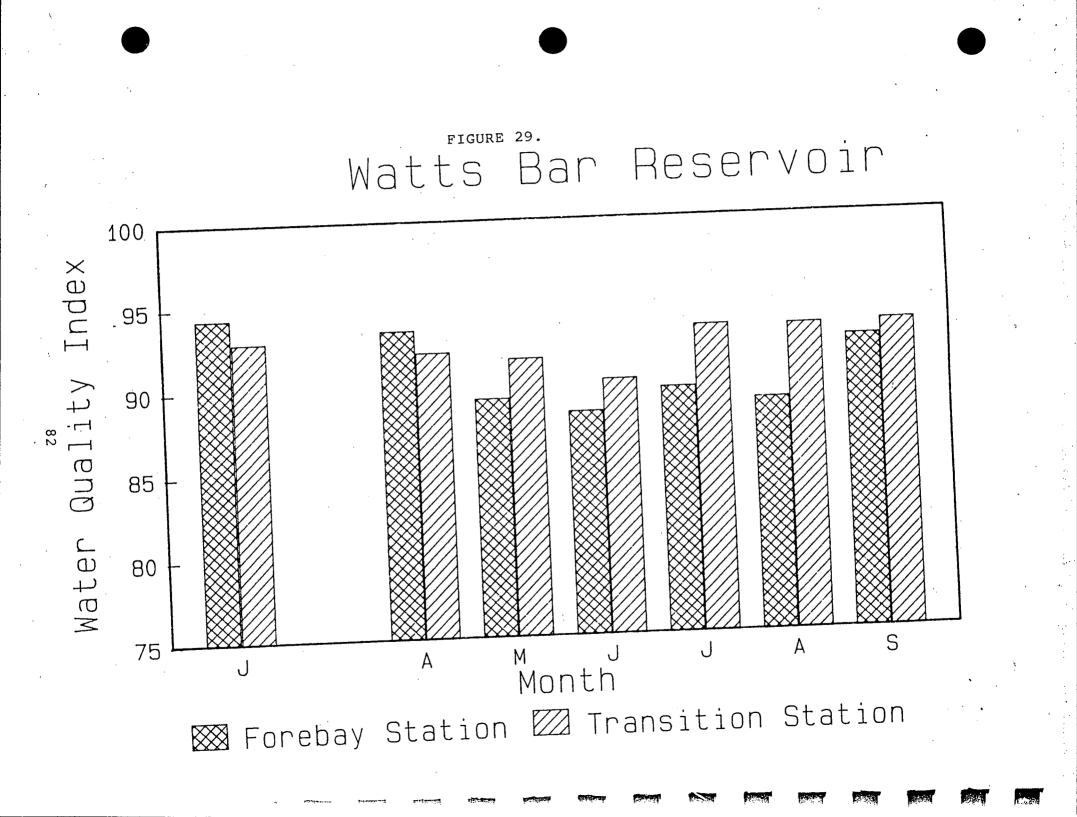
be among the highest of the mainstem Tennessee River reservoirs, in WY 1990. True color values, averaged 16 and 15 PCU's, at the forebay and transition zones, respectively, falling about in the mid-range of values measured at mainstem Tennessee river Vital Signs monitoring locations. Two of fourteen samples collected for fecal coliform organisms were positive; however, neither of these positive occurrences exceeded a water-contact recreation guideline of 200/100 ml.

Water Quality Indices -The results for the TVA RWQI (Figure 29) show that at the forebay, the very high surface pH's, ranging from 8.8 to 9.2, and coincident high chlorophyll-a concentrations during the May through August period, had the greatest effect in lowering the computed RWQI values. Overall, RWQI values were high, averaging 91 at the forebay and over 92 at the transition zone and indicating very good reservoir water quality.

Fort Loudoun Reservoir

In situ Measurements -In situ measurements of temperature, dissolved oxygen (DO), pH and conductivity show the reservoir is well mixed early in the year but develops a fairly strong thermal stratification from July through September (Appendix A-9, Table 15). Surface water temperatures ranged from 7.3 °C in January to 28.8 °C in July in the forebay, and from 7.0 °C to 27.4 °C for the same months at the transition zone. In June, a warming in surface temperature of almost 3.5 °C was noted between the transition zone (23.4 °C) and the forebay (26.8 °C).

Values for DO at the 1.5 meter depth ranged from 12.7 mg/l in July to 6.0 mg/l in September at the forebay, and from 11.6 mg/l in April to 5.7 mg/l in September at the transition zone. Contour plots of temperature and DO versus depth for the period January through the end of September for both the forebay (figure 30) and the transition zone (figure 31) depict the seasonal variation and summer time stratification of Fort Loudoun reservoir. In July, at the forebay, the temperature decreased



		For	rebay	······		Transition Zone					
Variable	N	Mean	Min	Max	•	<u>N</u>	Mean	Min	Max		
Temperature (° C) Dissolved Oxygen, (mg/l) pH (s.u.) Conductivity, (µmhos/cm)	100 100 100 100	19.5 7.1 7.6 181.7	6.8 0.5 6.6 70.0	28.8 12.8 9.0 230.0		75 75 75 75	19.7 7.7 7.6 199.2 0.32	6.9 4.2 6.9 150.0 0.11	27.4 12.6 8.7 240.0 0.66		
Organic - N (mg/l) Ammonia - N (mg/l) Nitrate+Nitrite - N (mg/l) Total Nitrogen (mg/l) Total Phosphorous (mg/l) TN/TP Ratio Dissolved Ortho - P (mg/l) Total Organic Carbon (mg/l) Soluble Organic Carbon (mg/l) Chlorophyll-a (µg/l)	18 18 18 18 18 18 18 18 18 18 9	0.27 0.05 0.37 0.7 0.04 19.1 0.015 2.1 1.9 11.9	0.09 0.01 0.05 0.3 0.02 6.1 0.002 0.9 0.8 1.0	0.46 0.19 0.70 1.1 0.09 46.0 0.030 3.0 2.7 20.0		18 18 18 18 18 18 18 18 18 18 18	$\begin{array}{c} 0.32\\ 0.05\\ 0.50\\ 0.9\\ 0.05\\ 16.7\\ 0.019\\ 2.3\\ 2.1\\ 11.5\end{array}$	0.01 0.22 0.4 0.03 8.4 0.009 1.1 0.9 3.0	0.09 0.65 1.2 0.08 21.3 0.040 3.9 3.9 20.0		
Secchi depth (m) Turbidity (NTU) Suspended Solids (mg/l) True Color (PCU) Apparent Color (PCU) Fecal Coliform (#/100 ml)	7 18 18 18 18 8	1.2 10.9 10.4 18.1 23.5 25.0	0.5 2.0 3.0 5.0 7.0 10.0	2.0 25.0 42.0 35.0 43.0 130.0		7 18 18 18 18 8	0.8 16.8 16.1 21.1 27.0 11.3	0.3 4.0 6.0 5.0 10.0 10.0	1.0 46.0 34.0 45.0 50.0 20.0		

	Table 15
Fort Loudoun	Reservoir - Water Quality Summary
Reservoir	Vital Signs Monitoring, WY 1990

STORET RETRIEVAL DATE 91/03/06 VITAL SIGNS RESERVOIR MONITORING HY 90

/TYPA/AMBNT/STREAM/SOLIDS

1017 475358 35 06 26.0 085 12 20.0 2 CHICKAMAUGA RES. AT LIGHTED BUOY 47065 TENNESSEE HAMILTON HAMILTON 040801 TENNESSEE RIVER BASIN TENNESSEE RIVER 472.3 06020001021 0000.710 ON 131TVAC 0000 METERS DEPTH

191.0 190.0 191.0 188.0 175.0 176.0 176.0 175.0 175.0 175.0 175.0 174.0 174.0 174.0 181.0 181.0 181.0 181.0 182.0 182.0 182.0 182.0 182.0 182.0 183.0 183.0 183.0 183.0 183.0 182.0

DATE FROM TO	TIME OF DAY	MEDIUM	SMK OR DEPTH (M)	00078 TRANSP SECCHI METERS	00010 WATER TEMP CENT	00300 Do Mg/L	00400 PH SU	00094 CNDUCTVY FIELD MICROMHO	DATE FROM TO	TIME OF DAY	MEDIUM	SMK OR DEPTH (M)	00078 TRANSP SECCHI METERS	00010 HATER TEMP CENT	00300 DO HG/L	00400 Ph SU	00094 CNDUCTVY FIELD MICROMHO
10	DAT	rector	(11)	11212100								15		26.2	5.4	7.00	171.0
90/01/24	1119	HATER	0.3	.88	8.3	11.1	7.70	148.0	90/06/26			15.5		26.2	4.7	7.00	172.0
90/01/24			1.5		8.3	10.9	7.70	146.0	90/06/26 90/06/26			16		26.0	4.1	6.90	171.0
90/01/24	1125	HATER	4		8.3	10.8	7.70	149.0	90/06/26			17		25.7	3.1	6.80	171.0
90/01/24	1128	HATER	6		8.3	10.9	7.70	149.0	90/07/17			0.3	1.25	27.2	7.2	7.60	191.0
90/01/24	1131	MATER	8		8.3	10.8	7.70	151.0	90/07/17			1.5		27.1	7.1	7.60	191.0
90/01/24	1134	HATER	10		8.3	10.8	7.70	151.0	90/07/17			2.5		26.9	6.2	7.50	191.0
90/01/24	1137	MATER	12		8.3	10.8	7.70	151.0	90/07/17			4		26.8	5.9	7.50	192.0
90/01/24	1140	HATER	14		8.3	10.8	7.70	150.0 151.0	90/07/17			6		26.8	5.8	7.50	192.0
90/01/24	1143	HATER	16		8.3	10.8	7.70		90/07/17			8		26.8	5.9	7.50	191.0
90/04/17	1100	HATER	0.3	1.00	16.8	11.4	8.60	128.0	90/07/17			10		26.8	5.9	7.50	191.0
90/04/17	1101	MATER	1.5		16.4	11.3	8.50	128.0	90/07/17			12		26.8	5.7	7.50	191.0
90/04/17	1102	MATER	3		15.6	10.3	8.00	128.0 127.0	90/07/17			14		26.7	4.9	7.40	190.0
90/04/17	1104	WATER	3.5		15.2	10.2	7,90	127.0	90/07/17			16		26.5	4.7	7.40	
90/04/17	1106	HATER	4		15.2	10.2	7.90		90/07/17			18		26.5	4.0	7.30	
90/04/17	1110	NATER	6		15.0	10.1	7.90	125.0	90/08/23			0.3	1.36	28.0	8.2	7.90	
90/04/17	1114	MATER	8		14.9	10.1	7.90		90/08/21			1.5		27.9	7.5	7.70	
90/04/17	1118	HATER	10		14.5	10.0	7.80		90/08/2			4		27.7	7.2	7.50	
90/04/17	1120	HATER	12		14.4	9.9	7.80		90/08/2			6		27.7	7.0	7.50	
90/04/17	1122	HATER	14		14.1	9.4	7.70		90/08/2		-	8		27.7	6.9	7.50	
90/04/17	1124	MATER	16		14.1	8.9	7.60		90/08/2			10		27.6	6.6	7.40	
90/04/17	1127	MATER	17.5		14.0	7.9	7.40		90/08/2			12		27.6	6.2	7.30	
90/05/15	1058	HATER	0.3	. 92	21.8	11.0	8.70		90/08/2			14		27.5	5.7	7.20	
90/05/15	1100	HATER	1		20.6	10.2	8.50		90/08/2			16		27.5	5.5	7.10	
90/05/15	1100	NATER	1.5		20.3	9.2	. 8.00		90/08/2			17.5		27.5	5.3	7.10	
90/05/19	1102	HATER	4		19.8	8.3	7.70		90/09/1			0.3	1.60	26.1	5.7	7.50	
90/05/19	1108	HATER	6		19.6	8.0	7.50		90/09/1			1		26.1	5.7	7.40	
90/05/1	1110	MATER	8		19.5	7.8	7.40		90/09/1			1.5		26.0	5.5	7.40	
90/05/1	1115	MATER	10		19.2	7.6	7.40		90/09/1			2		26.0	5.4	7.40	
90/05/1	5 1120	HATER	12		19.1	7.3	7.30		90/09/1			3		25.9	5.4	7.40	
90/05/1	5 1125	WATER	14		19.0	7.2	7.30		90/09/1			4		25.9	5.3	7.40	
90/05/19	5 1130	HATER	16		18.9	7.0	7.20		90/09/1			5		25.9	5.2	7.40	
90/05/1	; 1135	HATER	17.5		18.9	6.9	7.20		90/09/1			6		25.8	5.2	7.30	
90/06/26	1028	WATER	0.3	1.30	26.5	8.4	7.70		90/09/1			7		25.8	5.2	7.30	
90/06/20	1030	HATER	1		26.6	8.1	7.60				5 HATER	8		25.8	5.2	7.30	
90/06/20	1031	WATER	1.5		26.6	7.8	7.60				6 HATER	9		25.8	5.2	7.3	
90/06/20			4		26.6	7.6	7.50		90/09/1			10		25.8	5.2	7.3	
90/06/20			6		26.6	7.3	7.40		90/09/1			11		25.8	5.2	7.3	
90/06/2	5 1036	HATER	8	· ·	26.6	6.7	7.30				7 HATER	12		25.8	5.2	7.3	
90/06/2	5 1037	HATER	10	-	26.6	6.5	7.20					12		25.8 25.8	5.2	- 7:3	b 183.0 182.0
90/06/2 90/06/2	{ 1038	HATEB	12		26.6	5:3	7:10	8 173:8	90/09/1	8 123	8 HATER 8 HATER	14		<i>c</i> 5.0	2.2		
70/06/2	5, IU40	WILL'	47											•			

A26

STORET RETRIEVAL DATE 91/03/06 VITAL SIGNS RESERVOIR MONITORING WY90

 475358
 1017

 35 06 26.0 085 12 20.0 2
 CHICKAMAUGA RES. AT LIGHTED BUOY

 47065
 TENNESSEE

 HAMILTON
 040801

 TENNESSEE RIVER BASIN
 040801

 TENNESSEE RIVER 472.3
 06020001021 0000.710 ON

 0000 METERS DEPTH
 06020001021 0000.710 ON

/TYPA/AMBNT/STREAM/SOLIDS

DATE FROM TO	TIME OF DAY	MEDIUM	SMK OR DEPTH (M)	00078 TRANSP SECCHI METERS	00010 WATER TEMP CENT	00300 Do Mg/L	00400 Ph SU	00094 CNDUCTVY FIELD MICROMHO
90/09/18 90/09/18 90/09/18	1240	WATER	15 16 17		25.8 25.8 25.7	5.4 5.4 5.4	7.40 7.40 7.40	182.0 184.0 183.0

PGM=RET

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PGM≖RET

STORET RETRIEVAL DATE 91/03/06 VITAL SIGNS RESERVOIR MONITORING HY90

ONITORING

 475356
 1017

 35 06 26.0 085 12 20.0 2
 CHICKAHAUGA RES. AT LIGHTED BUOY

 47065
 TENNESSEE

 HAMILTON
 47065

 TENNESSEE RIVER BASIN
 040801

 TENNESSE RIVER 472.3
 131TVAC

 0602000
 0000 METERS DEPTH

06020001021 0000.710 ON

TYPA/AMENT/STREAM/SOLIDS

DATE	TIME		SHK Or Depth	84002 Code General	00080 COLOR PT-CO	00081 AP COLOR PT-CO	82079 TURBIDTY LAB NTU	00530 RESIDUE TOT NFLT HG/L	00605 Org N N MG/L	00610 NH3+NH4- N TOTAL HG/L	00630 No21N03 N-TOTAL Hg/L	00665 PHOS-TOT HG/L P	00671 Phos-dis Ortho Mg/l P
FROM	OF DAY	MEDIUM	(8)	REMARKS	UNITS	UNITS	NIU			.050	.41	.040	.010
TO	UAT	11202011			15	25	16.0	8	.z10	.050		.030	.009
90/01/24	1125	VERT	4		10	15	11.0	6	.190 .160	.010		.020	.005
on/n1/24	1138	MATER	13.5		10	15	5.8	5.5	.140	.020		.020	.000
on/04/17	1106	VERT	4		10	20	7.9	Ð					
00/04/17	1123	HATER	14.5	01									
90/05/1	5 1058	MAIER	0.3	DZ			•				, .1	x .020	.003
90/05/11 90/05/11	5 1059	MATER	0.3	D3		28	3.0	3	.370		· .	5.030	
90/05/1	5 1100	VERT	4	D1	25	35		4	.410	`	· · ·	3.040	
90/05/1	5 1103	VERT	4	D2	30	32	5.0		.450			6 .030	
00/05/1	5 1109	VERI	4	D3 D1	30	33	5 8.0		.130		0.2		
90/05/1	5 1126	MATER	14.5	02	25	33		· ·	.160	, .06			
90/05/1	5 1127	MATER	14.5 14.5	03	25	33		·	.480	, .01		0 .030 3 .020	
90/05/1	5 1124	S MATER	14.5		20	21		·	.250		•	.02	.003
90/06/2	6 103	Z VERI	14.5		25	2	• <u> </u>		.18			.02	0.007
90/06/2	0 104	VERT	4		20 20		-	0 4	.29			16 .02	
90/07/	7 111	Z MATER	15		10	-		0 3	.28			19 .02	
90/08/2	21 111	9 VERI	4		10		5 S.				• 0	19 .02	1 A
90/08/3	21 112	7 HATER	14.6		10	1	5 3.	• <u>;</u>			30 ·	18 .02	
00/09/	18 123	2 VERT	4 14.4		2		5 4.	0 0			4 3223	8	31616
90/09/	18 123	9 HATER	1414			00683	1	32211	32212		•		FEC COLI
			SHK	84002	00680 T ORG C	DORG		CHLRPHYL	CHERPHY	C CREAT	A		HFH-FCBR
DATE	TI	Æ	OR	CODE	C	C	-	A UG/L	B DUG/L	UG/L	UG/1	-	/100ML
FROM			DEPTH	GENERAL REMARKS	HG/L	HG/L		CORRECTI	J 00/C				440
TO	DAY	r MEDIUR	(H)	REPARTS								•	1.0
		-	0.3				,		5	1	1	1 2	
90/01/	24 11	19 HATER 25 VERT	4			•	.6 .7		4	1	1	•	10K
90/01/	24 11	38 HATER	13.5		•	9	••		_	1	2	5	
90/04	17 11	00 HATER	0.3		1.	a 1	.7		5	1K	īк	1	10K
90/04/	/17 11	06 VERT	4		î.		.5		4	10			10K
90/04	/17 11	23 MATER	14.5										10K
90/05	/15 10	58 HATER	0.3								_	1K	
90/05	/15 10	59 HATER	0.3		· .			1	12	2	2	16	
90/05	/15 11	00 HATER		. 01	2.		2.0 1.9		16	1 .	3 5	6	
90/05	/15 11	02 VERT	4				2.0	:	19	4	2	4	•
90/05	/15 11	04 VERT		4 03			1.9		2 2	1 2	2	3	
90/05	/15 1]	126 HATER	14.			.0	2.1		2	ĩĸ	1K	1	10K
90/05	/15 1	127 WATER	14. 14.			.0	1.9		-		_	•	
90/05	:/15 1)	128 MATER	0.						24	2	3	5	
90/06	/26 1	028 WATER		4			2.1 1.9						10K
90/06	/26 1	032 VERT	14.	5	2	.0	1.7				1	z	
90/00	7/17 1	041 HATER 059 HATER	о.	3		2.1	1.9		11	1K	•		10K
90/0	7/17 1	104 VERT		4			1.7						TOV
90/0	7/17 1	112 MAILK		.5	•				13	1	1	2	
90/0	8/21 1	116 MATER		4	1	2.2	2.1			-			10K
90/0	8/21]	119 VERI			1	1.9	1.9					1	
90/0	8/21 1	127 HATER					1.8		5	1K	1K	*	
00/0	9/18]	228 HATER	•		:	2.0							
,0,0	0/10	1232 VERT		4 ·		1.9	1.8						

STORET RETRIEVAL DATE 91/03/06 VITAL SIGNS RESERVOIR MONITORING MY90

PGM=RET

475265 1053 35 18 00.0 085 04 33.0 2 CHICKAMAUGA RESERVOIR 47065 TENNESSEE 1 TENNESSEE RIVER BASIN 4 TENNESSEE RIVER 490.47 131TVAC 0000 METERS DEPTH

HAMILTON 040801 06020001025 0005.740 ON

/TYPA/AMBNT/STREAM/SOLIDS

A29

DATE	TIME OF		SMK OR DEPTH	00078 TRANSP SECCHI	00010 Mater Temp	00300 DO	00400 Ph	00094 CNDUCTVY FIELD	DATE FROM	TIME OF		SHK OR DEPTH	00078 TRANSP SECCHI HETERS	00010 HATER TEMP CENT	00300 DO HG/L	00400 PH SU	00094 CNDUCTVY FIELD MICROMHO
FROM TO	DAY	HEDIUH	(H)	HETERS	CENT	HG/L	SU	MICROMHO	то	DAY	MEDIUM	(H)	rie i eko	CLIII			
90/01/24			0.3	1.38	7.3	11.2	7.80	158.0	90/08/21 90/08/21			4 [`] 6		26.2 26.2	5.5 5.4	7.30 7.30	179.0 179.0
90/01/24			1.5		7.3	11.1	7.70	158.0 160.0	90/08/21			8		26.2	5.3	7.20	179.0 179.0
90/01/24	0937	MATER	4	•	7.2	11.1	7.70 7.80	162.0	90/08/21	1043	HATER	10		26.2	5.3	7.20 7.40	187.0
90/01/24			6		7.2 7.2	$11.1 \\ 11.1$	7.80	162.0	90/09/18	1001	HATER	0.3	1.25	25.0	5.5 5.4	7.40	187.0
90/01/24			8		7.2	11.1	7.80	161.0	90/09/18	1003	HATER	1.5		25.1 25.1	5.3	7.40	186.0
90/01/24			9 0.3	1.10	15.8	11.9	8.50	134.0	90/09/18			4		25.1	5.3	7.40	186.0
90/04/17 90/04/17			1.5	1.10	15.8	11.8	8.50	134.0	90/09/18			6		25.1	5.2	7.40	187.0
90/04/17			2.5		15.3	10.9	8.10	134.0	90/09/18			.8 10		25.1	5.2	7.30	187.0
90/04/17			3		14.8	10.0	7.70	136.0	90/09/18	3 1009	MAIER	10					
90/04/17			4		14.7	9.7	7.70	136.0									
90/04/17			6		14.3	9.4	7.60	134.0									
90/04/17			8		14.2	9.0	7.60	133.0 134.0									
90/04/17			10	_	14.1	8.7	7.50 8.40	134.0									
90/05/1			0.3	1.07	21.0	10.2	8.20	136.0									
90/05/1			1		20.8	9.4 8.9	7.90	135.0									•
90/05/1			1.5		20.4 20.0	8.5	7.70	136.0									
90/05/1			2		19.2	7.5	7.40	134.0									
90/05/1			4		18.8	7.4	7.40	133.0									
90/05/1			8		18.6	7.2	7.30	131.0									
90/05/19 90/05/19			10		18.4	7.0	7.30	130.0									
90/05/1			10.5		18.4	7.0	7.30	129.0							1		
90/06/2			0,3	1.25	24.6	8.9	7.90	175.0	•								3
90/06/2			1		24.5	8.0	7.80	176.0									
90/06/2			1.5		24.7	7.8	7.80	175.0									
90/06/2			2.5		24.6	7.1	7.50	176.0 175.0									
90/06/2			3		24.5	6.0	7.30 7.20										
90/06/2	6 0931	MATER	4		24.4	5.7 5.3	7.20										
90/06/2			6		24.3	5.2	7.20										
90/06/2			8		24.3 24.2	4.4	7.10										
90/06/2			10	1.25	24.2	4.9	7.50										
90/07/1			0.3	1.25	25.1	4.7	7.50										
90/07/1			1.5 4		25.1	4.5	7.50										
90/07/1			6		25.0	4.5	7.50										
90/07/1			8		25.0	4.4	7.50										
90/07/1 90/07/1			10		24.9	4.2	7.50										
90/07/1			10.5		24.9	4.2	7.50	190.0									
90/08/2	i 103 i 103	NATER	0.3	1.24	26.5	<u>6:1</u> 5:7	7:48	178:8									

PGH=RET

STORET RETRIEVAL DATE 91/03/06 VITAL SIGNS RESERVOIR MONITORING MY90 475265 1053 35 18 00.0 085 04 33.0 2 CHICKAMAUGA RESERVOIR 47065 TENNESSEE HAMILTON TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 490.47 131TVAC 06020001025 0005.740 ON 0000 METERS DEPTH

00(7)

/TYPA/AHBNT/STREAH/SOLIDS

1 - 4-4 - -

			SMK OR	84002 CODE	00080 COLOR	00081 AP COLOR	82079 TURBIDTY	00530 RESIDUE	00605 ORG N	00610 NH3+NH4-	00630 NO24NO3 N-TOTAL	00665 PHOS-TOT	00671 PHOS-DIS ORTHO
DATE FROM	TIME OF		DEPTH	GENERAL	PT-CO	PT-CO	LAB	TOT NFLT HG/L	N HG/L	N TOTAL HG/L	HG/L	MG/L P	HG/L P
TO	DAY	HEDIUH	(M)	REMARKS	UNITS	UNITS	NTU	NOV L				.030	.010
			4		10	20	12.0	8	.210	.050	.43 .43	.030	.010
90/01/24	0937	VERT	6.Z		10	20	18.0	10	.150	.050	.24	.020	.005
90/01/24	0991	RAIER	4		10	20	5.Z	4	.200	.010K .040	.30	.030	.010
90/04/17 90/04/17	1002	WATED	9.2000		10	20	10.3	7	.100	.040			
90/04/17	1002	HATER	0.3	01									
90/05/15	0931	HATER	0.3	DZ									
90/05/15	0932	HATER	0.3	D3				3	.330	.010	.19	.030	.006
90/05/15			4	Dl	20	31	4.0	3	.360	.020	.19	.020	.005
90/05/15			4	02	20	27 32	4.0	3	.350	.030	.19	.020	.010
90/05/15			4	D3	25	38	11.0	8	.160	.080	. 26	.030	.010
90/05/15	0955	HATER	9.8	01	30 35	43	10.0	9	.190	.070	. 26		.010
90/05/15	0956	MATER	9.8	D2	30	38	11.0	9	.270	.070	.25	.030	.004
90/05/15			9.8	03	25	30	6.0	5	.380	.010		.030	.005
90/06/26			4		30	35		6	.260	.020	.29		.010
90/06/26	0942	HATER	9.5		20	25		4	.210	.010	.28		
90/07/17	0936	VERT	4		15	20		Б	.250	.010	.28 .23		.010
90/07/17			9.6000		10	15		5	.190	.030	.29		
90/08/21	1036	VERT	4 9.3		10	15	4.0	5	.170	.030	.18		
90/08/21			7.5		ž	5	3.0		.160		.18		
90/09/18					ž	5	; 3.0	4	.110	.110			
90/09/18	3 1008	MATER	9.4		_								31616
				84002	00680	00681		32211	32212	32214	32218		FEC COLI
			SHK OR	CODE	TORGC	D ORG C		CHLRPHYL	CHLRPHYL			(MFM-FCBR
DATE	TIME	•	DEPTH	GENERAL	C	C		A UG/L	B	С	Å		/100HL
FROM	OF	HEDIUH	(M)	REMARKS	HG/L	HG/L		CORRECTO	UG/L	UG/L	UG/L		/ 100110
то	DAY	NEUTON		NET DATE OF									150
90/01/24	4 093	5 HATER	0.3			_	_	5	3	1 2	. :	2	
90/01/2			4		. 9	•		. 4				2	
90/01/24			6.2		.8	•	r	· •		-			10K
90/04/1			0.3		·		0	11	:	IK 1		1	
90/04/1	7 095	VERT	4		1.9			4		່	L	1	
90/04/1			9.2000		1.8		D	•					10K
90/05/1			0.3	D1		-							10 10
90/05/1			0.3	D2								_	10
90/05/1			0.3	D3 D1	2.2	ź.,	0	17				3	
90/05/1			4	D2	2.3			13		•		2	
90/05/1			4	D3	2.4			16		-	-	1K 2	
90/05/1			9.8	01	2.3		0	2		-	-	2 1K	
90/05/1			9.8	DŽ	2.1		9 .	3			-	2	
90/05/1			9.8	03	2.1		0	2	2	IK	1K	۰.	10K
90/05/1 90/06/2	5 075	A WITED	0.3										
90/06/2			4		2.4	2.	2						
			9.5		2.1	٤.	0						10K
90/06/2	7 0074	3 HATER	0.3					_	-		1K	1K	
90/07/1	7 047	A VEPT	4		2.1			5	5	1K	AN.		
		9 HATER	9.6000		1.9		7						10K
		2 HATER	0.3						-	1K	1	1	
90/08/2			4		z.:			· 1	5	11	•		
		1 HATER	9.3		1.	91.	. 9						10K
		1 HATER	0.3			-	-		4	1K	1K ·	1	
90/09/1			4	•	Ź.(•	•				
		8 HATER	9.4	•	2.	0 1.	. 7						

STURET RETRIEVAL DATE 91/03/06 VITAL SIGNS RESERVOIR MONITORING WY90

PGM=RET

475317 1089 35 38 10.0 084 47 06.0 2 OPP. LOWE BR. HATTS BAR RES. 47121 TENNESSEE MEIGS TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 531.0 131TVAC HQ 06010201002 0002.040 OFF 0000 METERS DEPTH

/TYPA/AMBNT/STREAM/SOLIDS

				SHK	00078	00010	00300	00400	00094				SMK	00078	00010	00300	00400	00094	• •
	DATE	TIME		OR	TRANSP	WATER	DO	PH	CNDUCTVY	DATE	TIME		OR	TRANSP	MATER	00	PH	CNDUCTVY	
	FROM	OF		DEPTH	SECCHI	TEMP			FIELD	FROM	OF		DEPTH	SECCHI	TEMP			FIELD	
	TO	DAY	MEDIUM	(H)	METERS	CENT	MG/L	SU	MICROMHO	то	DAY	HEDIUH	(M)	HETERS	CENT	HG/L	SU	MICROMHO	
		-															- 10	114.0	
	90/01/25	1542	HATER	0.3	1.38	7.0	11.4	7.80	155.0	90/05/16			25		17.5	5.3	7.10 9.20	167.0	
	90/01/25			1.5		7.0	11.3	7.90	154.0	90/06/27			0.3	1.75	27.6	11.4	9.20	166.0	
	90/01/25			4		7.0	11.3	7.80	156.0	90/06/27			1.5		26.9	11.5	9.00	168.0	
	90/01/25			6		7.0	11.3	7.80	158.0	90/06/27			2.5		26.5	10.7	8.90	169.0	
	90/01/25			8		7.0	11.2	7.80	158.0	90/06/27			3		26.4	10.0 9.0	8.70	170.0	
	90/01/25			10		7.0	11.2	7.80	158.0	90/06/27			4		26.1	8.2	8.60	171.0	
	90/01/25	1553	HATER	12		7.0	11.2	7.80	157.0	90/06/27			5		25.8	6.7	8.20	174.0	
	90/01/25	1555	MATER	14		7.0	11.2	7.90	157.0	90/06/27			5.5		25.5 25.2	5.9	7.90	177.0	
	90/01/25	1557	HATER	16		7.0	11.2	7.80	159.0	90/06/27			6		29.2	5.2	7.40	181.0	
	90/01/25	1559	HATER	18		7.0	11.2	7.90	159.0	90/06/27			6.5		24.4	4.4	7.20	184.0	
	90/01/25	1601	MATER	20		7.0	11.2	7.90	158.0	90/06/27			8		22.8	3.7	7.00	187.0	
	90/01/25	1603	MATER	22		7.0	11.2	7.90	158.0	90/06/27			10		22.6	3.4	7.00	186.0	
	90/01/25	1605	HATER	24		7.0	11.1	7.90	158.0	90/06/27			12		22.5	3.3	6.90	186.0	
	90/01/25	1607	MATER	25		7.0	11.1	7.90	157.0	90/06/27			14 16		22.3	3.2	6.90		
	90/04/18	1234	MATER	0.3	1.36	14.1	10.5	8.10	124.0	90/06/27			18		22.1	3.2	6.90	185.0	
	90/04/18	1235	MATER	1.5		14.1	10.4	8.10	124.0	90/06/27			20		21.9	2.3	6.90		
•	90/04/18	1236	MATER	- 4		14.1	10.3	8.10	124.0	90/06/27			20		21.9	1.8	6.80		
	90/04/18	1239	MATER	6		14.1	10.2	8.10	123.0	90/06/27			22		21.8	1.1	6.80		
	90/04/18	1240	MATER	8		14.0	10.1	8.10	123.0	90/06/27			24		21.7	.7	6.80		
	90/04/18	1241	HATER	10		14.0	10.1	8.10	123.0	90/06/27			25		21.6	.6	6.80		
	90/04/18			12		14.1	10.1	8.10	123.0	90/06/27 90/07/19			0.3	1.45	28.3	11.0	8.90		
	90/04/18			14		14.0	10.0	8.00	122.0	90/07/19			1.5	2112	27.7	11.0	8.90	185.0	
	90/04/18			16		14.0	10.0	8.00	122.0 124.0	90/07/19			4		27.1	10.1	8.90		
	90/04/18			18		14.0	10.0	8.00	122.0	90/07/19			4.5		27.0	9.4	8.70	188.0	
	90/04/18			20		14.0	9.9	8.00	123.0	90/07/19			5		26.6	6.5	8.40	193.0	
	90/04/18			22		14.0	9.8	8.00 8.00	123.0	90/07/19			6		26.4	6.2	8.20	194.0	
	90/04/18			24		13.9	9.8	8.80	153.0	90/07/19			7.5		26.1	5.3	7.90	196.0	
	90/05/16			0.3		21.1	10.7	8.80	153.0	90/07/19			8		25.9	4.4	7.80	199.0	
	90/05/16			1.5		21.0	10.6 10.3	8.70	154.0	90/07/19			8.5		25.2	3.2	7.50	200.0	
	90/05/16			4		20.6	9.7	8.50	154.0	90/07/19			10		25.0	3.0	7.30	202.0	
	90/05/16			5		20.4 20.2	8.9	8.20	156.0	90/07/19			12		24.9	3.0	7.40	203.0	£
	90/05/16			6 8		19.7	8.0	7.70	156.0	90/07/19			14		24.7	3.0	7.40		
	90/05/16			10		19.5	7.7	7.60	156.0	90/07/19			16		24.4	2.3	7.40		
	90/05/16			10		19.1	7.2	7.50	154.0	90/07/19			18		24.2	1.7	7.30		
	90/05/16			14		18.8	6.9	7.40	149.0	90/07/19	9 1201	L WATER	20		24.1	1.2	7.30		
	90/05/16			16		18.8	6.8	7.40		90/07/19	9 1202	2 HATER	22		24.0	.9	7.30		
	90/05/16			18		18.5	6.4	7.40		90/07/19	9 1202	2 HATER	24		23.8	.2	7.30		
	90/05/16			20		18.3	6.3	7.30	144.0	90/07/19	9 1204	HATER	25.5		23.6	, , · ?	7.30		
	20/05/16			224		17:7	5.2	7:20	146.0 143.0	90/08/2 90/08/2	3 1139	R MATER	9:3 1:5		28.0 28.0	11:1	8.90 8.80	3 178:8	j –
	90/05/16	1250	MATER	Z4		17.7	5.4	1.20	173.0	307 007 21			2.5					•	

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PGM=RET

STORET RETRIEVAL DATE 91/03/06 VITAL SIGNS RESERVOIR MONITORING WY 90

 475317
 1089

 35 38 10.0 084 47 06.0 2

 OPP. LOWE BR. MATTS BAR RES.

 47121
 TENNESSEE

 MEIGS

 TENNESSEE RIVER BASIN
 040801

 TENNESSEE RIVER 531.0

 131TVAC
 HQ 06010201002 0002.040 OFF

 0000 METERS DEPTH

/TYPA/AMBNT/STREAM/SOLIDS

DATE	TIME		SMK Or Depth	00078 TRANSP SECCHI	00010 WATER TEMP	00300 DO	00400 Ph	00094 CNDUCTVY FIELD
FROM	DAY	MEDIUM	(M)	METERS	CENT	MG/L	SU	MICROMHO
10	DAI	(IEUZOII						170 0
90/08/22	1134	WATER	2		27.8	10.4	8.70	178.0 179.0
90/08/22			2.5		27.7	10.0	8.70 8.50	177.0
90/08/22			3		27.6	8.7		183.0
90/08/22			3.5		26.8	8.0	8.10 7.50	185.0
90/08/22			4		26.0	6.2	7.50	185.0
90/08/22			6		25.0	5.0	7.00	184.0
90/08/22			8		24.7	4.7	7.00	183.0
90/08/22			10		24.5	4.5	7.00	184.0
90/08/22			12		24.4	4.5	7.00	183.0
90/08/22	1143	WATER	14		24.3	4.2	7.00	182.0
90/08/22	1144	WATER	16		24.3	4.1		182.0
90/08/22			18		24.2	4.1	6.90	181.0
90/08/22			20		24.2	3.9	6.90 6.90	181.0
90/08/22			22		24.1	3.8		182.0
90/08/22			24		24.1	3.7	6.90	
90/08/22			24.5		24.1	3.5	7.00	192.0
90/09/19			0.3	1.25	25.5	6.2	7.70	194.0
90/09/19			1.5		25.5	6.1	7.70	
90/09/19			4		25.5	5.9	7.60	194.0
90/09/19			6		25.6	6.0	7.70	195.0
90/09/19			8		25.6	5.9	7.70	194.0
90/09/19			10		25.6	5.9	7.60	195.0
90/09/19			12		25.6	5.9	7.70	194.0
90/09/19			14		25.6	5.8	7.70	195.0
90/09/19	1152	WATER	16		25.6	5.7	7.60	194.0
90/09/1			18		25.5	5.3	7.60	195.0
90/09/19			20		25.4	4.8	7.50	195.0
90/09/19			22		25.4	4.8	7.50	
			23.5	:	25.5	4.7	7.50	
90/09/1			24		25.4	2.6	7.50	199.0
90/09/1	9 1155	MAICK	24					

STORET RETRIEVAL DATE 91/03/06 VITAL SIGNS RESERVOIR MONITORING Hy90

/TYPA/AMBNT/STREAM/SOLIDS

475 35 36 10.0 084 47 06.0 2 OPP. LOWE BR. MATTS BAR RES. 47121 TENNESSEE MEIGE TENNESSEE RIVER BASIN 04080 TENNESSEE RIVER 531.0 1317VAC HO C HEIGS 040801 0000 HETERS DEPTH

PGH=RET

HQ 06010201002 0002.040 OFF

			·		00080	00081	82079	00530	00605	00610	00630 NO21NO3	00665 PHOS-TOT	00671 Phos-dis
			SHK	84002	00080	AP COLOR	TURBIDTY	RESIDUE	ORG N	NH3+NH4-	N-TOTAL		ORTHO
DATE	TIME		OR	CODE	COLOR PT-CO	PT-CO	LAB	TOT NFLT	N	N TOTAL HG/L	MG/L	HG/L P	HG/L P
FROM	OF		DEPTH	GENERAL	UNITS	UNITS	UTN	HG/L	HG/L	N 67 L	1.07 4	•	
то	DAY	MEDIUM	(H)	REMARKS	UNITS					.070	.47	.040	.010
					10	15	9.0	5	.190	.050	.47	.030	.010
90/01/25	1544	VERT	4		10	20	8.8	4	.170	.020	.30	.020	.006
90/01/25	1604	HATER	22.3		15	15	5.1	4	.180	.020	.31	.010	.005
90/04/18	1236	VERT	4		15	15	6.0	6	,170	.050			
90/04/18	1246	HATER	21.3										
90/05/16	1213	KATER	0.3	D1									
90/05/16	1214	HATER	0.3	D2						.020	.16	.020	.003
90/05/16	1215	HATER	0.3	D3	20	25	2.0	1K					.003
90/05/16	1217	VERT	4	D1	20	23		1K		.020			.003
90/05/16	1218	VERT	4	D2	25	30		2	.4ZO	.020			.010
90/05/16	1219	VERT	4	D3	30	33			.210	.090			.008
90/05/16	1248	HATER	22.2	D1	30	33			.230				.008
90/05/16	1249	HATER	22.2	D2		33			.270				.002
90/05/16	1250	HATER	22.2	D3	30	15			.500				
90/06/27	1149	VERT	4		10	10			.440		•		
90/06/27	7 1203	HATER	22.3		7	20			.500				
90/07/19	114	VERT	4		15	10			.200				
90/07/1	1 1 2 0 2	WATER	22.7		7				.310	.01			
90/08/2	7 1204 9 1174	VERT	4		10				.150	.01		-	
90/08/2	6 11A	WATER	21.8		10		-		.180	.06		-	
90/08/2	C 3344	VEDT	4		10			18	.210	.07	0.2	1 .00	
90/09/1	9 114	A MATER	21.1		5	1	0 11.0						31616
90/09/1	4 IIS	ANALER						32211	32212	32214	32218		FEC COLI
			SHK	84002	00680	00681		CHLRPHYL			L PHEOPHI	พ	HFH-FCBR
		-	OR	CODE	T ORG C	D ORG C		A UG/L	B	- c	· A		/100HL
DATE	TIM		DEPTH	GENERAL	C	C		CORRECTO		UG/L	UG/L		/100AL
FROM	OF			REMARKS	HG/L	HG/L		CORRECTO	00/2				20
TO	DAY	MEDIUH	1 (1)	Net Milete									20
			0.3					· .		1K	1K	1	
90/01/2	5 159	2 HATER	4		•		.7		•	1K	1	1	
90/01/2	5 154	4 VERI	22.3		•'		.7		,	1K	1	1	
90/01/2	5 160	4 HATER	4		1.		.5		5	1K	1K	z	10K
90/04/1	18 123	6 VERI	21.3		1.		.5		•				10K
90/04/3	18 124	6 HATER	0.3	01									10K
90/05/3	16 121	3 HATER	0.3										2011
90/05/	16 12	4 HATER	0.3					1	5	1K	2	1K	
90/05/	16 12	5 HATER	4		1.		.7	2		îк	2	1K	
90/05/	16 12	17 VERT	4		2.		,8	i		1K	2	1K	4
90/05/	16 12	L8 VERT	4		2.		.7		3	1	1	2	
90/05/	16 12	19 VERT	22.2		1.		.5		2	1K	1K	2	
90/05/	16 12	48 HATER	22.2		1.		5		2	1K	1	2	
90/05/	16 12	49 HATER	22.2		1.		5		4	1	2	2	
90/05/	16 12	50 WATER	22.0	-	2.		.4	-		-			10K
90/06/	27 11	49 VERT	22.3		1.	,8)	6						
90/06/	27 12	03 HATER	0.1						10	1K	1	1	
90/07/	19 11	47 HATER	0.5		2		2.5	-					10K
90/07/	19 11	48 VERT	22.		1	.8 1	1.6						TOK
90/07/	19 12	02 WATER	22.						17	1	2	2	
90/08/	22 11	30 WATER		4	2		2 .1			- ·			10K
90/08/	/22 13	38 VERT			. 1	.9	1.8						TOK
90708/	/22 11	48 HATER	21						9	1	1K	2	
90/09/	/19 13	46 MATER	0.		1		1.8		,	-			
90/09	/19 13	149 VERT		4			1.8						
90/09	/19 1	154 HATER	21.	*									

PGM=RET 1114C 476041 35 49 50.0 084 36 33.0 2 MATTS BAR RESERVOIR 47145 TENNESSEE ROANE 4/145 IENNESSEE TENNESSEE RIVER BASIN TENNESSEE RIVER 560.80 131TVAC 0000 METERS DEPTH 040801

HQ 06010201002 0043.170 OFF

TYPA/AMENT/STREAM/SOLIDS

STORET RETRIEVAL DATE 91/03/06 VITAL SIGNS RESERVOIR MONITORING HY90

DATE FROM TO	TIME OF DAY	MEDIUM	SMK Or Depth (M)	00078 TRANSP SECCHI METERS	00010 WATER TEMP CENT	00300 DO HG/L		00094 CNDUCTVY FIELD MICROMHO 162.0	DATE FROM TO 90/08/2	TIME OF DAY 2 1041	HEDIUN	SMK OR DEPTH (M) 4	00078 TRANSP SECCHI METERS	00010 MATER TEMP CENT 25.4 25.3	00300 DO HG/L 8.1 7.7	00400 PH SU 7.60 7.50 7.40	00094 CNDUCTVY FIELD MICRONHO 186.0 184.0 183.0	1))
90/01/21 90/01/21 90/01/21 90/01/21 90/04/11 90/04/11 90/04/11 90/04/11 90/04/11 90/04/11 90/04/11 90/04/11	5 1200 5 1202 5 1205 8 1107 8 1108 8 1109 8 1112 8 1115 8 1118 8 1121 8 1124	MATER HATER HATER HATER HATER HATER HATER HATER HATER HATER	0.3 1.5 6 0.3 1.5 6 8 10 12 13 0.3	1.12 .98 .92	7.8 7.8 7.8 15.5 15.5 15.4 14.8 14.8 14.5 14.3 14.2 14.3 21.1	11.111.011.011.210.910.610.09.79.79.49.310.0	7.90 7.90 7.90 8.30 8.30 7.90 7.80 7.80 7.80 7.70 7.70 8.30 8.20	162.0 165.0 167.0 146.0 143.0 133.0 130.0 127.0 127.0 126.0 144.0	90/09/3 90/09/3 90/09/ 90/09/	2 1046 2 1049 2 1053 2 1058 9 1044 19 1046 19 1046 19 1046 19 1046 19 1056	MATER WATER MATER MATER	8 10 12 12.5 0.3 1.5 4 6 8 10 12 13	1.00	24.9 24.7 24.7 24.8 24.8 24.8 24.8 24.8 24.8 24.8 24.8	7.0 6.5 6.4 6.2 6.2 6.2 6.2 5.9 5.9	7.30 7.30 7.60 7.60 7.60 7.60 7.60 7.60 7.60 7.6	208. 208. 208. 208. 208. 208. 208. 209.	
90/05/ 90/06/ 90/06/ 90/06/ 90/06/ 90/06/ 90/06/ 90/06/	<pre>16 1040 16 1042 16 1050 16 1055 16 1100 16 1112 27 103 27 103 27 103 27 103 27 103 27 103 27 103 27 103 27 103 27 104 27 104 27 104</pre>	 MATER 	1.5 4 6 8 10 12 13 0.3 1.5 2.5 3.5 5 6 8 10	1.33	20.0 20.3 20.2 20.0 19.9 19.9 26.2 25.7 25.5 25.2 24.3 23.8 23.1 22.6 22.4	9.7 8.8 8.5 8.4 8.2 8.1 12.0 11.5 10.7 10.0 8.4 7.9 7.1 6.7 6.2	7.80 7.80 7.70 7.70 9.10 8.90 8.40 7.70 7.50 7.50 7.30	144.0 144.0 142.0 142.0 142.0 142.0 142.0 142.0 192.0 197.0 200.0 202.0 0 202.0 0 201.0 0 201.0 0 205.0										
90/06/ 90/06/ 90/06/ 90/07/ 90/07/ 90/07/ 90/07 90/07 90/07	<pre>/27 104 /27 104 /27 104 /19 103 /19 103 /19 103 /19 104 /19 104 /19 104 /19 104 /19 104 /19 104 /19 104 /19 104</pre>	2 HATER 3 HATER 15 HATER 15 HATER 16 HATER 16 HATER 11 HATER 11 HATER 11 HATER 12 HATER 14 HATER 15 HATER 16 HATER 16 HATER 16 HATER 16 HATER 16 HATER 17 HATER 17 HATER 16 HATER 16 HATER 16 HATER 17 HATER 16 HATER 16 HATER 16 HATER 16 HATER 16 HATER 17 HATER 17 HATER 17 HATER 17 HATER 18 HATER	10 12 13 0.3 1.5 6 8 10 12 13 0.3 1.5		24.7 24.4 24.3 24.3 23.9 23.8 23.8 23.8	7.0 6.9 6.1 6.0 5.8	7.5	0 209.0 0 170.0 0 168.0 10 167.0 10 168.0 10 168.0 10 168.0 10 168.0 10 168.0 10 174.0 50 174.0				ч. Т					``.	

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PGH=RET

STORET RETRIEVAL DATE 91/03/06 VITAL SIGNS RESERVOIR HONITORING HY90

/TYPA/AHBNT/STREAH/SOLIDS

476041 1114C 35 49 50. 36 33.0 2 MATTS BAR RESERVOIR 47145 TENNESSEE ROANE TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 560.80 131TVAC HQ 060 0000 METERS DEPTH

HQ 06010201002 0043.170 OFF

SHK 84002 00080 00081 82079 00530 00605 00610 00630 00665 00671 PHOS-DIS DATE TIME OR CODE COLOR AP COLOR TURBIDTY RESIDUE ORG N NH3+NH4- NO24NO3 PHOS-TOT FROH OF DEPTH GENERAL PT-CO PT-CO LAB TOT NELT н N TOTAL N-TOTAL ORTHO то DAY MEDIUN (H) REMARKS UNITS UNITS NTU HG/L HG/L HG/L HG/L HG/L P HG/L P 90/01/25 1201 HATER 3.4 10 15 12.0 .160 .060 . 53 .030 .020 8 90/01/25 1202 VERT 15 10 .040 .020 4 12.0 9 .200 .060 .52 90/04/18 1109 VERT 4 15 15 8.8 8 .300 .020 .31 .020 .006 90/04/18 1122 HATER 12.2 20 25 13.0 10 .210 .030 .39 .030 .007 90/05/16 1038 MATER 0.3 D1 90/05/16 1039 MATER 0.3 D2 90/05/16 1040 HATER 0.3 **D**3 90/05/16 1042 VERT D1 25 .030 .21 .030 .004 30 6.0 5 .310 4 90/05/16 1043 VERT .004 D2 20 25 .020 .21 .030 4 7.0 5 .340 90/05/16 1044 VERT D3 20 23 .240 .020 . 21 .030 .004 4 7.0 5 90/05/16 1111 MATER .26 12.3 DÌ 25 30 12.0 10 .110 .050 .030 .004 .004 90/05/16 1112 MATER .030 12.3 DZ 20 23 15.0 12 .150 .050 .26 90/05/16 1113 HATER 12.3 03 20 10 25 12 .25 .030 .004 14.0 .180 .040 90/06/27 1037 VERT 15 .004 5.0 4 .520 .010K .19 .040 4 90/06/27 1044 HATER 12.5 10 15 20.0 16 .250 .030 .40 .040 .007 90/07/19 1039 VERT 7 15 .010K .19 .020 .004 .340 4 6.0 4 90/07/19 1042 HATER 12.2 15 20 9.0 7 .180 .020 . Z6 .020 .005 90/08/22 1041 VERT 10 15 3.0 5 .180 .010K .19 .020 .003 4 90/08/22 1050 HATER 11.7 10 .030 .030 .006 21 . 20 7 10.0 .210 90/09/19 1047 VERT 5 -10 4.0 Б .130 .070 .21 .020 .009 90/09/19 1051 HATER .070 12.2 5 10 .150 .25 .030 .010 6.0 8

DATE From To	TIME OF Day	HEDIUM	SHK OR DEPTH (H)	84002 CODE GENERAL REMARKS	00680 Torg C C Hg/L	00681 D ORG C C HG/L	32211 Chlrphyl A Ug/L Correctd	32212 CHLRPHYL B UG/L	32214 CHLRPHYL C UG/L	32218 PHEOPHTN A UG/L	31616 FEC COLI MFM-FCBR /100ML
90/01/25	1159	HATER	0.3							,	70
90/01/25			3.4		.9	.7	5	1K	1	1	,.
90/01/25	1202	VERT	4		1.0	. 9	5	1	ĩ	ī	
90/04/18			4		1.7	1.6	10	1K	. ī	īκ	
90/04/18	1122	HATER	12.2		1.6	1.5	7	1K	ī	2	
90/05/16	1038	HATER	0.3	D1			-		_	-	10
90/05/16	1039	MATER	0.3	DŻ							10K
90/05/16	1040	HATER	0.3	D3							10
90/05/16	1042	VERT	4	D1	1.8	1.6	14	1	1	1	
90/05/16	1043	VERT	4	DZ	1.8	1.6	14	ī	2	ī	
90/05/16	1044	VERT	4	03	1.9	1.7	14		ī	2	
90/05/16	1111	HATER	12.3	D1	1.7	1.5	6	ī	ī	4	
90/05/16			12.3	02	1.6	1.5	6	ī	ī	i i	
90/05/16	1113	HATER	12.3	D3	1.6	1.5	5	ī	5	ĩ	
90/06/27	1037	VERT	4		2.4	2.2	14	1	ī	3	
90/06/27	1044	HATER	12.5		1.8	1.7		-	-	-	
90/07/19	1036	KATER	0.3								10K
90/07/19	1039	VERT	4		2.0	1.8	10	1K	1	2	
90/07/19	1042	HATER	12.2		1.6	1.4			-	-	
90/08/22	1037	MATER	0.3								10K
90/08/22	1041	VERT	4		2.0	1.9	12	1	1	3	
90/08/22	1050	HATER	11.7		1.9	1.8		-	-	-	
90/09/19	1044	MATER	0.3								10K
90/09/19	1047	VERT	4		1.9	1.9	6	1K	1K	3.	IOR
90/09/19			12.2		1.8	1.9	•	-	**		
, . , . , . , . , . , . , , . , , , , ,	1001	TALES.	****		1.0	1.7					

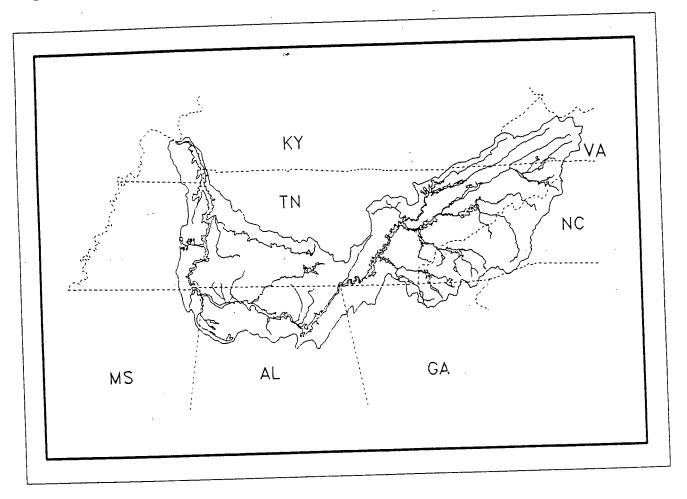
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Water Resources Division Chattanooga, Tennessee TVA/WR--92/1 July 1992

RESERVOIR VITAL SIGNS MONITORING - 1991 PHYSICAL AND CHEMICAL CHARACTERISTICS OF WATER AND SEDIMENT



WATER RESOURCES & ECOLOGICAL MONITORING

INTRODUCTION

In FY 1990, the Tennessee Valley Authority (TVA) initiated a Vital Signs Monitoring program on 12 TVA reservoirs (the nine mainstem Tennessee river reservoirs - Kentucky through Fort Loudoun, and three major tributary reservoirs - Cherokee, Douglas and Norris) as part of its Water Resources and Ecological Monitoring Program (TVA, 1989). In FY 1991, the Vital Signs Monitoring program was expanded to include Melton Hill and Tellico reservoirs to comprise TVA's Basic Vital Signs monitoring strategy, Table 1a, (TVA, 1991). In addition, ten non-navigable tributary impoundments were also added to the program in 1991 which received less intensive monitoring, called TVA's Limited Vital Signs monitoring strategy, Table 1b, (TVA, 1991b).

The objective of the Vital Signs program is to provide basi information on the health or integrity of the aquatic ecosystem in each TVA reservoir and to provide screening level informatior for describing how well each reservoir meets the "fishable and swimmable" goals of the Clean Water Act. This is the first tim in the history of the Agency that a commitment to a long term, systematic sampling of all major TVA reservoirs has been made.

The basis of the Vital Signs monitoring is examination of appropriate physical, chemical and biological indicators in three areas of each reservoir. These three areas are the forebay (the lacustrine region of the reservoir, immediately upstream of the dam), the transition zone (the mid-reservoir region where the water changes from free flowing to more quiescent, impounded water), and the inflow or headwater region of the reservoir. The information gathered is used to make seasonal and spatial assessments of each reservoir's health and the overall health of the reservoir system, and to provide guidance on the need to design and implement more detailed short-term studies where problems seem to exist. In addition, this information establishes a baseline for comparing with future water quality conditions and monitoring water quality trends for TVA

reservoirs.

The Vital Signs program employs several activities to assess reservoir health. They include physical and chemical characteristics of water and sediment, acute toxicity screening of water and sediment, benthic macroinvertebrate population assessments, and fish abundance, composition and health.

This report presents 1991 information on the physical and chemical characteristics of water and sediment collected on the present twenty four Vital Signs reservoirs. Sampling was conducted at the forebay and transition zone locations on the largest eighteen reservoirs. Only the forebay location was sampled on the remaining six, smaller reservoirs. Water and sediment samples were not collected at reservoir inflow locations because: (1) for several tributary reservoirs, TVA's ambient stream monitoring program (Parr, 1991) incorporates sampling at inflow locations; (2) for mainstem or run-of-the-river reservoirs, inflow water sampling locations are tailrace locations and are dominated by the effects of the impoundment immediately upstream, rather than the processes occurring in the downstream reservoir being sampled; and (3) the lack of suitable substrate often precluded the collection of sediment samples at reservoir inflow tailrace locations.

The Reservoir Vital Signs Monitoring locations sampled in 1991 are shown on Figure 1 and listed in Table 1.

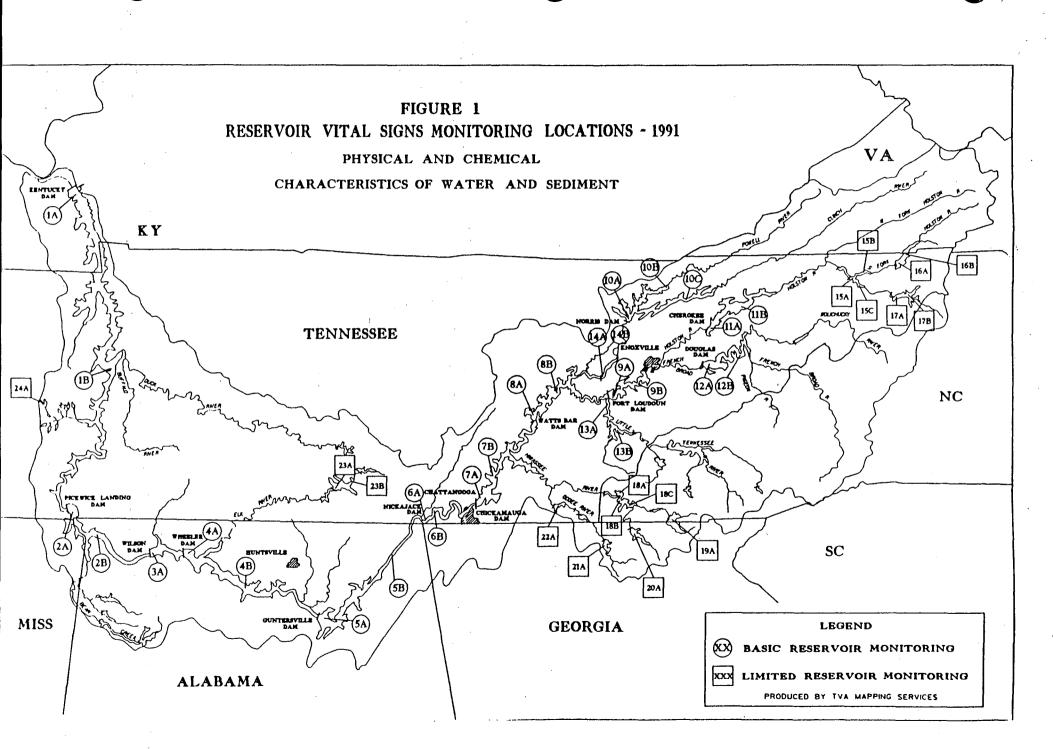


Table la

WATER QUALITY MONITORING LOCATIONS RESERVOIR VITAL SIGNS MONITORING, 1991

Basic Water Quality Monitoring Locations

	Foreb	Transition Zone Locations							
		Map ID	Storet		Map ID	Storet			
Reservoir	River Mile	Number	Station #	<u>River Mile</u>	<u>Number</u>	<u>Station #</u>			
	(Tennessee)	· · · · · · · · · · · · · · · · · · ·	· · · · · · · · · · · · · · · · · · ·	(Tennessee)					
	(10mme)	Mai	n Stream						
Kentucky	23.0	1A.	202832	112.0	1B.	475015			
Pickwick	207.3	2A.	476799	230.0	2B.	016923			
Wilson	260.8	3A.	016912						
Wheeler	277.0	4A.	016900	307.5	4B.	017012			
Guntersville	350.0	5A.	017261	396.8	5B.	017101			
Nickajack	425.5	6A.	476344	433.0	6B.	476239			
Chickamauga	472.3	7A.	475358	490.5	7B.	475265			
Watts Bar	531.0	8A.	475317	560.8	8B.	476041			
Fort Loudoun	603.2	9A.	475602	624.6	9B.	475603			
fort boudban									
		Tr	ibutary ·						
Norris	CRM 80.0	10A.	476009	PRM 30.0	10B.	477187			
NOLTID				CRM 125.0	10C.	477186			
Cherokee	HRM 53.0	11A.	475025	HRM 76.0	11B.	475028			
Douglas	FBRM 33.0	12A.	475081	FBRM 60.7	12B.	475993			
Tellico	LTRM 1.0	13A.	476260	LTRM 21.0	13B.	476295			
Melton Hill	CRM 24.0	14A.	477064	CRM 45.0	14B.	476194			
Mercon niri			;						

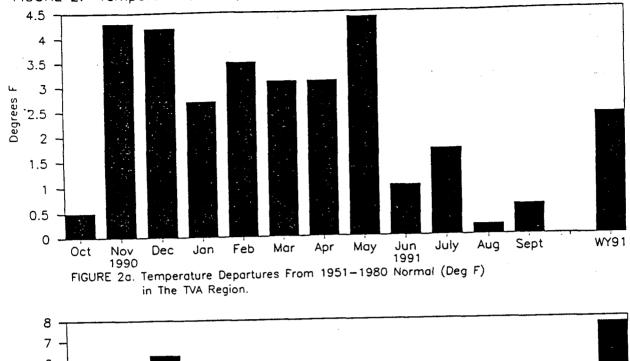
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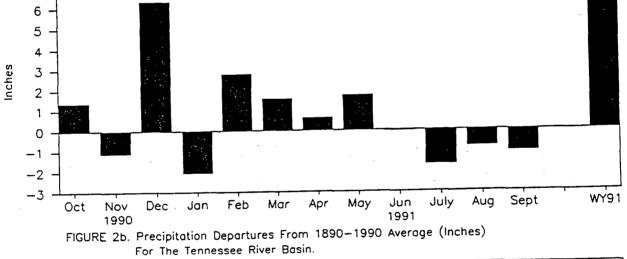
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HYDROLOGIC OVERVIEW OF WATER YEAR 1991

Seasonal variations in atmospheric temperature and rainfall have a direct impact on water quality. Consequently, many water quality characteristics (temperature, dissolved oxygen, conductivity, turbidity, suspended solids, etc.) exhibit seasonal effects. During the dry season, when runoff is minimal. streamflow is derived principally from the base flow of groundwater. Groundwater contains greater concentrations of dissolved minerals than does surface drainage because of increased water/soil/rock contact and longer groundwater residence time. During the wet season, streamflow is principally derived from rapid overland runoff that allows little time for mineral dissolution. Consequently, lower concentrations of most dissolved constituents are added to a river during heavy rainfall and subsequent high flows. However, periods of intense rainfall and high overland flows wash off or "flush" a watershed and transport soil particles to streams. This carries large loads of nonpoint source pollutants (nutrients, suspended solids, turbidity, etc.) to streams and rivers. Therefore, examination of atmospheric temperature, rainfall and runoff patterns during Water Year (WY) 1991 aids in interpretation of the Reservoir Vital Signs Monitoring data.

Atmospheric Temperature - The average annual temperature in the TVA Region is approximately 60 degrees Fahrenheit (°F), with January usually being the coldest month and July the hottest. During WY 1991, atmospheric temperatures in the TVA Region averaged about 2.4 degrees Fahrenheit (°F) warmer than for the normal period, 1951-1980, (USDOC, 1990 and 1991). All twelve months of WY 1991 experiencing above normal temperatures. The greatest monthly departures (about 4 °F above normal) were in November and December 1990 and May 1991, Figure 2a. January through April also had departures about three degrees Fahrenheit above normal. The result was that the seven month period





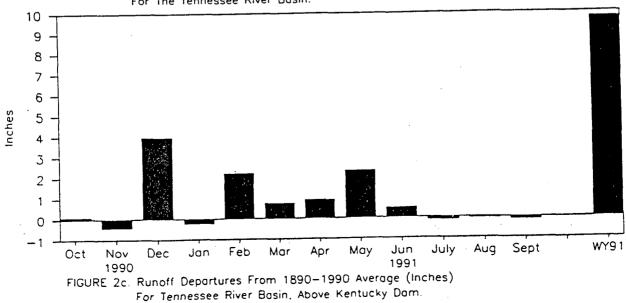


FIGURE 2. Temperature, Precipitation, and Runoff – Tennessee River Basin, WY-91

(November 1990-May 1991) was unusually warm - averaging over 3.5 degrees above normal. Although August was near normal, spells of hot, dry weather occurred in the summer, with conditions somewhat worse to the west and south of Chattanooga. This pattern persisted through September.

Rainfall - The Tennessee River basin averages about 52 inches of precipitation annually. However, there are large variations in the spatial distribution of precipitation. The range is from a high of about 93 inches in the mountains of southwestern North Carolina near Highlands to a low of about 37 inches in the shielded valleys of these same mountains near Asheville, North Carolina. Elsewhere in the Valley, precipitation usually ranges within five to ten inches of the basin average. March is usually the wettest month and October the driest. WY 1991 was wetter than the previous year, but resembled the pattern of a wetter than average cool season and a The precipitation in dryer than average warm season, Figure 2b. WY 1991 for the Tennessee River basin was slightly in excess of 59 inches, a departure of 15 percent above the 101-year long term average, with that portion of the basin downstream of Chattanooga generally wetter than the portion of the basin upstream of Chattanooga. Precipitation in November 1990 and January 1991 was below average, but the other months in the October 1990 through May 1991 period were wet enough to exceed the long term average for this eight month period by 11 inches. For the Tennessee River basin, December 1990 was the wettest December on record averaging 11.1 inches of precipitation, which was 6.3 inches above the long term average for December. The period of June through September was about 3.5 inches drier than average for the Tennessee River basin (TVA 1991c).

<u>Streamflow</u> - Streamflow varies seasonally with rainfall, although during the spring and summer evapotranspiration reduces the amount of runoff somewhat. Watersheds that receive 50 to 60

inches of precipitation annually average about 20 to 30 inches of In a normal year, the discharge of the Tennessee River (approximately 64,000 cfs) corresponds to about 22 inches of runoff. runoff distributed over the 40,900 square mile drainage basin. A larger amount of runoff occurs during the wet winter and spring months when precipitation events are frequent, temperatures are low, and there are no leaves on deciduous vegetation. Consequently soil absorption, evaporation and transpiration losses are low at that time of year, and both runoff and streamflow are higher than during the summer and fall months. In WY 1991 there was an unusually high amount of precipitation and runoff, particularly in December and during the period February through May. Much of this runoff was held in storage in tributary reservoirs and later released such that during the subsequent dry period of June through September, even though rainfall was substantially below normal, streamflow of the Tennessee River at Kentucky Dam was near normal (Figures 2b and The net result for WY 1991 was an annual 15% excess in precipitation with resultant total runoff that was approximately ten inches above the long term mean of 22.4 inches. Mean flows during 1991 for each of the Vital Signs reservoirs reflect the higher-than-average annual runoff (Table 2).

1.3 meters, 5.2 NTU's, and 4.4 mg/l, respectively; transition zone Secchi depths, turbidity, and suspended solids averaged 1.4 meters, 5.4 NTU's, and 5.4 mg/l, respectively. True color values averaged about 11-12 PCU's at the forebay and transition zones. These data show the light transparency of Nickajack Reservoir to fall within the mid-range of observations at the Vital Signs monitoring locations.

Additional Comments - Water quality data collected as part of the Vital Signs monitoring program for Nickajack Reservoir in 1990 and 1991 have shown the physical/chemical water quality characteristics of the forebay and the transition zone to be very similar. Because of the well mixed hydrologic conditions of Nickajack Reservoir and the relatively short distance between the forebay (TRM 425.5) and the transition zone (TRM 433.0), future Vital Signs monitoring will be conducted only at the forebay.

Chickamauga Reservoir

In situ Measurements - Measurements of temperature, dissolved oxygen (DO), pH and conductivity show Chickamauga Reservoir to be generally well mixed and lacking any strong thermal stratification (Appendix A-7, Table 13). Surface temperatures ranged from 8.3°C in January to 29.9°C in July in the forebay; and from 7.1°C to 27.6°C for the same months at the transition zone.

Values for DO at the 1.5 meter depth ranged from 10.4 mg/l in April to 6.0 mg/l in August at the forebay; and from 10.1 mg/l in January to 5.5 mg/l in August at the transition zone. Contour plots of temperature and DO versus depth for the period January through the end of September for both the forebay (Figure 17) and the transition zone (Figure 18) depict the seasonal variation and rather weak stratification of Chickamauga Reservoir in 1991.

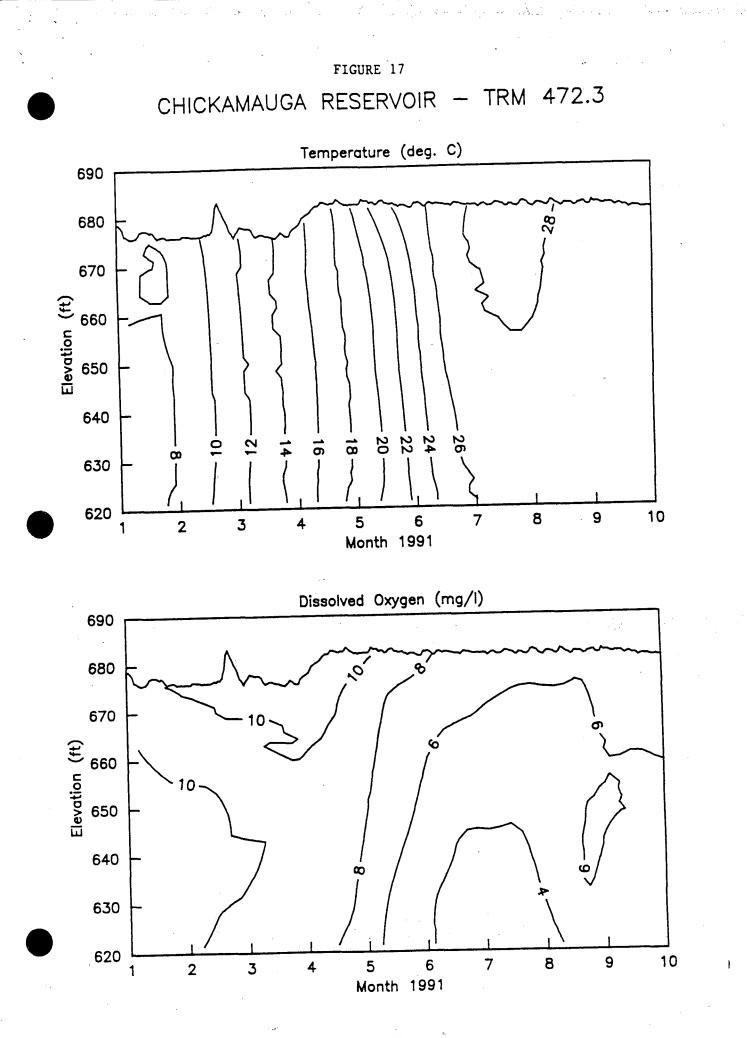
Values of pH ranged from 7.0 to 8.2 on Chickamauga Reservoir. Conductivities ranged from 117 to 182 μ mhos/cm, and averaged about 160 μ mhos/cm. Comparison of pH and conductivity at the transition zone with upstream pH and conductivity at Watts

	Table	13	-
Chickamauga	Reservoir -	Water	Quality Summary
Reservoir	Vital Signs	Monit	oring, WY 1991

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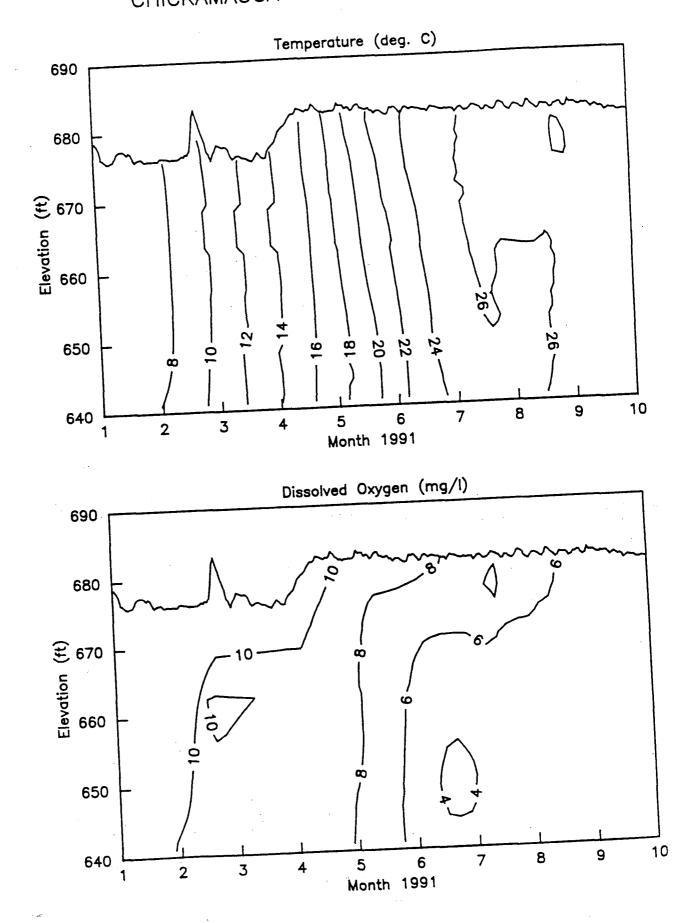
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VariableNHeanMinMaxNIteanTemperature ('C)78 22.52 7.90 29.90 55 21.18 7.10 27.60 Dissolved Oxygen (mg/1)78 6.93 3.00 11.10 55 7.14 3.40 10.30 pH (s.u.)78 6.93 3.00 11.10 55 7.52 7.00 8.10 Conductivity (umhos/cm)78 159.06 137.00 182.00 55 156.93 117.00 177.00 Conductivity (umhos/cm)18 0.21 0.03 0.37 18 0.19 0.06 0.38 Ammonia - N (mg/1)18 0.24 0.11 0.50 18 0.49 0.35 0.59 Nitrate+Nitrite - N (mg/1)18 0.25 0.020 0.030 18 0.24 0.024 0.020 0.030 Total Phosphorus (mg/1)18 0.008 0.002 0.020 18 0.09 0.004 0.224 Dissolved Ortho - P (mg/1)18 1.84 1.70 2.00 18 1.88 1.70 2.24 Soluble Organic Carbon (mg/1) 9 6.78 2.00 10.00 9 7.22 1.00 13.00 Chiorophyl1-a (ug/1) 6 1.45 1.16 1.75 7 1.40 1.13 1.7 Sechi dept (m) 6 1.45 1.16 1.75 7 1.40 1.13 1.7 Sechi dept (m) 6 1.45 1.1	,		2			T=-	neition 2	one (TRM	490.47)
VariableNMeanMinMaxITemperature ('C)78 22.52 7.90 29.90 55 21.18 7.10 27.60 Dissolved Oxygen (mg/1)78 6.93 3.00 11.10 55 7.14 3.40 10.30 pH (s.u.)78 7.46 7.00 8.20 55 7.52 7.00 8.10 Conductivity (umhos/cm)78 159.06 137.00 182.00 55 156.93 117.00 177.00 Organic - N (mg/1)18 0.21 0.03 0.37 18 0.19 0.06 0.38 Ammonia - N (mg/1)18 0.24 0.11 0.50 18 0.49 0.35 0.59 Total Nitrogen (mg/1)18 0.25 0.202 0.030 18 0.024 0.020 0.020 Total Phosphorus (mg/1)18 0.025 0.020 0.030 18 0.099 0.004 0.024 Dissolved Ortho - P (mg/1)18 0.08 0.002 0.020 17 1.78 1.60 2.00 Soluble Organic Carbon (mg/1) 18 1.76 1.60 1.90 17 1.78 1.60 2.00 Secchi depth (m) 6 1.45 1.16 1.75 7 1.40 1.13 1.7 Truchidity (NTU) 18 6.78 1.00 11.00 18 4.17 1.00 7.0			Forebay		.3)			Min	<u>Max</u>
Temperature (°C)78 22.52 7.90 29.90 55 21.13 1.4 3.40 10.30 Dissolved Oxygen (mg/1)78 6.93 3.00 11.10 55 7.14 3.40 10.30 pH (s.u.)77 7.46 7.00 8.20 55 7.52 7.00 8.10 Conductivity (umhos/cm)78 159.06 137.00 182.00 55 156.93 117.00 177.00 Organic - N (mg/1)18 0.21 0.03 0.37 18 0.19 0.06 0.38 Ammonia - N (mg/1)18 0.24 0.11 0.50 18 0.25 0.14 0.47 Nitrate+Nitrite - N (mg/1)18 0.24 0.11 0.50 18 0.24 0.020 0.030 Total Nitrogen (mg/1)18 0.025 $0.26.5$ 18 20.8 11.7 29.5 Total Phosphorus (mg/1)18 0.08 0.002 0.030 18 0.024 0.020 0.034 Dissolved Ortho - P (mg/1)18 1.84 1.70 2.00 18 1.88 1.70 2.00 Soluble Organic Carbon (mg/1) 18 1.76 1.60 1.90 17 1.78 1.60 2.00 Soluble Organic Carbon (mg/1) 6 1.45 1.16 1.75 7 1.40 1.13 1.7 Secchi depth (m) 6 1.45 1.16 1.75 7 1.40 1.13 1.7 Secchi de	Variable	N		<u>Min</u>	Max	_11			27 60
Suspended Solids (mg/1) 12 10.42 5.00 15.00 12 17.50 10.00 25.0	Temperature ('C) Dissolved Oxygen (mg/l) pH (s.u.) Conductivity (umhos/cm) Organic - N (mg/l) Ammonia - N (mg/l) Nitrate+Nitrite - N (mg/l) Total Nitrogen (mg/l) Total Phosphorus (mg/l) Total Phosphorus (mg/l) TN/TP Ratio Dissolved Ortho - P (mg/l) Total Organic Carbon (mg/l) Soluble Organic Carbon (mg/l) Chlorophyll-a (ug/l) Secchi depth (m) Turbidity (NTU) Suspended Solids (mg/l) True Color (PCU)	78 78 77 78 18 18 18 18 18 18 18 18 18 18 18 18 18	<u>Mean</u> 22.52 6.93 7.46 159.06 0.21 0.05 0.24 0.50 0.025 20.5 0.008 1.84 1.76 6.78 1.45 5.72 4.78 10.42 17.92	<u>Min</u> 7.90 3.00 7.00 137.00 0.03 0.01 0.11 0.30 0.020 15.0 0.002 1.70 1.60 2.00 1.16 3.00 1.00 5.00 10.00	29.90 11.10 8.20 182.00 0.37 0.09 0.50 0.60 0.030 26.5 0.020 2.00 1.90 10.00 1.75 12.00 11.00 15.00 30.00	55 55 55 18 18 18 18 18 18 18 18 18 18 17 9 7 18 18 18 12 12	21.187.147.52156.930.190.050.250.490.02420.80.0091.881.787.221.404.564.1710.0017.50	3.40 7.00 117.00 0.06 0.01 0.14 0.35 0.020 11.7 0.004 1.70 1.60 1.00 1.13 3.00 1.00 5.00 10.00	10.30



CHICKAMAUGA RESERVOIR - TRM 490.5

FIGURE 18



Bar Dam forebay shows that pH and conductivity are lowered by the effect of the Hiwassee River inflows to Chickamauga Reservoir about 9 miles upstream of the transition zone.

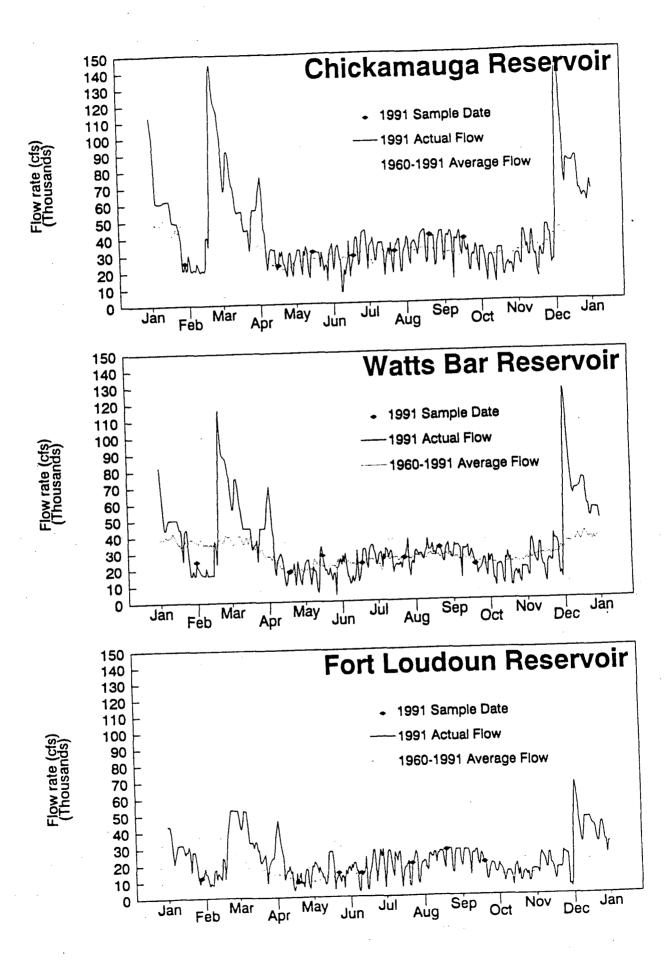
<u>Biochemical Measurements</u> - In Chickamauga Reservoir, during 1991, concentrations of organic nitrogen, ammonia nitrogen, nitrite plus nitrate nitrogen, and total phosphorus averaged 0.21, 0.05, 0.24 and 0.025 mg/l, respectively at the forebay; and 0.19, 0.05, 0.25 and 0.024 mg/l, respectively, at the transition zone. Average total nitrogen concentrations were the lowest measured among Vital Signs monitoring locations on the Tennessee River in 1991. In addition, both total phosphorus and dissolved ortho phosphorus concentrations were also among the lowest observed at any of the Vital Signs monitoring locations on the Tennessee River.

TN/TP ratios ranged from 12 to 30 in Chickamauga Reservoir, indicating extended periods of phosphorus limiting conditions. The highest chlorophyll-a concentrations were measured in May ranging from 8-10 μ g/l at the forebay and 11-13 μ g/l at the transition zone. The surface concentrations of chlorophyll-a averaged approximately 7 μ g/l at both the Chickamauga Reservoir forebay and the transition zone in 1991.

Organic carbon concentrations, total and soluble, in Chickamauga Reservoir were quite low, averaging between 1.9 and 1.8 mg/l, at both the forebay and the transition zone.

Physical Measurements - Figure 19 illustrates the average daily discharge from Chickamauga Reservoir in 1991 with water quality sampling dates superimposed. None of the water quality surveys conducted in 1991 on Chickamauga Reservoir occurred on days with unusually high or low flows. Consequently the variation in those measures of water clarity (Secchi depth, turbidity, suspended solids) is quite small. Forebay Secchi depth, turbidity, and suspended solids measurements averaged 1.4 meters, 5.7 NTU's, and 4.8 mg/l, respectively. Transition zone Secchi depths, turbidity, and suspended solids averaged 1.4 meters, 4.6 NTU's, and 4.2 mg/l, respectively. In addition, true

FIGURE 19



color values, averaged about 10 PCU's, at the forebay and transition zones, respectively. Together, these values indicate the light transparency of Chickamauga Reservoir to be high compared with the other mainstem Tennessee River reservoirs.

<u>Watts Bar Reservoir</u>

In situ Measurements - Temperature, dissolved oxygen (DO), pH and conductivity measurements show the reservoir to be well mixed early in the year and developing just a moderate degree of thermal stratification at the forebay in July and August (Appendix A-8, Table 14). Surface water temperatures ranged from 7.2°C in January to 30.2°C in July in the forebay, and from 7.7°C to 28.4°C for these same months at the transition zone.

Values for DO at the 1.5 meter depth ranged from 12.8 mg/l in April (due to high photosynthetic activity) to 8.1 mg/l in September at the forebay, and from 11.2 mg/l in January to 6.6 mg/l in September at the transition zone. Contour plots of temperature and DO versus depth for the period January through the end of September for both the forebay (Figure 20) and the transition zone (Figure 21) depict the seasonal variation with a weak summer time thermal stratification and a rather strong oxycline in the forebay of Watts Bar Reservoir. From the surface to the bottom of the reservoir, at the forebay, the data show an 8°C decrease in temperature in May, and about a 10 mg/l decrease in DO in June and July; with near bottom DO concentrations in the hypolimnion of Watts Bar forebay less than 1 mg/1.

Values of pH ranged from 7.0 to 9.2 on Watts Bar Reservoir. In April, May, June, and July near surface values of pH in the forebay were high, equal to or exceeding 9.0 and with DO saturation values ranging from 125-150%, giving evidence of very high rates of photosynthesis.

Conductivities ranged from 126 to 208 μ mhos/cm and averaged about 175 μ mhos/cm, with highest conductivities coinciding with lower streamflows in January.

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Table 14 Watts Bar Reservoir - Water Quality Summary Reservoir Vital Signs Monitoring, WY 1991

Variable Temperature (°C) Dissolved Oxygen (mg/l) pH (s.u.) Conductivity (umhos/cm) Organic - N (mg/l) Ammonia - N (mg/l) Nitrate+Nitrite - N (mg/l) Total Nitrogen (mg/l) Total Phosphorus (mg/l) TN/TP Ratio Dissolved Ortho - P (mg/l) Total Organic Carbon (mg/l) Soluble Organic Carbon (mg/l) Chlorophyll-a (ug/l) Secchi depth (m) Turbidity (NTU) Suspended Solids (mg/l) True Color (PCU) Apparent Color (PCU) Fecal Coliform (#/100 ml)	<u>N</u> 124 124 124 124 18 18 18 18 18 18 18 18 18 18	Forebay Mean 21.32 6.80 7.79 169.78 0.20 0.04 0.28 0.52 0.021 29.0 0.008 1.97 1.77 11.78 1.51 4.39 5.67 9.19 18.44 10.00	(TRM 531. Min 7.00 0.30 7.00 126.00 0.03 0.01 0.01 0.22 0.009 16.3 0.002 1.70 1.50 6.00 1.26 1.00 2.00 2.00 5.00 10.00	30.20 13.40 9.20 188.00 0.44 0.12 0.51 0.61 0.030 57.0 0.020 2.40 2.00 19.00 1.73 10.00 15.00 25.00 60.00	<u>Tra</u> <u>N</u> 66 66 66 18 18 18 18 18 18 18 18 18 18 18 18 18	<u>Mean</u> 20.93 8.11 7.76 175.58 0.22 0.03 0.34 0.60 0.027 23.2 0.009 1.87 1.71 7.78 1.10 6.00 7.56 11.06 16.25 10.00	MIN 7.60 4.40 7.20 126.00 0.03 0.01 0.17 0.45 0.020 11.5 0.002 1.40 1.30 5.00 0.75 3.00 2.00 5.00	$\begin{array}{c} 110.2 \\ 28.40 \\ 11.70 \\ 8.80 \\ 208.00 \\ \hline 0.78 \\ 0.07 \\ 0.61 \\ 0.99 \\ 0.040 \\ 49.5 \\ 0.020 \\ 2.30 \\ 2.00 \\ 13.00 \\ 1.46 \\ 13.00 \\ 15.00 \\ 35.00 \\ 35.00 \\ 35.00 \\ \end{array}$
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FIGURE 20 WATTS BAR RESERVOIR - TRM 531.0

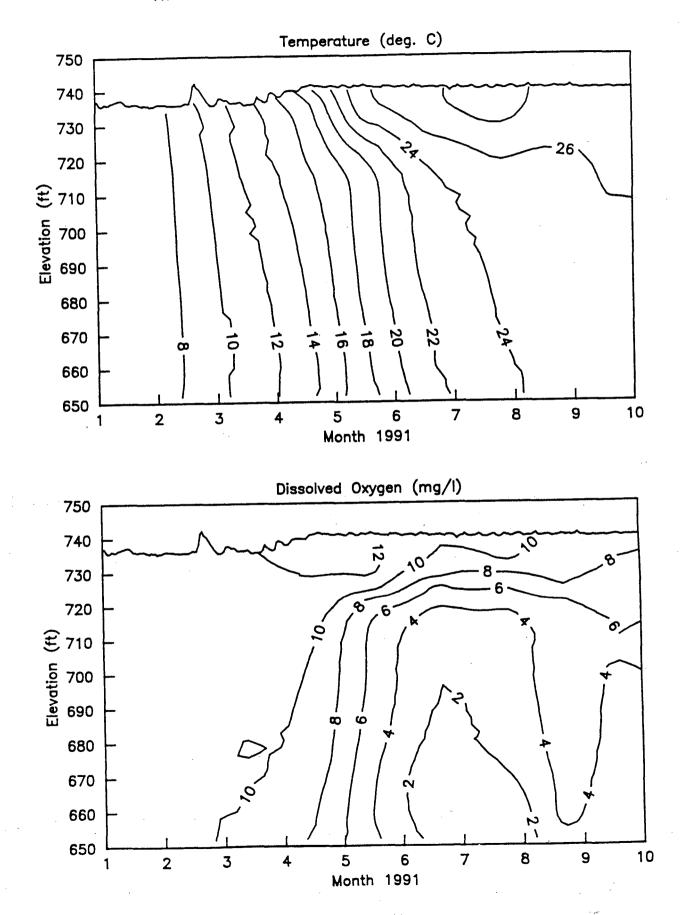
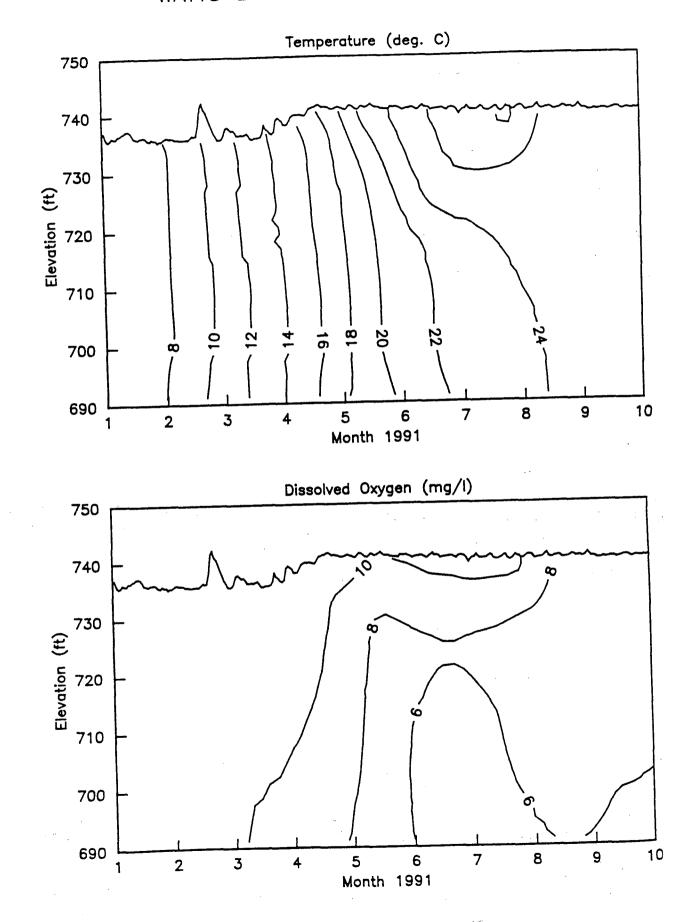


FIGURE 21

WATTS BAR RESERVOIR - TRM 560.8



<u>Biochemical Measurements</u> - In Watts Bar Reservoir during 1991, concentrations of organic nitrogen, ammonia nitrogen, nitrite plus nitrate nitrogen, and total phosphorus averaged 0.20, 0.04, 0.28 and 0.021 mg/l, respectively at the forebay; and 0.22, 0.03, 0.34 and 0.027 mg/l, respectively, at the transition zone. The average total phosphorus concentrations observed at the forebay on Watts Bar Reservoir were lower than any of the other Tennessee River Vital Signs monitoring locations. The average dissolved ortho phosphorus concentrations of 0.008 and 0.009 mg/l, respectively, at the forebay and transition zones were essentially identical to the average concentrations of dissolved ortho phosphorus in Chickamauga Reservoir, and were among the lowest observed at any of the Tennessee River Vital Signs monitoring locations in 1991.

TN/TP ratios ranged from 12 to 57 in Watts Bar Reservoir, indicating (at least) intermittent periods of phosphorus limiting conditions. Watts Bar forebay's lowest observed TN/TP ratios were 16; however, high chlorophyll-a concentrations were measured in the forebay of Watts Bar Reservoir. The highest chlorophyll-a concentrations were measured in August, 19 μ g/l at the forebay, and in July, 13 μ g/l at the transition zone. Surface concentrations of chlorophyll-a pigment averaged about 12 μ g/l at the Watts Bar Reservoir forebay, and about 8 μ g/l at the transition zone in 1991.

Organic carbon concentrations (both total and soluble) in Watts Bar Reservoir were low, averaging 2.0 and 1.8 mg/l, at the forebay and 1.9 and 1.7 mg/l at the transition zone.

<u>Physical Measurements</u> - Figure 19 illustrates the average daily discharge from Watts Bar Reservoir in 1991 with sampling dates superimposed. Forebay Secchi depth, turbidity, and suspended solids measurements averaged 1.5 meters, 4.4 NTU's, and 5.7 mg/l, respectively. Transition zone Secchi depths, turbidity, and suspended solids averaged 1.1 meters, 6.0 NTU's, and 7.6 mg/l, respectively. These values indicate the light transparency of Watts Bar Reservoir forebay to be among the

highest of the mainstem Tennessee River reservoirs, in 1991. True color values, averaged 9 and 11 PCU's, at the forebay and transition zones, respectively, also falling among the lower values measured at mainstem Tennessee river Vital Signs monitoring locations. の時間になるので、

Fort Loudoun Reservoir

In situ Measurements - Measurements of temperature, dissolved oxygen (DO), pH and conductivity show the reservoir is well mixed early in the year but develops a fairly strong thermal stratification from June through September (Appendix A-9, Table 15). Surface water temperatures ranged from 7.5°C in January to 28.2°C in July at the forebay, and from 6.8°C to 29.6°C for the same months at the transition zone. In July, at the forebay, the temperature decreased over 8°C from water surface to bottom. The reservoir remained stratified through September.

Values for DO at the 1.5 meter depth ranged from 12.3 mg/l in April (algal bloom) to 8.9 mg/l in August at the forebay, and from 14.6 mg/l in July (algal bloom) to 5.7 mg/l in August at the transition zone. Contour plots of temperature and DO versus depth for the period January through the end of September for both the forebay (Figure 22) and the transition zone (Figure 23) depict the seasonal variation and summer time stratification of Fort Loudoun Reservoir. Hypolimnetic DO concentrations measured in the forebay of Fort Loudoun Reservoir in 1991 were higher than in 1990. For example, in the hypolimnion in 1990, DO's were measured below 2 mg/l in July and August; but, in 1991 no DO's were measured below 3 mg/l.

Values of pH ranged from 6.7 to 9.1 on Fort Loudoun Reservoir. At the forebay, values of pH exceeding 8.5 and DO saturation values exceeding 120% were measured each month, April through September, giving evidence of much photosynthetic activity. During these same months, the same pattern of high pH and high DO saturation was observed, although to a lesser extent, at the transition zone.

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 CHICKAMAUGA RES. AT LIGHTED BUOY

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 PGM=RET 06020001021 0000.710 ON

/TYPA/AMBNT/STREAM/SOLIDS

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91/01/28 91/04/16	1107 0958	WATER	12.8		.030	. 020	1.8	1.9	2.00	1.00	1.00	1.00	
91/04/16 91/05/15	1008	WATER WATER	13.8	01	.020 .030	.005	1.8 1.7	1.7 1.6	5.00	1.00	1.00	1.00	
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STORET RETRIEVAL DATE 92/05/16 475265 1053 35 18 00.0 085 04 33.0 2 CHICKAMAUGA RESERVOIR 47065 TENNESSEE HAMILT TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 490.47 PGM≖RET HAMILTON 040801 /TYPA/AMBNT/STREAM/SOLIDS 06020001025 0005.740 ON 131TVAC 0000 NETERS DEPTH

INDEX 1021500 007720 MILES 0953.80 0046.50 4 DATE TIME FROM OF	SMK OR DEPTH	84002 CODE GENERAL	00078 TRANSP SECCHI METERS	31616 FEC COLI MFM-FCBR /100ML	00080 COLOR PT-CO UNITS	00081 AP COLOR PT-CO UNITS	82079 TURBIDTY LAB NTU	00530 RESIDUE TOT NFLT MG/L		00610 NH3+NH4- N TOTAL MG/L	00630 N02&N03 N-TOTAL MG/L
TO DAY MEDIUM	(H) 0.3	REMARKS	1.68	,		15	4.0	1 K	.060	.060	. 46
91/01/28 0940 WATER 91/01/28 0947 VERT	4				10 10	15	3.0	1 K	.070	.050	
91/01/28 0952 WATER 91/04/16 0858 WATER 91/04/16 0904 VERT	7.8 0.3 4		1.13	20	5	10 10	5.0 6.0	3 6	. 150 . 100	.070 .080	.34
91/04/16 0911 WATER 91/05/15 0908 WATER	9.8 0.3 0.3	D 1 D 2	1.38	10K 10K							27
91/05/15 0909 WATER 91/05/15 0910 WATER	0.3	D 3		10K			4.0	4	.330	.030 .020	. 23
91/05/15 0920 VERT 91/05/15 0921 VERT	4	D 1 D 2					4.0 4.0	3	. 150	.050	. 22 . 23
01/05/15 0922 VERT	4 9.6000	D3 D1					6.0 6.0	6	.180 .180	.060	. 24
91/05/15 0924 WATER 91/05/15 0925 WATER	9.6000	D 2					6.0	Ĩ.	.140	.040	. 23
91/05/15 0926 WATER 91/06/18 0817 WATER 91/06/18 0821 VERT	9.6000	D 3	1.25	10K	10 15	20 25	4:0 7.0	5 7	.220 .200	.010K .040	. 25
91/06/18 0825 WATER 91/07/23 0855 WATER	9 0.5		1.26	. 10K	. 10	20	3.0	3	.380	.020 .060	. 17 . 26
01/07/23 0901 VERT	9.4				10	20	4.0	4	. 180		
91/07/23 0912 WATER 91/08/20 0950 WATER 91/08/20 0956 VERT	0.5		1.78	10K	15 10	20 25	4.0 5.0	4 7	.240 .240	.030	. 21 . 22
91/08/20 1005 WATER 91/09/17 0935 WATER	9.4 0.5		1.30	10	15	20	3.0	3	.180	.050 .080	. 16
91/09/17 0940 VERT 91/09/17 0945 WATER	4 9.2000				5			5	.130	.000	• • •
DATE TIME	SMK OR DEPTH	84002 CODE GENERAL	00665 PHOS-TOT	ORTHO	00680 T ORG C C	00681 D ORG C C	32211 CHLRPHYL A UG/L	32212 CHLRPHYL B UG/L	32214 CHLRPHYL C UG/L	32218 PHEOPHTN A UG/L	
FROM OF TO DAY MEDIUM	(M)	REMARKS	MG/L P	MG/L P	MG/L	MG/L	CORRECTO		1.00	1.00	
91/01/28 0947 VERT	4		.030	.020	2.1		1.00	1.00			
91/01/28 0952 WATER 91/04/16 0904 VERT 91/04/16 0911 WATER	7.8 4 .9.8 0.3	D 1	.030 .030 .030	.006	1.8	1.8	4.00	2.00	3.00	3.00	
91/05/15 0908 WATER 91/05/15 0909 WATER	0.3	D 2						r.			
91/05/15 0910 WATER 91/05/15 0920 VERT	0.3	03 D1	.020	.004	1.9			1.00K 1.00K		1.00	
91/05/15 0921 VERT	4	D 2 D 3	.020	.004	1.9	1.8	11.00	1.00K		1.00	
91/05/15 0922 VERT 91/05/15 0924 WATER	9.6000	D 1	.020	.007	1.7				,		
91/05/15 0925 WATER 91/05/15 0926 WATER	9.6000 9.6000 4	D 2 D 3	.020 .020 .020	.006	1.7	1.6	11.00	1.00%	1.00	3.00	
91/06/18 0821 VERT 91/06/18 0825 WATER	9		.020		1.9	1.7	14.00		1.00	1.00	
91/07/22 1200 WATER 91/07/23 0901 VERT	0.3	•	. 020		2.0 1.8			1.00K		2.00	
91/07/23 0912 WATER	9.4		.020	.010	. 1.9	1.8	3.00	1.00%	1.00%	1.00	
91/08/20 0956 VERT	Q , 4		.030		1.8			1.00%	1,00	2.00	

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DATE FROM TO	T I ME OF DAY	MEDIUN	SMK OR DEPTH (M)	00060 STREAN FLOW CFS	00010 WATER TEMP CENT	00300 D0 MG/L	00301 DO Satur Percent	00400 Рк Su	00094 CHDUCTVY FIELD MICROMHO	DATE FROM TO
91/07/2			8		24.9	3.4	40.5\$	7.30	181	
91/07/2			10		24.8	3.3	39.35	7.30	182	91/09/19 91/09/19
91/07/2			12		24.6	3.3	39.35	7.30	184	91/09/19
91/07/2		VALER	14		24.5	3.2	37.65	7.30	184	91/09/19
91/07/2			18	•	24.3	2.9	34.15	7.20	181	91/09/19
91/07/2			. 20		24.2	2.7	31.85	7.20	181	91/09/19
91/07/2			22		23.9	1.5	17.65	7.20	181	91/09/19
91/07/24	1145	WATER	23.5		23.8		10.65	7.10	176	
91/07/24			24		23.7	. 4	4.75	7.00	171	
91/07/24			26		23.6	. 3	3.55	7.00	173	
91/08/21			0.5	31525	26.5	9.5	115.95	8.70	180	
91/08/21 91/08/21			1.5		26.5	9.3	113.45	8.70	180	
91/08/21			1.3		26.4	9.1	111.05	8.70	181	
91/08/21					26.3 26.2	8.2 7.8	100.05	8.50	181	
91/08/21			5.5		26.0	6.4	78.05	8.40	182	
91/08/21			6		25.5	5.1	60.75	7.70	184	
91/08/21			8		25.1	4.7	56.05	7.50	183	
91/08/21			10		25.0	4.7	56.05	7.50	182	
91/08/21			12		24.9	4.6	\$4.85	7.40	181	
91/08/21			15		24.8	4.6	54.8\$	7.40	181	
91/08/21 91/08/21			16 18		24.8	4.6	54.85		- 181	
91/08/21			20		24.7 24.7	4.5	53.68 53.68	7.40	179	
91/08/21			žž		24.7	4.4	52.4\$	7.40	179 178	
91/08/21			24		24.7	111	52.48	7.40	181	
91/08/21	1156	VATER	25.5		24.6	4.4	52.48	7.40	181	
91/09/19			0.5	20779	26.7	8.6	106.25	8.70	183	
91/09/19			1		26.7	8.6	106.25	8.60	183	
91/09/19			1.5		26.7	8.1	100.05	8.60	183	
91/09/19			4		26.5	7.6	92.75	8.40	184	
91/09/19			- 5		26.4	7.6	92.75	8.10	183	
91/09/19 91/09/19			5.5		26.4	6.5	79.38	8.10	185	
91/09/19					26.3 26.2	6.5	79.35	8.10	185	
91/09/19			9.5		26.1	6.1 6.0	74.45 73.25	7.90 7.80	185	
91/09/19			10		25.8	4.5	54.98	7.70	186	
91/09/19			iž		25.5	3.8	45.28	7.60	186	
									100	

INDEX 1021500 007720 00920 MILES 0953.80 0046.50 531.00

SHK 00060 00010 00300 00301 00400 00094 CNDUCTAT FIELD MICROMHO TIME OR DEPTH STREAM WATER 00 00 - PH FLOW OF DAY HEDIUM TEMP SATUR PERCENT MG/L su (8) /19 1227 WATER /19 1229 WATER /19 1231 WATER /19 1233 WATER /19 1235 WATER /19 1235 WATER /19 1242 WATER 25.5 25.2 25.2 25.0 25.0 25.0 25.0 14 3.8 45.28 7.50 187 16 18 20 22 24 26 3.6 3.6 3.4 3.3 42.98 42.98 40.58 39.38 31.08 26.28 7.40 7.40 7.30 7.40 7.30 7.40 187 187 188 188 188 188 2.6

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RIEVAL DATE 92/05/16 STORET 1089 475317 35 38 10.0 084 47 06.0 2 OPP. LOWE BR. WATTS BAR RES. MEIGS 47121 TENNESSEE TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 531.0 06010201002 0002.040 ON 131TVAC 0000 METERS DEPTH

INDEX 1021500 007720 00920 MILES 0953.80 0046.50 531.00

MEDIUM

TIME

0 F

DAY

DATE

FROM

10

PGM=RET

00078

TRANSP

SECCHI

METERS

84002

CODE

GENERAL

REMARKS

SHK

OR

DEPTH

(M)



31616

FEC COLL

MFM-FCBR

/100ML

/TYPA/AMBNT/FISH/STREAM/SOLIDS

00081

AP COLOR

PT-CO

UNITS

00080

COLOR

PT-CO

UNITS

82079

TURBIDTY

LAB

NTU

00630

NOZ&NO3

N-TOTAL

MG/L

.51

.49

. 31

.31

. 31

. 38

.37

. 39

. 14

. 33

.01

.41

. 03

: 34

.03

. 25

.08

. 32

00610

NH3+NH4-

N TOTAL

MG/L

.020

.030

.030

.020

.040

.090

.090

.120

.010K

.060

.010K

.010K

.050

.020

.070

.020

.040

2.00

32218

PHEOPHIN

A

UG/L

.050

00605

ORG N

N

MG/L

.080

.030

.110

.240

.200

.110

. 110

.100

.280

. 220

.200

. 110

.360

.170

.440

.250

.420

. 220

1.00

00530

RESIDUE

TOT NELT

MG/L

10K 1.32 4 0.3 4.0 91/01/31 1254 WATER 10 15 4 4.0 91/01/31 1257 VERT - 4 20 15 91/01/31 1309 WATER 22.1 10 . 1.26 D 1 91/04/18 1238 WATER 0.3 - 10K D 2 0.3 91/04/18 1239 WATER 10 4 D 3 2.0 0.3 5 91/04/18 1240 WATER 2 3 D 1 91/04/18 1243 VERT 4 1.0 2 5 4 D 2 4 2.0 5 91/04/18 1244 VERT 2 10 91/04/18 1245 VERT D 3 4 6.0 2 -5 9 D 1 6.0 91/04/18 1250 WATER 91/04/18 1251 WATER 23 5 2 5 23 D 2 5 6.0 2 03 23 91/04/18 1252 WATER 10K 1.70 2 4.0 91/05/16 1503 WATER 0.3 9 9.0 91/05/16 1507 VERT 4 91/05/16 1523 WATER 22.7 10K 1.65 4 2.0 0.3 15 91/06/19 0959 WATER 15 15 10.0 91/06/19 1001 VERT - 4 6.0 25 22 91/06/19 1014 WATER 10K 1.73 2 2.0 0.5 20 91/07/24 1120 WATER 5 9 7.0 4 60 91/07/24 1126 VERT 15 23.1 91/07/24 1142 WATER 10K 1.30 3 2.0 0.5 20 91/08/21 1127 WATER 5 7 5.0 25 91/08/21 1133 VERT 20 91/08/21 1152 WATER 23 10K 1.60 3 0.5 20 2.0 15 91/09/19 1213 WATER 5 5.0 6 30 91/09/19 1218 VERT 10 91/09/19 1237 WATER 23.5 32214 32211 32212 00680 00681 00671 00665 CHLRPHYL CHLRPHYL SHK 84002 CHLRPHYL PHOS-DIS T ORG C D ORG C PHOS-TOT 0.8 CODE В C A UG/L TIME С DATE ORTHO С GENERAL UG∕L DEPTH MG/L CORRECTO UG/L OF FROM MG/L MG/L P MG/L P REMARKS MEDIUM (M) DAY 10 1.00K 6.00 2.0 1.9 .020 .010 91/01/31 1257 VERT 1.8 1.8 .010 .020 91/01/31 1309 WATER 22.1 01 91/04/18 1238 WATER 0.3

D 2 91/04/18 1239 WATER 0.3 1.00K 2.00 1.00 D 3 10.00 91/04/18 1240 WATER 91/04/18 1243 VERT 0.3 1.7 2.1 .004 1.00 .010 1.00 D 1 1.00 9.00 - 4 1.8 .004 2.1 1.00K .010 1.00 D 2 1.00K 4 9.00 91/04/18 1244 VERT 2.1 1.7 .004 .010 03 91/04/18 1245 VERT 4 1.6 .007 1.8 .020 91/04/18 1250 WATER 91/04/18 1251 WATER D 1 23 1.5 1.7 .030 .007 23 DZ 1.5 .008 1.7 1.00 .030 1.00K 1.00K 23 D 3 10.00 1.8 91/04/18 1252 WATER 2.0 .002 .009 4 91/05/16 1507 VERT 1.6 1.7 .009 .020 2.00 1.00 2.00 91/05/16 1523 WATER 22.7 17.00 2.0 2.2 .002K .010 91/06/19 1001 VERT 1.6 1.9 .010 1.00 .030 2.00 1..00 91/06/19 1014 WATER 22 14.00 2.0 2.4 .004 .020 ۷. 1.7 1.9 91/07/24 1126 VERT .010 .020 2.00 1.00 1.00 19.00 91/07/24 1142 WATER 23.1 1.9 .007 2.1 .030 91/08/21 1133 VERT 1.7 .010 1.8 .030 2.00 1.00 1.00 23 12.00 91/08/21 1152 WATER 2.0 2.3 .009 .030 91/09/19 1218 VERT 4 1 0 2 1 020 070 01/00/10 1737 HATED 73 5

STORET RETRIEVAL DATE 92/05/16 476041 1114C 35 49 50.0 084 36 35.0 2 WATTS BAR RESERVOIR 47145 TENNESSEE ROANE 1ENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 560.80 1311VAC HO 0601 0000 METERS DEPTN

040801 0 Ng 06010201002 0043.170 OFF

PGN=RET

/TYPA/AMBNY/STREAN/SOLIDS

	0000 821												•						
	INDEX 10 MILES DP	21500 53.80 (007720	00920 60.80 SMK	00060	00010	00300	00301	00400 PH	00094 CNDUCTVV	MILES OF DATE	21500 00772 53.80 0046.5 TIME OF	0 00920 0 560.80 SMK OR DEPTH	00060 S1REAM FLOW	00010 UATER TEMP	00300	00301 D0 SATUR PERCENT	00400 PH SU	00094 CNDUCIVT Field / Micronho
	DATE FROM TO	TIME OF DAT	NEDIUM	OR DEPTH (M)	STREAM FLOW CFS	WATER TEMP CENT	DO NG/L	SATUR PERCENT	SU 7,70	FIELD NICRONHO 206	FROM TO 91/07/24	DAY MEDIU 1018 WATER		CFS	CENT 27.3 26.5	НС/L 9.4 8.6	116.15 104.95 98.85	8.50 8.30 8.10	191 193 194
	91/01/3 91/01/3 91/01/3	1 1144	WATER WATER	0.5 1 1.5		7.7 7.7 7.7 7.7	11.4 11.3 11.2 11.2	95.85 95.05 94.15 94.15	7.70 7.80 7.80 7.80	207 207 207 208	91/07/24 91/07/24 91/07/24	1019 WATER 1020 WATER 1021 WATER 1025 WATER	3		26.0 25.2 24.4 24.0	8.1 7.5 6.7 6.5	89.35 78.85 76.55	7.70 7.70 7.70 7.70	194 194 192 194
	91/01/3 91/01/3 91/01/3 91/01/3	1 1151 1 1155 1 1200	WATER WATER WATER	6 8 10		7.7 7.7 7.6 7.6	11.2 11.1 11.3 11.1	94.15 93.35 93.35 93.35	7.70 7.70 7.70 8.20	206 206 206 126	91/07/24 91/07/24 91/07/24	1027 WATER 1029 WATER 1033 WATER 1035 WATER	10 12 13 0.5		23.8 23.5 23.5 24.5	6.4 6.1 5.9 7.2	75.35 70.15 67.85 86.75	7.60 7.60 7.70 7.70	194 195 180 179
	91/01/3 91/04/1 91/04/1 91/04/1	8 1049 8 1052 8 1052	WATER WATER WATER	0.3 1.5		17.9 17.3 16.6 16.4	11.7 10.8 10.3 10.1	123.25 111.35 106.25 101.05	7.90 7.70 7.50 7.40	130 135 139 143	91/08/2 91/08/2 91/08/2	1 1010 WATER 1 1012 WATER 1 1014 WATER 1 1016 WATER	1.5		26.6 26.6 26.6 26.3	7.0 6.9 6.6	82.45 81.25 80.05 77.65	7.70 7.70 7.70 7.60	180 181 181 181
	91/04/1 91/04/1 91/04/1 91/04/1	8 1056 8 1057 8 1058	WATER WATER WATER	6 8 10 12		16.3 16.0 15.8 15.8	9.9 9.7 9.4 9.2	99.05 97.05 94.05 92.05	7.30 7.20 7.20 7.20	169	91/08/2 91/08/2 91/08/2	1 1018 WATER 1 1020 WATER 1 1022 WATER 1 1030 WATER	8 10 12		24.3 24.3 24.3 24.3	6.6 6.5 6.5	77.68 76.58 76.58 75.38	7.60 7.60 7.60 7.60 7.80	181 181 179 193
)	91/04/1 91/04/1 91/05/1 91/05/1	<pre>8 1103 6 1342 6 1343</pre>	WATER WATER WATER	13 0.3 1.5		15.7 23.4 22.6 22.0	8.8 9.8 9.9 9.6	88.05 112.65 113.85 109.15	8,20 8,20 8,00 7,60	152 151 148	91/09/1 91/09/1 91/09/1	1 1035 WATER 9 1110 WATER 9 1113 WATER 9 1113 WATER 9 1114 WATER	0.5 1 1.5		24.9 24.9 24.9 24.9 24.9	6.6 6.6 6.5	78.65 78.65 78.65 77.45	7,00 7,70 7,80 7,60	193 192 193
	91/05/1 91/05/1 91/05/1 91/05/1	6 1347 6 1349 6 1351	WATER WATER WATER	3.1	÷	20.8 20.4 19.9 19.7	8.0 7.6 7.4 7.2	88.95 82.65 80.45 78.35	7.50 7.40 7.40 7.40	142 140 139	91/09/1 91/09/1 91/09/1	9 1116 WATER 9 1120 WATER 9 1122 WATER 9 1124 WATER	8 8 10		24.9 24.9 24.9 24.8	6.3 6.3 6.2 6.1	73.85	7.60 7.70 7.60	194 193 195
	91/05/ 91/05/ 91/05/ 91/05/	16 1356 16 1400 16 1405	WATER VATER VATER	10 12 13		19.6 19.6 19.6 26.5	7.1 7.0 7.0 11.1	76.18	7.40	140 140 177	91/09/1 91/09/1	9 1126 WATE 19 1132 WATE			24.7	5.7	67.98	7.70	, ,,,
	91/06/ 91/06/ 91/06/ 91/06/	19 0848 19 0849 19 0850 19 0851	WATER WATER WATER WATER	0.3		26.1 26.0 25.5 24.7	9.8 9.6 8.8 7.9	117.18 104.88 94.05	8.6 8.3 8.0	0 175 0 172 0 168									·
	91/06/ 91/06/ 91/06/	19 0854 19 0855 19 0855 19 0855	UATER UATER S VATER S VATER	4.8 6 8 10		23.1 22.4 22.2 22.2	5.4 5.7 5.0	59.18 56.88 55.78	7.6 7.4 7.4 7.4	0 161 0 161 0 161									
	91/06/ 91/06/	19 0850 19 0851 24 101	6 WATER 9 WATER 5 WATER 7 WATER	12 12.5 0.5 1		22.2 28.4 27.9	4. 10. 9.	4 131.65		0 185									,
	,,,,,,																		

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PGM=RET

SIURE REIRIEVAL DAIE 92/05/16 47 35 40.0 084 36 33.0 2 WATTS BAR RESERVOIR 47145 TENNESSEE ROANE TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 560.80 1311VAC HQ 060

HQ 06010201002 0043.170 OFF

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/TYPA/AMBNT/STREAM/SOLIDS

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INDEX 1021500 007720 00920 MILES 0953.80 0046.50 560.80

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TIME OF DAY	MEDIUM	SMK OR DEPTH (M)	84002 CODE GENERAL REMARKS	00078 TRANSP SECCHI METERS	31616 FEC COLI MFM-FCBR /100ML	00080 COLOR PT - CO UNITS	00081 AP COLOR PT-CO UNITS	82079 TURBIDTY ŁAB NTU	00530 RESIDUE TOT NFLT MG/L	00605 0rg n n Mg/l	00610 NH3+NH4- N TOTAL MG/L	00630 N028N03 N-TOTAL MG/L
1148 1205 1049	VERT WATER WATER	0.5 4 10.6 0.3	D 1	1.00 .75	10 10K	10 10	15 15	6.0 5.0	7 8	.060 .060	.060 .040	.61 .60
1051 1053 1054 1055 1055	WATER VERT VERT VERT WATER	0.3 4 4 12.1	D 3 D 1 D 2 D 3 D 1 D 2		10	22222	5 5 5 5	5.0 5.0 7.0 6.0 6.0	7 7 8 9	.120 .110 .120 .030 .090	.040 .030 .020 .050 .070	. 30 . 31 . 31 . 40 . 41 . 42
1102 1342 1349	WATER WATER VERT	12.1 0.3 4 12.2	D 3	1.22		-	5	6.0 6.0 10.0	8 5 10	. 290 . 230	.010 .050	. 28 . 41
2 0848 2 0851 2 0856	WATER VERT WATER	0.3 4 12 0.5		1.00		20 35	20 35	4.0 13.0	6 15	. 400 . 220	.020	. 4 1
1021 1034 1010	VERT WATER WATER	4 12.1 0.5 4			10K	10 15	25 20	6.0 4.0	7 5	. 160 . 330	.060	. 22
1025 1110 1116	WATER WATER VERT	11.9 0.5 4 12.3		1.20	10K		35 20 25	7.0 4.0 5.0	5 7	. 320	.020 .020 .040	. 28
TIHE OF DAY	HEDIUM	SMK DR DEPTH (M)	84002 CODE GENERAL REMARKS	00665 Рнос-тот Mg/l р	00671 PHOS-DIS ORTHO MG/L P	00680 T ORG C C MG/L	00681 D ORG C C Mg/L	32211 CHLRPHYL A UG/L CORRECTD	32212 CHLRPHYL B UG/L	32214 CHLRPHYL C UG/L	32218 PHEOPHIN A UG/L	
1205 1049	WATER WATER	4 10.6 0.3 0.3	D 1 D 2	.030 .040	.020 .020	2.0 1.9	1.9 1.9	5.00	1.00	1.00	2.00	
3 1051 3 1053 3 1054 3 1055 3 1055 3 1100	WATER VERT VERT VERT WATER	0.3 4 4 12.1	D 3 D 1 D 2 D 3 D 1 D 2	.020 .020 .020 .020 .020 .020	.005 .006 .006 .009 .009	1.9 1.7 1.6 1.5 1.4	1.6 1.5 1.5 1.4 1.3	6.00 6.00 5.00	1.00K	1.00 1.00 1.00	1.00K 1.00 1.00	
3 1102 5 1349 5 1401	WATER VERT WATER	12.1	D 3	.020 .030 .030	.004 .009	2.1 1.9	1.8 1.8	9.00 9.00		1.00		
0856	WATER	4 12 4		.030 .040 .020	.007	2.1	1.8 1.8	13.00		1.00	2.00	
4 1034 1 1016	WATER VERT	12.1		.020	.007	1.9	1.8	9.00	1.00	1.00	3.00	
9 1116	VERT	11.9 4 12.3		.040 .030 .030	.010 .010 .010	2.2	2.0	8.00	1.00	1.00	2.00	
	1148 OF 1148 1050 1053 1053 1053 1053 1053 1053 1053 1053 1053 1053 1053 1053 1053 1053 10102 10102 10102 10102 10102 10102 10102 10102 10102 10102 10102 10102 10102 10102 11116 11116 10102 111102 111102 111102 11102 11102 111102 111102 111102 11102 111102 11102 11102 111102 111102 111102 111102	11 ME OF DAY MEDIUM 1145 WATER 1148 VERT 1205 WATER 1050 WATER 1051 WATER 1053 VERT 1054 VERT 1055 VERT 1105 WATER 1105 VERT 1105 WATER 1105 WATER 1101 WATER 1342 WATER 1401 WATER 0856 WATER 0856 WATER 1021 VERT 1021 VERT 1034 WATER 1021 VERT 1021 VERT 1021 VERT 1021 VERT 1025 WATER 1021 VERT 1025 WATER 1100 WATER 1110 WATER 1110 WATER 1110 WATER 1110 WATER	11ME OR DEPTH DAY MEDIUM (M) 1145 WATER 0.5 1148 VERT 4 1205 WATER 0.3 1050 WATER 0.3 1051 WATER 0.3 1055 VERT 4 1101 WATER 12.1 1102 WATER 12.1 1102 WATER 12.1 1104 WATER 12.1 1105 WATER 12.1 1102 WATER 12.1 1104 WATER 12.1 1105 WATER 12.2 0856 WATER 12.2 1015 WATER 12.1 1025 WATER 1.9 1010 WATER 12.1 1025 WATER 1.9 11101 WATER 1.9 <td>SMK B4002 OR OF DEPTH GENERAL GENERAL DAY HEDIUH (M) REMARKS 1145 WATER 0.5 GENERAL 1145 WATER 0.5 GENERAL 1145 WATER 0.3 D1 1050 WATER 0.3 D2 1051 WATER 0.3 D3 1055 VERT 4 D1 1054 VERT 4 D2 1055 VERT 4 D3 1100 WATER 12.1 D1 1101 WATER 12.1 D3 1102 WATER 12.1 D3 1342 WATER 0.3 D3 0851 VERT 4 D3 0856 WATER 12.2 D848 0856 WATER 0.5 CDE 1015 WATER 12.3 CDE 1014 VERT 4 D2</td> <td>TIME SHK 84002 00078 OF DEPTH GENERAL SECCHI DAY HEDIUH (M) REMARKS METERS 1145 WATER 0.5 1.00 1148 VERT 4 1.00 1205 WATER 10.6 1.00 1050 WATER 0.3 D1 .75 1050 WATER 0.3 D2 .75 1050 WATER 1.10 4 D2 1055 VERT 4 D1 .75 1050 WATER 1.21 D3 </td> <td>SHK 84002 OF 00078 DEPTH 31616 FEC COLI GENERAL GENERAL SECCHI MFM-FCREN 31616 FEC COLI MFM-FCR MFTFS 1145 WATER 0.5 1.00 10 1145 WATER 0.5 1.00 10 1145 WATER 0.5 1.00 10 1145 WATER 0.3 D1 .75 10K 1049 WATER 0.3 D2 .00 10 1053 VERT 4 D2 .00 10 1055 VERT 4 D2 .00 10 1055 VERT 4 D2 .00 10K 1054 WATER 12.1 D2 .00 10K 1105 WATER 12.1 D3 1.22 10K 1322 WATER 1.20 10K .00 10K 1054 WATER 12.1 D3 1.20 10K 1055 VERT 4 .00 .00 .00</td> <td>TIME OF DAY SMK HEDIUM 84002 CODE CODE (M) 00078 TRAMSP ECCHI METERS 31616 FEC COLI MEM-FCBI /100ML 00080 PT.CO PT.CO PT.CO UNITS 1145 WATER 0.5 1.00 10 1146 VERT 4 10 10 1146 VERT 4 10 10 1146 VERT 4 10 10 1050 WATER 0.3 D1 .75 10K 1051 VERT 4 D2 2 2 1055 VERT 4 D2 2 2 1054 VERT 4 D2 2 2 1055 VERT 4 D2 2 2 1101 VATER 12.1 D3 1.22 10K 2 1104 VATER 0.3 1.20 10K 20 0851 VERT 4 20 35 1.46 10K 1010 VATER 0.5 1.46 10K</td> <td>TIME SMK 84002 00078 31616 00080 00081 DAY HEDIUM (H) REMARKS TRAMSP FEC COLI COUR DOUBD AP COLOR 1145 WATER 0.5 1.00 10 UNITS UNITS 1145 WATER 0.6 10 10 15 1205 WATER 0.3 D1 .75 10K 100 15 1051 WATER 0.3 D2 10X 2 5 5 1055 VERT 4 D1 2 5 5 1055 VERT 4 D2 2 5 5 1010 WATER 12.1 D2 2 5 5 1104 WATER 12.2 D3 1.22 10K 5 5 1104 WATER 12.1 D3 1.22 10K 5 20 0846 WATER 12.1 D3<</td> <td>IIME OF DAY HEDIUM SHK OF PT HEDIUM 84.002 CODE FEC COLI DEPTH DEPTH DEPTH DEPTH DEPTH DEPTH DEPTH DEPTH DEPTH DEPTH DEPTH DEPTH DEPTH 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Resource Group River Basin Operations Water Resources

RESERVOIR VITAL SIGNS MONITORING - 1992 PHYSICAL AND CHEMICAL CHARACTERISTICS OF WATER AND SEDIMENT

Prepared by Joeeph P. Fehring and Dennis L. Meinert

Chattanooga, Tennessee October 1993

INTRODUCTION

The Tennessee Valley Authority (TVA) initiated a Reservoir Monitoring Program in 1990 as part of its Water Resources and Ecological Monitoring Activities. In these first three years (1990-1992), the Reservoir Monitoring Program has undergone continual change and improvement. Initially, in 1990, only 12 TVA reservoirs were examined, the nine mainstream Tennessee River reservoirs (Kentucky through Fort Loudoun) and three major tributary storage reservoirs Cherokee, Douglas, and Norris (Dycus and Meinert, 1991). In 1991, the Reservoir Monitoring Program was expanded to 24 reservoirs, to include the system's only two tributary reservoirs with navigation locks (Tellico and Melton Hill) and ten other smaller tributary reservoirs (Dycus and Meinert, 1992).

The two objectives of the Reservoir Monitoring Program are to provide basic information on the "health" or integrity of the aquatic ecosystem in TVA reservoirs (referred to as Vital Signs Monitoring) and to provide screening level information for describing how well each reservoir meets the "fishable" and "swimmable" goals of the Clean Water Act (referred to as Use Suitability Monitoring).

The basis of Vital Signs Monitoring is examination of appropriate physical, chemical, and biological indicators monitoring tools - at one or more strategic locations in each reservoir, i.e. the forebay immediately upstream of the dam; the transition zone (the mid-reservoir region where the water changes from free flowing to more quiescent, impounded water; and the inflow or headwater region of the reservoir. The monitoring tools comprised: basic physical/chemical water quality sampling; sediment quality and toxicity testing; benthic macroinvertebrate community evaluations; and fish community evaluations. A summary report (Meinert and Dycus, 1993) presents results of TVA's 1992 Reservoir Monitoring Program, to include all Vital Signs (physical/chemical, sediment, benthic and fish communities) and

Use Sutiability Monitoring.

This technical data summary presents the 1992 Vital Signs physical/chemical water quality sampling information only. The basic group of 24 reservoirs were again monitored, however, several sampling location changes were made prior to commencement of 1992 sampling activities. Transition zone sampling locations were relocated further downstream to less riverine environs on Kentucky, Wheeler, and Guntersville reservoirs. Due to the similarities in water quality between forebay and transition zone the transition zone sampling was dropped on Nickajack reservoir. Forebay sampling locations on Chatuge and Nottely reservoirs were relocated slightly downstream and mid-reservoir sampling locations were added to better define ecological conditions on these two reservoirs.

MATERIALS AND METHODS

Reservoir Characteristics - The physical characteristics of a reservoir (volume, surface area, depth, hydraulic residence time, etc.) have a great effect on its intrinsic physical and chemical processes and water quality characteristics. The Vital Signs reservoirs are very broadly categorized as either run-ofthe-river or tributary reservoirs, with generally large differences in their morphologic and hydraulic characteristics (Table 1). Primary differences include the greater depths and longer hydraulic residence times of tributary reservoirs, resulting in the development of strong thermal stratification and its associated physical/chemical processes. Short average residence times found on most of the main stream, run-of-theriver reservoirs (usually less than 30 days) results in wellmixed riverine conditions. Thermal stratification of the run-ofthe-river reservoirs rarely exists and then for only short periods of time under conditions of low flow and intense solar heating. However, tributary reservoirs with their longer residence times (typically greater than 100 days) develop strong thermal stratification that begins in the spring and ends with seasonal cooling and mixing in the late autumn. During this summer stratification, inflowing water may be cooler than the epilimnetic surface water and may enter the reservoir as a cold underflow to the hypolimnion rather than mixing with the epilimnion. This results in an even longer residence time of the epilimnetic water than is indicated by the calculated average reservoir volume/flow quotient, particularly if the turbine power intakes draw water exclusively from below the thermocline.

A major effect of this thermal stratification in tributary reservoirs is the depletion of hypolimnetic dissolved oxygen. The pattern of hypolimnetic dissolved oxygen depletion is variable both spatially and temporally for any one reservoir and among reservoirs. The general pattern, however, is that after the onset of thermal stratification, an anoxic zone first begins

Table 1

CHARACTERISTICS OF VITAL SIGNS RESERVOIRS

Reservoir	ainage Area . miles)	Length ^a (miles)	Surface Area [®] (acres) 1000's	Depth at Dam [*] (ft)	Volume ^a <u>(ac-ft)</u> 1000's	Average Reservoir Flow 1960-92 (cfs)	Average Hydraulic Residence Time-1992* (Days)	CY 1992 Reservoir Flow _(cfs)_	
		Run-of	f-the-Rive		oirs				
Kentucky	40,200	184.0	160.3	88	2,839	63,182	28.6	49,960	
Pickwick	32,820	53.0	43.1	84	924	56,505	9.3	50,057	
Wilson	30,750	15.5	15.5	108	634	53,305	6.5	48,840	
Wheeler	29,590	74.0	67.1	66	1,050	50,956	11.1	47,838	
Guntersville	24,450	76.0	67.9	65	1,018	41,562	12.5	41,086	
Nickajack	21,870	46.0	10.4	60	241	35,593	3.5	35,122	
Chickamauga	20,790	59.0	35.4	83	628	34,174	9.4	33,608	
	17,300	72.0/24.0 ^b	39.0	105	1,010	27,788	19.7 9.3	25,846	
Fort Loudoun	9,550	50.0	14.6	94	363	15,742	33°	19,664 6,301°	
Tellico	2,627	33.2	16.5	80	415	6,365°	16.5	3,670	
Melton Hill	3,343	44.0	5.7	69	120	4,424	10.5	5,070	
 1		Tribu	tary Rive	r Reserv	oirs				
Norris	2,912	73.0/53.0	34.2	202	2,040	4,070	325.7	3,158	
Cherokee	3,428	54.0	30.3	163	1,481	4,529	177.1	4,215	
Douglas	4,541	43.0	30.4	127	1,408	6,879	96.0	7,398	•
Boone	1,840	17.4/15.3 ^b	4.3	129	189	2,510	37.3	2,555	
South Holston	703	24.0	7.6	239	658	983	346.3	958	
Watauga	468	16.0	6.4	274	569	700	351.6	816	
Hiwassee	968	22.0	6.1	255	422	2,077	95.2	2,234	
Chatuge	189	13.0	7.0	124	234	453	241.8	488	
Nottely	214	20.0	4.2	167	170	405	180.4	478	
Blue Ridge	232	11.0	3.3	156	193	614	155.4	626	
Ocoee #1 (Parksville)	595	7.5	1.9	115	85	1,415	30.6	1,400	
Tims Ford	529	34.0	10.6	143	530	967	358.7	745	
Normandy	195	17.0	3.2	83	110	343	161.7		
Beech	16	5.3	0.9	32	11	14 ^d	280 ^d		

Measurements based on normal maximum pool and average flows. a.

Major/minor arms of reservoir.

c. Estimated based on releases from Chilhowee Dam, and adjusted based on the additional drainage area between Tellico (2627 sq miles) and Chilhowee (1977 sq miles) dams.

d. Estimated

to develop in the transition zone of the reservoir. The large input and settling out of allochthonous organic matter (leaves and detritus washed in by the tributary streams) causes high rates of oxygen uptake in the transition zone and results in the initial depletion of dissolved oxygen in this area of the reservoir. The zone of anoxic water gradually extends both upstream and downstream, until it reaches the free-flowing river upstream and the dam downstream. At the same time the anoxic zone develops vertically and laterally. In the worst case, the entire hypolimnion may become anoxic and remain so until reaeration during autumn overturn. The hypolimnetic anoxia promotes the dissolution and release of metals, minerals, and nutrients from the reservoir sediment and bottom water, resulting in elevated concentrations of reduced forms of iron, manganese, sulfate, nitrate, and phosphate. When reservoir destratification occurs in autumn, the hypolimnion is mixed with surface water and replenished with oxygen.

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Epilimnetic processes are also affected by thermal stratification, particularly in deeper tributary reservoirs. With the onset of spring stratification, primary production can be quite high, particularily in the transition zone. The phytoplankton community frequently receives amples supplies of nutrients from periodic spring rainfall/runoff events, and with lower water velocities and increased rates of sedimentation, improved water clarity and light penetration often results in episodes of high algal productivity as evidenced by high chlorophyll-a concentrations, high pH values and supersaturation of dissolved oxygen. The longer residence times of tributary reservoirs results in water clarity usually being more a function of algal cell abundance and chlorophyll pigment and less a function of inorganic suspended material as is the case in mainstream reservoirs with shorter residence times and higher In mainstream reservoirs, the shorter detention velocities. times and higher velocities inhibit the deposition of suspended material, resulting in generally lower water clarity and light

availability and often limiting algal productivity.

Epilimnetic water clarity often increases moving downstream into the forebay area of the reservoir and into the late summer as rainfall/runoff events occur less frequently. Even though water clarity increases, nutrients become depleted and primary production often is reduced as much of the organic matter produced during photosynthesis decomposes and settles below the photic zone. Bacterial decomposition consumes available dissolved oxygen (respiration) and releases carbon dioxide, thereby lowering the pH. Unlike mainstream reservoirs, this decomposition process in tributary reservoirs often results in two oxygen minima in the water column - one in the epilimnion at or near the thermocline and one in the hypolimnion.

Consequently, physical/chemical differences in water quality between run-of-the-river and tributary reservoirs are expected in examining the 1992 Vital Signs data. In the Results and Discussion section of this report, water quality characteristics are discussed and comparisons are made among the Vital Signs reservoirs to point out these differences. Because physical/chemical water quality conditions are different, this does not imply that one reservoir is good and another is poor, it merely is a reflection of the unique physical and chemical differences between each. For example, one particular reservoir, Fort Loudoun, exhibits both run-of-the-river and tributary reservoir characteristics. Although it is located on the main stream of the Tennessee River, it is the most upstream reservoir and is formed by the confluence of the French Broad and Holston Major tributary reservoir impoundments on both these Rivers. river systems, Douglas Dam at mile 32.3 of the French Broad and Cherokee Dam at mile 52.3 on the Holston, release cool, hypolimnetic water low in dissolved oxygen during the summer. Depending upon the quantity and duration of water released from Douglas and Cherokee dams and the temperature of the impounded Fort Loudoun water, this cool water may flow under the warmer water impounded in Fort Loudoun Reservoir resulting in thermally

stratified conditions. Consequently, Fort Loudoun can intermittently exhibit the characteristics of either a run-of-the-river reservoir or a tributary reservoir.

Overview - Physical/chemical variables were measured at a total of forty seven locations on the twenty four Vital Signs reservoirs (Figure 1, Table 2). The Vital Signs water quality monitoring activities on these reservoirs, followed either a "basic" or "limited" sampling strategy (Tables 2 and 3). The basic sampling strategy included monthly water quality surveys (January and April through September) at the forebay and transition zone locations on fourteen TVA reservoirs: the nine mainstem Tennessee River reservoirs; and Cherokee, Douglas, Norris, Melton Hill and Tellico reservoirs. The limited sampling strategy included monthly water sampling (April through October) for a smaller list of parameters at the forebay locations (and at mid-reservoir locations on larger reservoirs) on ten nonnavigable tributary impoundments.

Water quality measurements, sample collections and sample handling followed standard practices accepted by the Environmental Protection Agency (EPA, 1979 and 40 CFR 136), US Geological Survey (USGS, 1977), and American Public Health Association (AWWA, WPCF, APHA, 1989) as specified in TVA quality assurance/quality control (QA/QC) procedures (TVA, 1990 and TVA, 1987). TVA laboratory analyses conformed to established EPA, USGS, and APHA QA/QC procedures (TVA, 1989b).

Details on the physical/chemical analyses and measurements on the water samples are summarized in Tables 3 and 4. The 1992 data are tabulated in the Appendix A - Physical/Chemical Characteristics of Water. All the data are stored and are available on EPA's water quality data storage and retrieval (STORET) computer system.

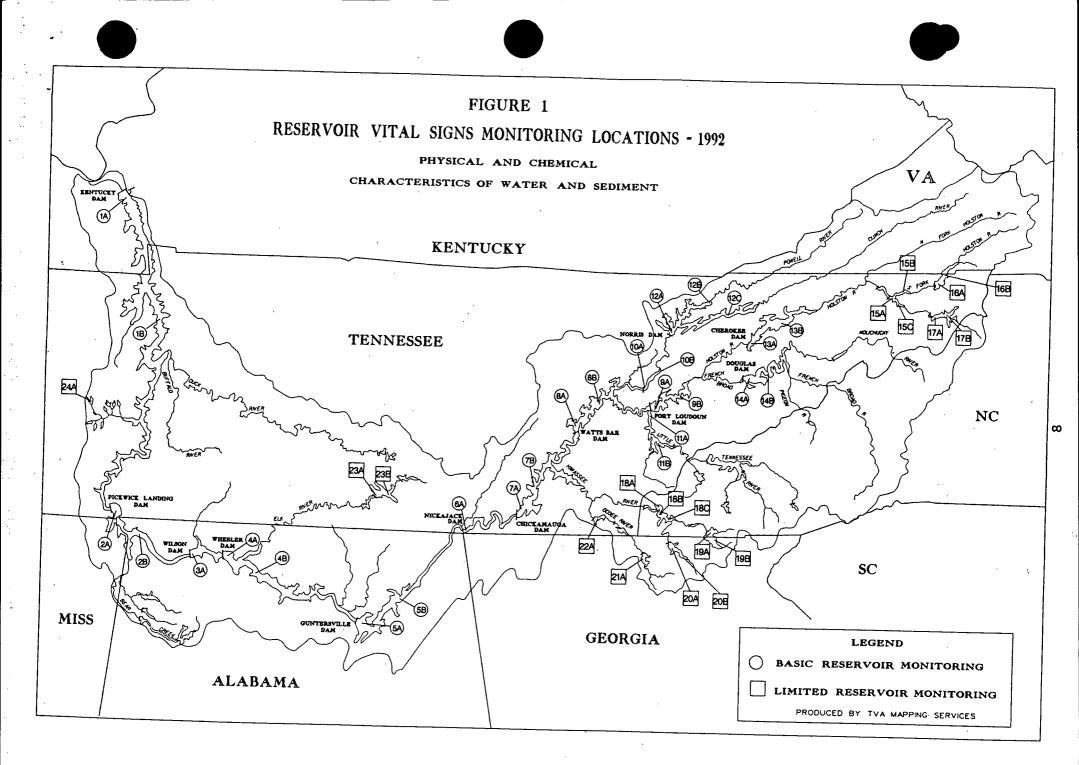


Table 2

WATER QUALITY MONITORING LOCATIONS RESERVOIR VITAL SIGNS MONITORING, 1992

Basic Water Quality Monitoring Locations

	Foreb	ay Locat	ions	Transition Zone Locations					
		Map ID	Storet		Map ID	Storet			
Reservoir	<u>River Mile</u>	Number	<u>Station #</u>	<u>River Mile</u>	<u>Number</u>	<u>Station #</u>			
	(Tennessee)			(Tennessee)					
	F	Run-of-th	e-River Rese	ervoirs					
					. –				
Kentucky	23.0	1A.	202832	85.0	1B.	477403			
Pickwick	207.3	2A.	476799	230.0	2B.	016923			
Wilson	260.8	3A.	016912						
Wheeler	277.0	4A.	016900	295.9	4B.	017009			
Guntersville	350.0	5A.	017261	375.2	5B.	017522			
Nickajack	425.5	6A.	476344						
Chickamauga	472.3	7A.	475358	490.5	7B.	475265			
Watts Bar	531.0	8A.	475317	560.8	8B.	476041			
Fort Loudoun	605.5	9A.	475602	624.6	9B.	475603			
	(603.2		477404)						
Melton Hill	CRM 24.0	10A.	477064	CRM 45.0	10B.	476194			
Tellico	LTRM 1.0	11A.	476260	LTRM 21.0	11B.	476295			
		• Tributa	ry Reservoi	rs					
Norris	CRM 80.0	12A.	476009	PRM 30.0	12B.	477187			
				CRM 125.0	12C.	477186			
Cherokee	HRM 53.0	13A.	475025	HRM 76.0	13B.	475028			
Douglas	FBRM 33.0	14A.	475081	FBRM 60.7	14B.	475993			
Dougras	I DIGI 33.0	T-117 •	475001		T17.				

Table 2 (continued)

WATER QUALITY MONITORING LOCATIONS RESERVOIR VITAL SIGNS MONITORING, 1992

Limited Water Quality Monitoring Locations

	Forebay_Locat		Mid-Rese	ervoir Locations
Reservoir	Map ID <u>River Mile</u> <u>Number</u>	Storet <u>Station #</u>	<u>River Mile</u>	Map ID Storet <u>Number Station #</u>
	Tr	ibutary		
Boone	SFHR 19.0 15A.	475858	SFHR 27.0	15B. 476221
South Holston Watauga Hiwassee Chatuge	SFHR 51.0 16A. WRM 37.4 17A. HIRM 77.0 18A. HIRM 121.1 19A.	475859 475576 370001 370059	WRM 8.3 SFHR 62.5 WRM 44.0 HiRM 85.0 HiRM 90.0 HiRM 125.6	15C.47599716B.47557317B.47557718B.37015418C.370163
Nottely Blue Ridge Ocoee #1 Tims Ford Beech	NRM21.520A.ToRM54.121A.ORM12.522A.ERM135.023A.	130073 130032 475684 477072	HiRM 125.6 NRM 31.0 ERM 150.0	19B. 130071 20B. 120806 23B. 475768
200011	BRM 36.0 24A.	475876		

Abbreviations: BRM - Beech River Mile CRM - Clinch River Mile ERM - Elk River Mile FBRM - French Broad River Mile HiRM - Hiwassee River Mile HRM - Holston River Mile LTRM - Little Tennessee River Mile

NRM - Nottely River Mile ORM - Ocoee River Mile PRM - Powell River Mile SFHR - South Fork Holston River Mile ToRM - Toccoa River Mile WRM - Watauga River Mile



Table 3

RESERVOIR VITAL SIGNS - 1992 WATER QUALITY MONITORING STRATEGY

Description	Sample Collection Depths	Monitoring <u>Basic</u>	Strategy ^b Limited
<u>Field Measurements</u> Hydrolab [®] (temperature, pH, dissolved oxygen, and conductivity)	0.3,1.5,4,6,8ª etc.	monthly	monthly
Secchi Depth		monthly	monthly
Fecal Coliform	0.3 (surface grab)	monthly	NA
<u>Laboratory Measurements</u> Chlorophyll-a	composite ^c	monthly	monthly
Nutrients Organic Nitrogen Ammonia Nitrogen Nitrite & Nitrate Nitrogen Total Phosphorus Dissolved Ortho phosphorus	composite (& bottom) ^d composite (& bottom) composite (& bottom) composite (& bottom) composite (& bottom)	monthly Ap monthly Ap monthly Ap	ril & August ril & August ril & August ril & August ril & August
Organic Carbon Total Organic Carbon	composite (& bottom)	monthly Ap	ril & August
Color and Solids Color Suspended Solids	composite (& bottom) composite (& bottom)	monthly monthly	NA NA

In situ Hydrolab measurements were made at the depths indicated and at regularly spaced intervals (2-4 meters) from the water surface to the bottom of the water column. These measurements were also made at intermediate depths any time the temperature changed by more than 2°C or the dissolved oxygen changed by more than 1 mg/l.

^b Basic monthly is January and April through September. Limited monthly is April through October.

^c Composite indicates a photic zone composite sample with the photic zone defined as four meters or twice the Secchi depth, whichever is greater.

^d Bottom grab samples were only collected as part of the basic sampling strategy. Bottom indicates a grab sample collected three meters above the bottom at forebay locations and one meter above the bottom at transition zone locations.

Table 4

RESERVOIR VITAL SIGNS MONITORING - WATER PHYSICAL/CHEMICAL MEASUREMENTS, 1992

EPA Storet			Detection
<u>Code</u>	Description	Units	Limits
	Field Measure	ements	
	<u>ricia meddar</u>	<u> </u>	
00010	Hydrolab	٥C	
00010 00300	Temperature Dissolved Oxygen	mg/l	
00400	pH	Std. units	
00094	Conductivity	micromhos	
00078	Secchi Depth	meters	0.1 meters
31616	Fecal Coliform	colonies/100mL	1/100 mL
	Laboratory Meas	urements	
32211	Chlorophyll-a	μg/l	1 µg/l
	Nutrients		0.00
00605 00610	Organic Nitrogen Ammonia Nitrogen	mg/l mg/l	0.02 mg/l 0.01 mg/l
00610	Nitrite & Nitrate Nitrogen		0.01 mg/l
00665	Total Phosphorus	mg/l	0.002 mg/l
00671	Dissolved Ortho phosphorus	mg/l	0.002 mg/l
	Our and a Comban		
00680	Organic Carbon Total Organic Carbon	mg/l	0.2 mg/l
		<i></i>	
	Color and Solids		
00080	Color	PCU	1 PCU
00530	Suspended Solids	mg/l	1 mg/l

<u>Quality Assurance/Quality Control</u> - The Reservoir Vital Signs Monitoring program includes three unique QA/QC measures. These are: (1.) collection of triplicate sets of samples at each reservoir sampling location; (2.) preparation of sample container (and/or field) blanks (defined below) each collection day; and, (3.) preparation of sample filtration blanks (defined below) with each set of filtered samples. These data in tabular form are given in the appendix.

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•Triplicates - On one occasion during the year, water samples from each reservoir were collected and analyzed in triplicate to assess sample collection, analysis, and natural variability. Triplicate data shown in Appendix A are identified by the remarks codes "D1", "D2", and "D3", respectively, for each individual measurement in a set of triplicates. Each individual water quality parameter has its own inherent naturally occurring variability, and different methods of analysis have varing degrees of precision and accuracy which are reflected in these data.

•Container Blanks - A container blank is defined as a sample container which has been filled with water taken directly from a Reagent Grade I water system without transference to any intermediary container. A field blank is defined as a sample container which has been filled with Reagent Grade I water which has been temporally stored in a transport carboy or large container for use in the field. A set of container (or field) blanks was collected by each survey party on each monthly survey. Container blanks sample bottles are filled and handled in the same manner and analyzed for the same variables as actual reservoir water quality samples. This assesses the degree of contamination associated with the sample bottles and/or the sample handling processes. The container blank information for the last three years (Table 5) show that true color and suspended solids have a very low frequency of detection, less than 3%. However, the results also indicate that contamination of samples

due to ammonia nitrogen is quite frequent. Approximately 16% of the "blank" ammonia nitrogen containers show detectable concentrations greater than or equal to twice the laboratory detection limit of 0.01 mg/l. An acceptable level of sample bottle contamination would be less than 10%.

Table 5 shows that total organic carbon also is frequently detected in blank samples, with approximately two-thirds of the container blanks showing possible evidence of contamination, in 1992. However, because of instrument variability (± 0.1 mg/l) at or near the instrument's operational detection limit of 0.2 mg/l, and the variable quality of reagent grade water used in the field, carbon values equal to or less than 0.4 mg/l (twice the detection limit) have been judged to be indicative of contaminate-free containers. Considering only total organic carbon values greater than 0.4 mg/l (twice the detection limit), about 10% of the organic carbon blank containers showed contamination in 1992.

The quality of sample containers, chemical preservatives, Reagent Grade I water, and field and laboratory handling all need to be examined to minimize and eliminate possible sources of contamination.

•Filtration Blanks - Filtration blanks were prepared with each set of water samples whenever dissolved phosphorus samples were collected. Filtration blanks were prepared in the field by filtering and handling Reagent Grade I water in exactly the same manner as the ambient reservoir water was handled and filtered. The filtration blank samples were analyzed in the same manner as ambient water quality samples to assess variability due to field sample filtration techniques. The data are summarized in Table 5 and show that approximately 32% of the dissolved ortho phosphorus filtration blank samples had detectable concentrations of phosphorus greater than the laboratory detection limit of 2 μ g/l. This was about twice the percentage of contamination for (unfiltered) dissolved phosphorus container blanks (15.6%), indicating contamination due to sample filtration and handling.

<u>Recommendations</u> - Because of the low frequency of contamination of container blanks for true color and suspended solids, collection of container blanks for these parameters should be discontinued in the 1993 Vital Signs Reservoir Monitoring Program. Continued attention needs to be given to elimination sources of ammonia nitrogen contamination and contamination introduced by the handling and filtration of dissolved phosphorus samples.

TABLE 5

QUALITY ASSURANCE/QUALITY CONTROL DATA SUMMARY RESERVOIR VITAL SIGNS MONITORING 1990-1992

	<u>Organic-N</u> (mg/l)	<u>Ammonia-N</u> (mg/l)	Nitrite + <u>Nitrate-N</u> (mg/l)	Total <u>Phosphorus</u> (µg/l)	Dissolved <u>Ortho-P</u> (µg/l)		True <u>Color</u> (PCU)	Suspended <u>Solids</u> (mg/l)	
Detection Limit (DL)	0.02	0.01	0.01	2	2	0.2	1	- 1	
1990		. !	CONTAINER	BLANK SAMPL	.ES				
Number of Samples % > DL % > 2xDL	70 27.1 5.7	70 34.3 15.7			30 3.3 0.0	73 72.4 21.9	70 1.4 1.4	70 0.0 0.0	
1991 Number of Samples % ≥ DL% % ≥ 2xDL	100 29.0 11.0	101 30.7 20.8			65 30.8 7.7	100 72.0 32.0	69 2.9 2.9	81 0.0 0.0	
1992 Number of Samples % Exceeding DL % ≥ 2xDL	92 30.4 3.3	94 48.9 16.0	94 6.4 3.3		90 15.6 4.4	93 66.7 10.8	9 0.0 0.0	9 0.0 0.0	
		Ē	ILTRATION	BLANK SAMP	LES				
1990 Number of Samples % ≥ DL % ≥ 2xDL					118 20.3 5.1	·			
1991 Number of Samples % ≥ DL% % ≥ 2xDL					153 26.8 9.2				
1992 Number of Samples % Exceeding DL % ≥ 2xDL					129 31.8 8.5				

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HYDROLOGIC OVERVIEW OF WATER YEAR 1992

Seasonal variations in atmospheric temperature and rainfall have a direct impact on water quality. Consequently, many water quality characteristics (temperature, dissolved oxygen, conductivity, water clarity, suspended solids, etc.) exhibit seasonal effects. During those times of the year when runoff is minimal (normally August-October) streamflow is derived principally from the base flow of ground water. Ground water contains greater concentrations of dissolved minerals than does surface drainage because of increased water/soil/rock contact and longer ground water residence time. During those times of the year when runoff is higher (normally January-March) streamflow is principally derived from rapid overland runoff that allows little time for mineral dissolution. Consequently, lower concentrations of most dissolved constituents are added to a river during heavy rainfall and subsequent high flows. However, periods of intense rainfall and high overland flows wash off or "flush" a watershed and transport soil particles to streams. This carries large loads of nonpoint source pollutants (nutrients, suspended solids, fecal bacteria, etc.) to streams and rivers. From a water quality perspective, low streamflows not only often result in higher water conductivity and higher water clarity, but also lakes and rivers are less able to dilute and assimilate the anthropogenic wastes discharged to them. Since low streamflows often occur during the warmer summer months, the problem of low streamflows can be critical. Warmer water temperatures combined with low streamflows enhance biological activity and thermal stratification, resulting in the potential for dissolved oxygen deficit problems and impacts on aquatic life. One of the important benefits of the TVA reservoir system is the ability to maintain adequate streamflow through the reservoir system during extended periods of low rainfall and low runoff by the controlled release of water. Such was the case in the summer of 1992 (June through October), when rainfall was below normal, but streamflow

through the system was regulated at just slightly above average levels, thereby helping to maintain a healthy aquatic river system. Consequently, examining atmospheric temperature, rainfall, and runoff patterns during 1992 aids in interpretation of the Reservoir Vital Signs Monitoring data.

Atmospheric Temperature

Average annual temperature in the TVA region is approximately 60 degrees Fahrenheit,°F (15.6 degrees Celsius,°C) with January usually being the coldest month and July the hottest. According to U.S. Department of Commerce climatic data, atmospheric temperatures in the TVA region averaged about 0.3° F (0.2° C) cooler than normal in 1992 (USDOC, 1992). January and February were unusually warm with 2.6° F (1.4° C) and 4.9° F (2.7° C) above normal, respectively, Figure 2-a. However, the rest of the months were near or below normal. May, June, August, and October had departures greater than -1.0° F (-0.6° C). This resulted in a cooler than normal growing season.

<u>Rainfall</u>

The Tennessee River basin averages about 51-52 inches (1295-1320 millimeters [mm]) of precipitation annually. However, there are large variations in the spatial distribution of precipitation. The range is from a high of about 93 inches (2360 mm) in the mountains of southwestern North Carolina near Highlands to a low of about 37 inches (940 mm) in the shielded valleys of these same mountains near Asheville, North Carolina. Elsewhere in the Valley, precipitation usually ranges within five to ten inches of the basin average. March is usually the wettest month and October the driest.

Rainfall in the Tennessee Valley in 1992 averaged 43.4 inches (1102 mm), about 8 inches (204 mm) less than the long term 100-year average, (a departure of about minus 15 percent) and about 12.7 inches (323 mm) less rainfall than 1991 (TVA 1992). Following a wet October-December 1991, each of the first five

months of 1992 was more than an inch (25 mm) below the long term average, with the greatest departure being -2.4 inches (-61 mm) in April, as shown in Figure 2-b. Consequently, the period January-May 1992 ranked as one of the ten driest on record in the Tennessee Valley. In spite of this rainfall deficit, all TVA reservoirs were at summer pool levels by the end of May. Rainfall during the summer (June through October) was slightly below normal (0.1 inches). Rainfall was rather evenly distributed in the Tennessee Valley in 1992 with that portion east of Chattanooga receiving about 43.5 inches (1105 mm) and that portion west of Chattanooga receiving about 43.3 inches (1100 mm).

Extreme precipitation events for 1992 were few. April 20-22 brought heavy rains along the Tennessee-North Carolina border resulting in seven inch plus (>180 mm) storm totals. A freakish snowstorm hit May 5-8 dropping as much as 60 inches (1524 mm) of snow at Mt. Pigsah, North Carolina. Hurricane Andrew remnants dropped over five inches (127 mm) of rain in a 24 hour period on August 27 in northwest Georgia.

<u>Streamflow</u>

Streamflow varies seasonally with rainfall, although during the spring and summer evaporation, transpiration, and infiltration reduce the amount of runoff. Watersheds that receive 50 to 60 inches of precipitation annually average about 20 to 30 inches of runoff. In a normal year, the discharge of the Tennessee River (approximately 64,000 cfs) corresponds to about 22 inches of runoff distributed over the 40,900 square mile drainage basin. A larger amount of runoff occurs during the wet winter and spring months (January-April) when precipitation events are frequent, temperatures are low, and there are no leaves on deciduous vegetation. Consequently, soil absorption, evaporation, and plant transpiration losses are low at that time of year, and both runoff and streamflow are higher than during the summer and fall months. In 1992, runoff was about an inch

(25 mm) below normal, with the first six months of the year having below normal runoff and the last six months having above average runoff, Figure 2-c. The abnormally dry spring and low runoff (January-May) of 1992, combined with the spring filling of the tributary reservoirs (resulting in the release of little water from the tributary reservoirs) resulted in low flows in the mainstem Tennessee River reservoirs, particularly in April. Consequently, an unusual episode of early spring thermal stratification on the tributary and many of the mainstem reservoirs was observed in April 1992. Higher, more normal flows in May and June (after the filling of the tributary reservoirs and higher amounts of rain) resulted in the destratification of the mainstem reservoirs and a return to more normal reservoir conditions. The impacts of the early spring stratification on the water quality of several reservoirs is discussed in the following chapter.

The net result for the Tennessee Valley in 1992 was an annual 15 percent deficit in precipitation with resultant total runoff that was approximately 0.9 inches below the long-term mean of 22.5 inches. Mean streamflows during 1992 for each of the Vital Signs reservoirs reflect the lower than average annual runoff (table 1).

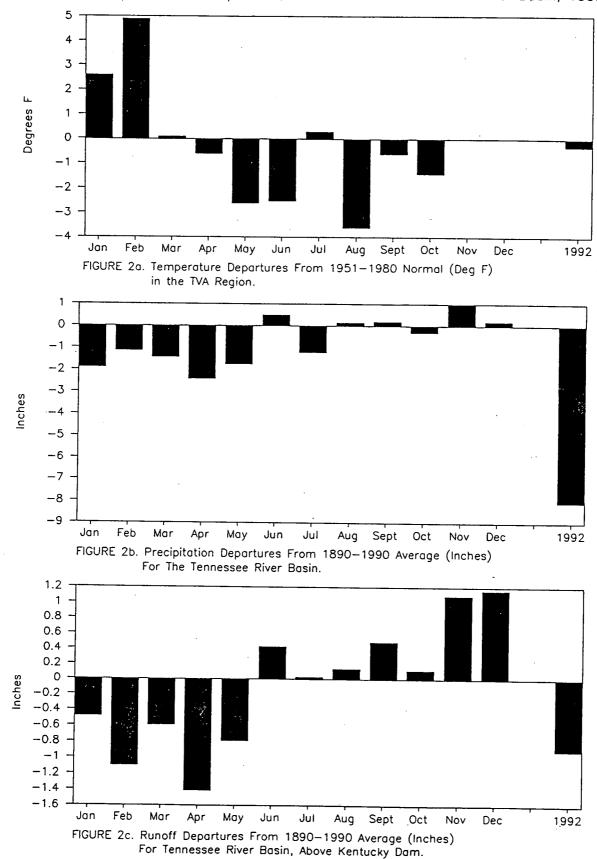


FIGURE 2. Temperature, Precipitation, and Runoff - Tennessee River Basin, 1992

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forebay, but pHs did not exceed 7.8 at the transition zone. There was about a 23 percent decrease in average nitrite plus nitrate concentrations from 0.31 mg/l at the transition zone to 0.24 mg/l at the forebay (along with corresponding increases in organic nitrogen and organic carbon) suggesting the photosynthetic uptake of nutrients and primary production processes occurring in the lower end of Guntersville Reservoir.

At the forebay, the highest chlorophyll-a concentration of 12 ug/l was measured in August (average summer chlorophyll-a concentration was 6-7 ug/l in 1992). At the transition zone chlorophyll-a concentrations were lower, averaging about 4 ug/l. TN/TP ratios frequently exceeded 20 at both the forebay and transition zone, indicating conditions when phosphorus concentrations may have limited photosynthesis. Water clarity on Guntersville Reservoir in 1992 was the highest among the mainstem Tennessee River reservoirs, with average Secchi depths of 1.8 and 1.6 meters at the forebay and transition zone, respectively.

Nickajack Reservoir

Surface water temperatures ranged from $6.8\circ$ C in January to 27.6°C in July in the forebay; and values for DO at the 1.5-meter depth ranged from 11.4 mg/l in January to 5.5 mg/l in September at the forebay.

The riverine character of Nickajack Reservoir, with an average hydraulic residence time of only three to four days (table 3.1), results in it being the best mixed of any of the Vital Signs reservoirs. Temperature and DO data reflect a lack of stratification in Nickajack reservoir in 1992, with the exception that in April, during low flow conditions on the Tennessee River, a maximum temperature differential (surface to bottom) of 3.8°C was measured at the forebay. During the same month, the maximum DO differential was 2.8 mg/l. In all other months the reservoir was well mixed. The minimum DO measured in Nickajack Reservoir in 1992 was 5.0 mg/l, at the bottom of the forebay, in July. DOS of 2.7, 3.8, and 4.5 mg/l were measured in

the releases of water from Chickamauga Dam in July and August.

Values of pH and conductivity varied over a rather narrow range, from 7.1-8.2 and from about 160-190 umhos/cm, respectively. At the forebay, the highest chlorophyll-a concentration of 9 ug/l was measured in April and averaged about 4 ug/l in 1992. Values of pH of 8.2 and DO saturation of 105-110 percent (which were the highest pH and DO saturations observed in Nickajack reservoir in 1992) were measured in April coincident with the high chlorophyll-a observation at the forebay.

Chickamauga Reservoir

Surface temperatures ranged from 6.8°C in January to 28.0°C in July in the forebay and from 6.1°C to 26.1°C for the same months at the transition zone. Values for DO at the 1.5-meter depth ranged from 11.4 mg/l in January to 5.4 mg/l in September at the forebay and from 11.5 mg/l to 4.6 mg/l for these same months at the transition zone. The 4.6 mg/l concentration of DO at the 1.5-meter depth is the lowest in-reservoir DO measured at the 1.5-meter depth on any of the Vital Signs reservoirs in 1992, and is less than the State of Tennessee minimum water quality criteria for fish and aquatic life of 5.0 mg/l. The lowest measured DO in Chickamauga Reservoir in 1992 was 2.8 mg/l, found at the bottom of the forebay in July.

Like many other mainstem Tennessee River reservoirs, Chickamauga is generally well mixed and lacks any strong thermal stratification. However, the low flows of the Tennessee River system in April and early May facilitated the development of a weak thermocline and oxycline in these months at both the forebay and transition zone sampling locations, in 1992. Maximum temperature differentials (surface to bottom) of 4.5°C and 3.0°C were observed at the forebay, in April and May, respectively. At the transition zone, in April and May, maximum temperature differentials of 2.4°C and 3.0°C, respectively, were measured. During these same two months, oxygen differentials of 3.2 mg/l

and 5.8 mg/l, respectively, were measured at the forebay; and, 3.3 mg/l and 4.7 mg/l, respectively, were measured at the transition zone. (The larger oxygen differentials measured in May were a result high DOs at the water surface during a period of high photosynthetic activity.) Minimum DOs measured in Chickamauga Reservoir in 1992 were 2.8 mg/l and 3.5 mg/l, at the bottom of the forebay and the transition zone, respectively, in July.

Values of pH ranged from 7.0 to 8.6. Conductivity ranged from about 155 to 195 umhos/cm, and averaged about 170 umhos/cm. Comparison of pH and conductivity at the transition zone with upstream pH and conductivity at Watts Bar Dam forebay indicates these are lowered by the soft water inflows of the Hiwassee River to Chickamauga Reservoir, about nine miles upstream of the transition zone.

Average total nitrogen concentrations in Chickamauga Reservoir were among the lowest measured at Vital Signs Monitoring locations on the Tennessee River in 1992. In addition, both total phosphorus and dissolved ortho phosphorus concentrations were also among the lowest observed at any of the Vital Signs Monitoring locations on the Tennessee River.

The highest chlorophyll-a concentrations were measured in May, 12 ug/l and 7 ug/l, respectively, at the forebay and transition zones. Concentrations of chlorophyll-a averaged 6-7 ug/l at the forebay and 4-5 ug/l at the transition zone in 1992.

Watts Bar Reservoir

Surface water temperatures ranged from 6.0°C in January to 27.3°C in July in the forebay and from 6.2°C to 26.3°C for these same months at the transition zone. Values for DO at the 1.5-meter depth ranged from 11.6 mg/l in January (as well as 11.6 mg/l in April due to high photosynthetic activity) to 6.3 mg/l in September at the forebay; and, from 11.4 mg/l in January to 5.8 mg/l in September at the transition zone. The minimum

observed DO concentration in Watts Bar Reservoir in 1992 was 0.6 mg/l at the bottom of the forebay in July.

Temperature and dissolved oxygen data show that during the summer of 1992, Watts Bar Reservoir developed a moderate degree of both thermal and oxygen stratification in the forebay. Surface to bottom temperature differentials ($\Delta T's$) were 7.0°C in April (during the period of low flows) and exceeded 6°C in May and June. DO versus depth data showed a rather strong oxycline to develop in the forebay of Watts Bar Reservoir from May through August. During these four months surface to bottom differences in DO were consistently greater than 7.0 mg/l, and near bottom DO concentrations in the hypolimnion were less than 1 mg/l in July. The transition zone was much more well mixed during the summer of 1992. Maximum $\Delta T's$ were 4.1°C (in April) and the minimum bottom DO measured was 5.5 mg/l (in September).

Values of pH ranged from 6.7 to 9.1 on Watts Bar Reservoir. Through out the summer (April-August) near surface values of pH in the forebay were often high, exceeding 8.5, with DO saturation values commonly exceeding 100 percent, indicating high rates of photosynthesis.

The average total phosphorus concentrations observed in Watts Bar Reservoir (0.029 mg/l at the forebay and 0.033 mg/l at the transition zone) were among the lowest of the Tennessee River Vital Signs Monitoring locations. In addition, the average dissolved ortho phosphorus concentrations of 0.008 mg/l and 0.010 mg/l, respectively, at the forebay and transition zones were also among the lowest observed at any of the Tennessee River Vital Signs Monitoring locations in 1992.

The highest chlorophyll-a concentrations were measured in June at the forebay (14 ug/l) and in May at the transition zone (14 ug/l). Surface concentrations of chlorophyll-a averaged about 7 ug/l at the forebay and about 8 ug/l at the transition zone in 1992. The high TN/TP ratios observed at the transition zone indicate the possibility of phosphorus limitation on primary productivity.

Forebay Secchi depth and suspended solids measurements averaged 1.4 meters and 4.9 mg/l, respectively. These values indicate the light transparency of Watts Bar Reservoir forebay to be relatively high compared with other mainstem Tennessee River reservoirs in 1992.

Fort Loudoun Reservoir

Temperature and dissolved oxygen (DO) data show the establishment of stratification (both a thermocline and oxycline) in the forebay portion of the reservoir which persisted throughout most of the summer (April through August) of 1992. Surface water temperatures ranged from 6.6° C in January to 28.7° C in July at the forebay and from 6.1° C to 29.6° C for the same months at the transition zone. Maximum thermal stratification occurred in the forebay in June when surface to bottom temperature differentials (Δ T's) were 8.2° C, and in the transition zone in April when Δ T's of 9.9° C were observed.

In Fort Loudoun Reservoir in 1992, DO at the 1.5-meter depth ranged from 11.5 mg/l in August (algal bloom) to 5.3 mg/l in September at the forebay; and from 14.0 mg/l in January to 5.4 mg/l in September at the transition zone. The minimum DO observed in Fort Loudoun Reservoir in 1992 was 1.8 mg/l at the bottom of the forebay during August. Maximum surface to bottom dissolved oxygen differentials ($\Delta DO's$) exceeded 7 mg/l each month, May through August, at the forebay. The transition zone was better mixed with $\Delta DO's$ exceeding 3 mg/l observed only in July, and a minimum bottom DO of 5.1 in August.

Values of pH ranged from 6.5 to 9.1. At the forebay, pH values exceeding 8.5, and DO saturation values exceeding 110 percent were measured from April through August giving evidence of substantial photosynthetic activity. During April, May, and July, a similar pattern of high pHs and high DO saturations was observed, although to a lesser extent, at the transition zone.

Conductivity ranged from 90 to 255 umhos/cm, averaging about

195 umhos/cm at the forebay and 215 umhos/cm at the transition The slightly lower conductivities measured at the forebay zone. area were caused by the mixing of the inflows from the Little Tennessee River, via the Tellico Reservoir canal with the higher conductivity water of the Tennessee River. For example, during summer, water from Tellico Reservoir is usually colder than the surface water of Fort Loudoun Reservoir causing it to flow under the warmer water of Fort Loudoun Reservoir. This was the case in September, 1992, when water surface conductivity was greater than 200 umhos/cm and near bottom conductivity was about 110 umhos/cm in the forebay of Fort Loudoun Reservoir. In the spring, the water from Tellico Reservoir may be warmer than the water of Fort Loudoun Reservoir and often flow across the top and "float" on the surface of the Fort Loudoun Reservoir. Such was the case in April 1992, when the Fort Loudoun forebay had surface conductivity less than 100 umhos/cm and near bottom conductivity near 200 umhos/cm. Other months (e.g., May, June, July, etc.) give evidence of partially mixed "lenses" of low conductivity water from Tellico Reservoir merging with the higher conductivity water from Fort Loudoun Reservoir forebay at one or more depths.

Nutrient concentrations (total nitrogen and total phosphorus) were high at both the forebay and the transition zone. The average nitrite plus nitrate nitrogen concentrations of 0.55 mg/l (forebay) and 0.41 mg/l (transition zone) were the highest average concentrations of this nutrient measured in 1992 at any of the Tennessee River Vital Signs Monitoring locations. These high concentrations of nitrogen are due to a combined effect of the wastewater discharges in the Knoxville metropolitan area and the inflows to Fort Loudoun Reservoir from the Holston and French Broad rivers, which also have relatively high nitrogen concentrations.

The highest chlorophyll-a concentrations in the forebay occurred in May (17 ug/l) and June (18 ug/l) and in the transition zone in April (18 ug/l). Surface concentrations of chlorophyll-a averaged about 11 ug/l and 8 ug/l, at the forebay

TABLE 8

Guntersville Forebay

Guntersville Transition

Variable	N	Mean	Min	Max	N	Mean	Min	Max
Variable Temperature (C) Dissolved Oxygen (mg/l) Percent Saturation pH (s.u.) Conductivity (umhos/cm) Organic - N (mg/l) Ammonia - N (mg/l) Nitrate+Nitrite - N (mg/l) Total Nitrogen (mg/l) Total Phosphorus (mg/l) Dissoved Ortho - P (mg/l) TN/TP Ratio Total Organic Carbon (mg/l) Chlorophyll-a (ug/l) Secchi depth (m) Suspended Solids (mg/l)	73 73 73 73 73 73 18 18 18 18 18 18 18 18 18 18 18 18 18	21.5897.37180.7187.509 $165.1940.3040.0510.2430.5980.0320.01519.1962.6295.7141.8433.571$	$\begin{array}{c} 6.930\\ 2.930\\ 37.089\\ 6.840\\ 147.000\\ 0.110\\ 0.020\\ 0.090\\ 0.330\\ 0.030\\ 0.003\\ 8.250\\ 1.900\\ 1.000\\ 1.500\\ 1.000\\ 1.000\end{array}$	Max 28.300 10.710 115.976 8.350 186.000 0.620 0.090 0.540 0.880 0.040 0.030 29.333 4.300 12.000 2.250 7.000 15.000	N 53 53 45 53 14 14 13 13 14 14 13 14 14 7 7 7 14	Mean 22.603 7.120 79.321 7.548 176.865 0.238 0.050 0.312 0.595 0.033 0.019 18.570 2.507 3.571 1.571 4.214 10.000	Min 7.260 5.410 67.500 7.200 157.000 0.080 0.020 0.170 0.360 0.020 0.005 9.000 1.900 1.000 1.000 1.000 5.000	Max 28.530 10.660 101.111 7.830 194.000 0.370 0.130 0.540 0.920 0.040 0.030 26.300 3.500 9.000 2.000 8.000 15.000
True Color (PCU) Fecal Coliform (#/100 ml)	18 7	10.000 10.000	5.000 10.000	10.000	7	10.000	10.000	10.000

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Nickajack Forebay (TRM 425.5)

Variable	N	Mean	Min	Max
Temperature (C) Dissolved Oxygen (mg/l) Percent Saturation pH (s.u.) Conductivity (umhos/cm) Organic - N (mg/l) Ammonia - N (mg/l) Nitrate+Nitrite - N (mg/l) Total Nitrogen (mg/l) Total Phosphorus (mg/l) Dissoved Ortho - P (mg/l) TN/TP Ratio Total Organic Carbon (mg/l) Chlorophyll-a (ug/l) Secchi depth (m) Suspended Solids (mg/l) True Color (PCU)	85 85 85 36 36 36 36 36 36 36 36 36 36 36 36	$\begin{array}{c} 20.618\\ 7.169\\ 76.312\\ 7.436\\ 174.303\\ 0.200\\ 0.060\\ 0.312\\ 0.573\\ 0.033\\ 0.020\\ 17.928\\ 2.104\\ 4.071\\ 1.383\\ 4.143\\ 10.893\\ 10.893\end{array}$	6.700 5.000 62.069 7.100 158.000 0.070 0.010 0.360 0.020 0.006 11.750 1.800 1.000 1.210 2.000 5.000 10.000	27.700 11.500 108.421 8.200 191.000 0.490 0.140 0.520 0.850 0.050 0.040 25.667 2.300 11.000 1.600 7.000 15.000 10.000
Fecal Coliform (#/100 ml)	7	10.000	10.000	10.000

Chickamauga Forebay (472.3)

Variable	N	Moan			Chickamau	iga Trans:	ition (TRI	490.47)
Temperature (C) Dissolved Oxygen (mg/l) Percent Saturation pH (s.u.) Conductivity (umhos/cm) Organic - N (mg/l) Ammonia - N (mg/l) Nitrate+Nitrite - N (mg/l) Total Nitrogen (mg/l) Total Phosphorus (mg/l) Dissoved Ortho - P (mg/l) TN/TP Ratio Total Organic Carbon (mg/l) Chlorophyll-a (ug/l) Secchi depth (m) Suspended Solids (mg/l) True Color (PCU) Fecal Coliform (#/100 ml)	N 81 81 81 81 81 81 18 18 18 18	Mean 20.680 7.057 75.224 7.470 171.148 0.225 0.053 0.251 0.529 0.028 0.009 20.904 2.121 5.167 1.348 4.429 11.071 10.000	Min 6.800 2.800 34.146 7.000 155.000 0.080 0.010 0.100 0.230 0.020 0.022 7.000 1.800 1.000 1.130 2.000 10.000 10.000	Max 28.000 11.500 126.667 8.600 194.000 0.520 0.100 0.520 0.740 0.060 0.020 33.000 2.300 12.000 1.500 7.000 15.000	N 51 51 51 51 51 51 12 12 12 12 12 12 12 12 12 12 12 12 12	Mean 19.071 7.474 77.881 7.566 166.890 0.265 0.060 0.285 0.610 0.035 0.011 18.704 2.133 4.000 1.283 5.250 10.417 10.000	Min 6.100 4.500 52.941 7.300 155.000 0.100 0.020 0.150 0.330 0.020 0.002 8,250 1.800 2.000 1.210 3.000 5.000 10.000	Max 25.300 11.700 11.700 117.021 8.400 190.000 0.430 0.090 0.520 0.760 0.050 0.020 28.500 2.500 7.000 1.470 8.000 15.000 10.000

Watts Bar Forebay (TRM 531)

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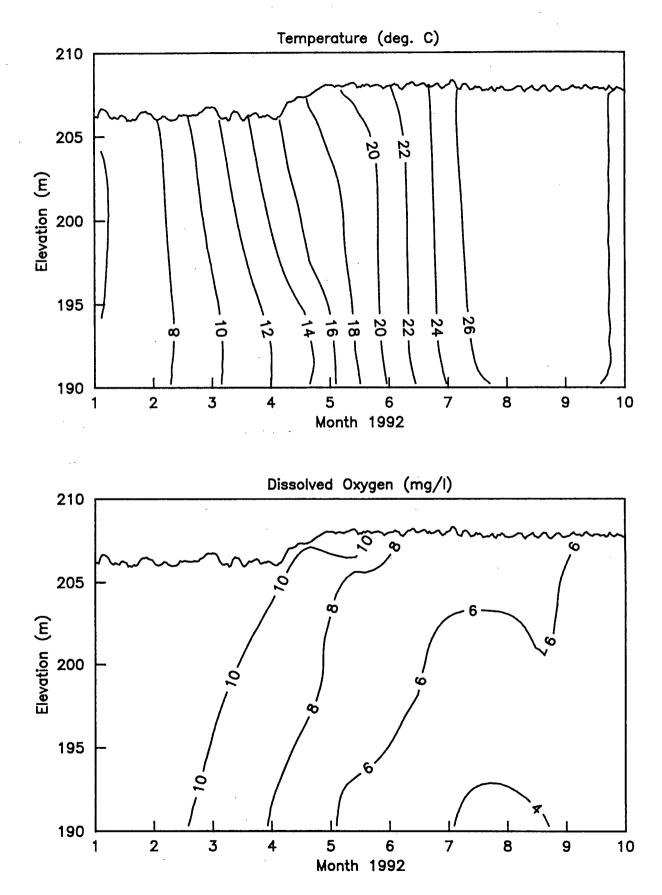
Variable N Temperature (C) Dissolved Oxygen (mg/l) Percent Saturation pH (s.u.) Conductivity (umhos/cm) Organic - N (mg/l) Ammonia - N (mg/l)Nitrate+Nitrite - N (mg/l) Total Nitrogen (mg/l) Total Phosphorus (mg/l) Dissoved Ortho - P (mg/l) TN/TP Ratio Total Organic Carbon (mg/l) Chlorophyll-a (ug/l) Secchi depth (m) Suspended Solids (mg/l) True Color (PCU) Fecal Coliform (#/100 ml)

Watts Bar Transition (TRM 560.8)

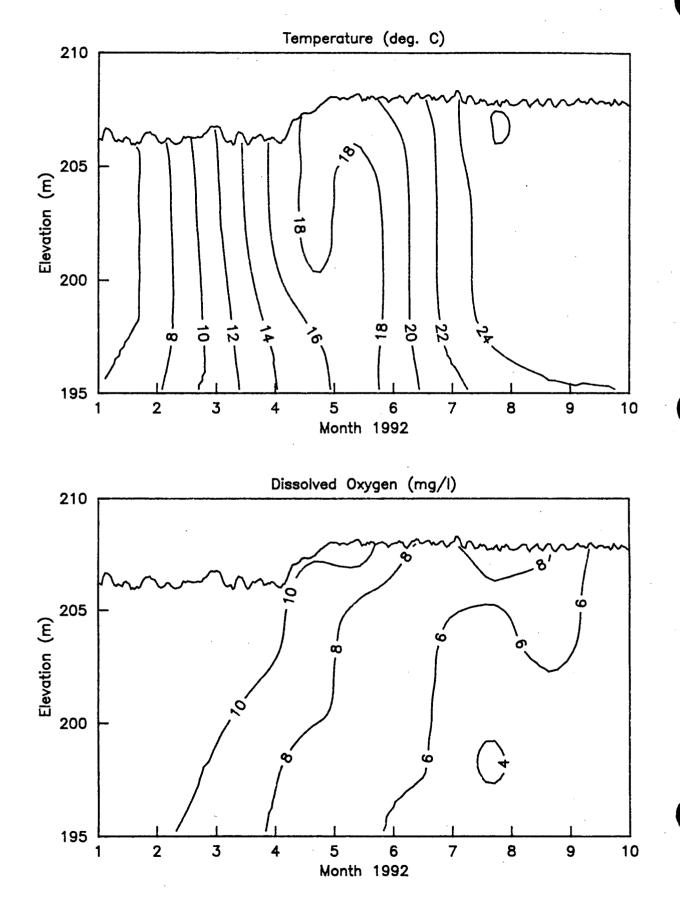
ſ	Mean	Min	Max
124 124 124 124 18 18 18 18 18 18 18 18 18 18 18 18 18	$19.307 \\ 6.856 \\ 71.527 \\ 7.685 \\ 172.769 \\ 0.307 \\ 0.025 \\ 0.281 \\ 0.613 \\ 0.029 \\ 0.008 \\ 21.440 \\ 2.100 \\ 7.286 \\ 1.393 \\ 4.917 \\ \end{array}$	Min 6.000 0.600 6.897 6.700 137.000 0.100 0.010 0.100 0.370 0.020 0.002 14.666 1.700 4.000 1.100 3.000	Max 27.300 11.800 142.683 9.100 200.000 0.630 0.060 0.570 0.780 0.040 0.030 38.000 2.800 14.000 1.770 10.000
16 7	11.250 10.000	5.000 10.000	15.000 10.000

N	Mean	Min	Max
68 68 68 14 14 14 14 14 14 14 14 14 14 12 7 7 22 22 7	19.478 8.354 88.222 7.810 187.276 0.312 0.033 0.354 0.699 0.033 0.010 43.198 2.114 7.857 1.103 9.083 10.833 10.000	6.100 5.500 63.218 7.400 150.000 0.170 0.010 0.540 0.002 0.002 12.600 1.900 4.000 0.800 4.000 5.000 10.000	26.300 11.500 125.000 8.700 213.000 0.600 0.090 0.720 0.940 0.050 0.020 325.000 2.300 14.000 1.270 17.000 15.000 10.000

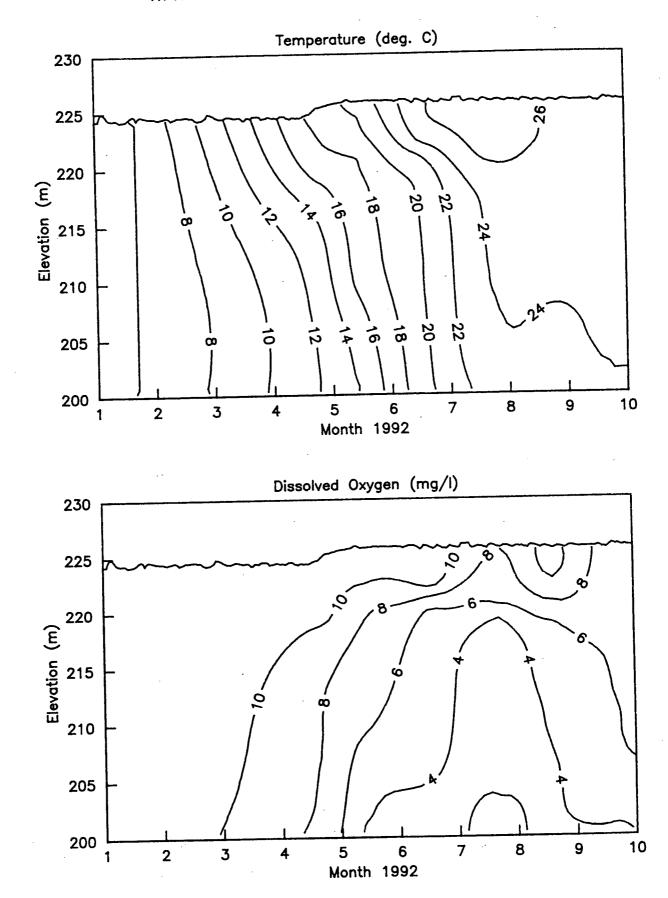
CHICKAMAUGA RESERVOIR - TRM 472.3



CHICKAMAUGA RESERVOIR - TRM 490.5

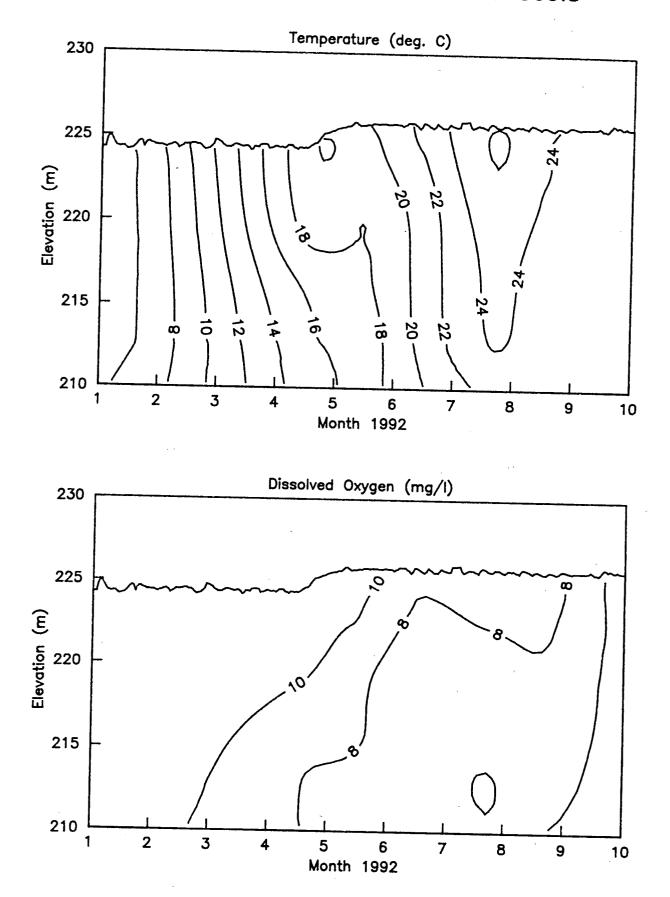


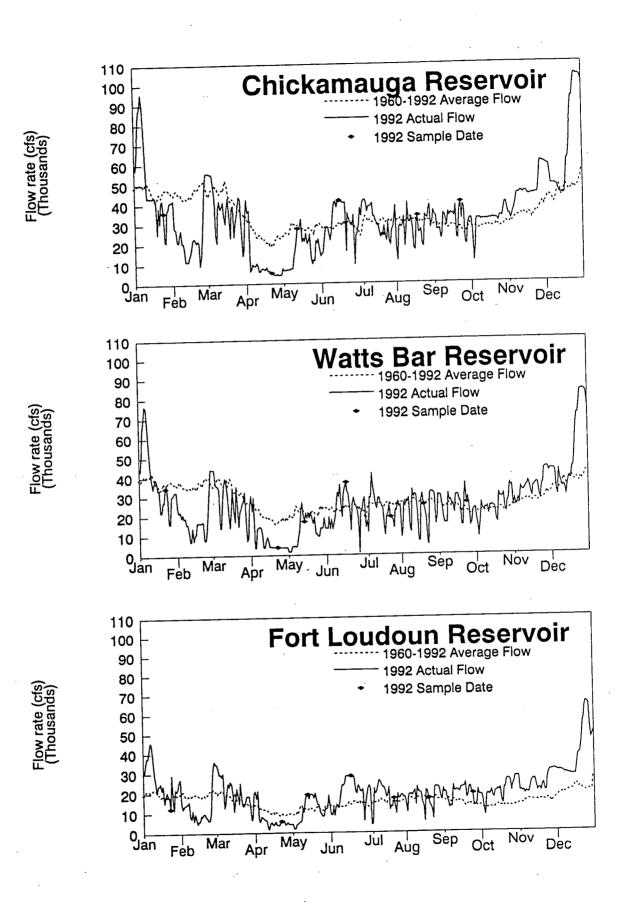
WATTS BAR RESERVOIR - TRM 531.0

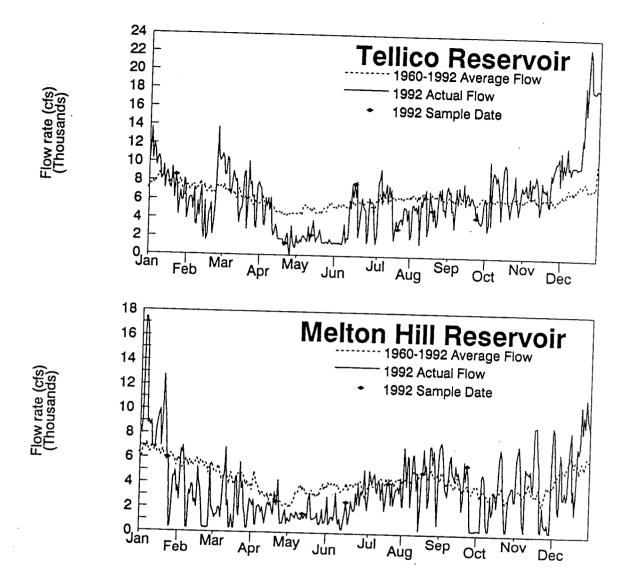


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WATTS BAR RESERVOIR - TRM 560.8







PGM≃RET

475358 1017 35 06 26.0 085 12 20.0 2 CHICKAMAUGA RES. AT LIGHTED BUOY 47065 TENNESSEE HAMILTON TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 472.3 131TVAC 06020001021 0000.710 ON 0000 METERS DEPTH

/TYPA/AMBNT/STREAM/SOLIDS

A-23

DATE FROM TO 92/01/24	TINE OF DAT	MEDIUN	SMK OR DEPTH (H) 0.5	00060 STREAM FLOW CFS	00010 WATER TEMP CENT	00300 D0 Hg/L	00301 DO SATUR PERCENT	00400 Ph Su	00094 CNDUCTVY Field Micronho	DATE From To	TIME OF Day	MEDIUN	SMK OR DEPTH (M)	00060 STREAM FLOW CFS	OGO10 WATER TEMP CENT	00300 .D0 Hg/L	00301 Do Satur Percent	00400 Ph Su	00094 CNDUCTVY FIELD MICRONHO
 92/01/24 92/01/24 92/01/24 92/01/24 92/01/24 92/01/24 92/01/24 92/01/24 92/01/24 92/01/24 92/01/24 92/04/21	1000 1001 1002 1004 1005 1006 1007 1008 1009 1010 1035	WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER	0.5 1 1.5 6 8 10 12 13 14 15 0.5	36108 5421	6.8 6.8 6.8 6.8 6.8 6.8 6.8 6.8 6.8 6.8	11.5 11.5 11.4 11.4 11.3 11.3 11.3 11.2 9.9	94.38 94.38 93.48 92.68 92.68 92.68 92.68 92.68 91.88 91.88 91.88	7.50 7.50 7.50 7.50 7.40 7.40 7.40 7.40 7.40 7.40 7.40 7.4	168 168 169 169 169 167 168 168 168 168 168	92/06/15 92/06/15 92/06/15 92/06/15 92/06/15 92/07/21 92/07/21 92/07/21 92/07/21 92/07/21 92/07/21	1147 1149 1151 1153 1108 1110 1112 1114 1118 1120	WATER WATER WATER WATER WATER WATER WATER WATER WATER	10 12 14 14.2 0.5 1 1.5 5.5 6	24342	22.6 22.6 22.6 28.0 27.8 27.8 27.4 27.4	6.8 5.7 5.6 7.0 7.0 6.3 7.0 5.3 5.3 5.3	69.05 66.75 65.55 64.45 96.25 92.45 88.65 79.75 65.45	7.40 7.40 7.30 7.30 7.90 7.80 7.70 7.50 7.30 7.30	173 174 174 165 165 165 165
92/04/21 92/04/21 92/04/21 92/04/21 92/04/21 92/04/21 92/04/21 92/04/21 92/04/21 92/05/12 92/05/12 92/05/12 92/05/12	1039 1041 1045 1047 1049 1051 1055 1057 1058 1105 1104 1106 1108	WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER	1.5 6 10 12 13 14 0.5 1.5 2.5	21454	18.0 17.7 17.5 16.8 16.5 16.1 14.3 14.3 14.3 14.3 14.3 14.3 14.3 19.8 19.8 19.2	9.7 9.3 8.5 8.1 7.3 7.3 6.7 11.5 10.3 9.9 7.9	102.1s 101.1s 95.9s 87.6s 87.6s 76.5s 70.2s 70.2s 126.7s 114.1s 112.0s 107.6s 84.0s	7.90 7.80 7.80 7.50 7.50 7.40 7.30 8.60 8.60 8.40 8.30 8.20 8.20 7.80	160 160 160 158 159 158 156 167 167 167 167	92/07/21 92/07/21 92/07/21 92/07/21 92/07/21 92/07/21 92/08/18 92/08/18 92/08/18 92/08/18 92/08/18 92/08/18	1124 1126 1128 1130 1132 1134 1134 1107 1110 1110 11113 1116 1122 1125 1128 1128 1128	WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER	8 10 12 14 15 5 16 16 5 0 5 1 1 5 6 8 10 12	33592	27.3 27.0 26.8 26.6 26.6 27.1 27.1 27.1 27.1 27.1 27.1 27.1 27.1	5.98 4.85 4.85 5.38 6.3 6.21 6.21 6.25 5.5	65.48 60.55 59.33 55.63 44.43 40.75 82.75 80.23 77.85 76.55 76.55 76.55 74.15 70.45	7.30 7.20 7.10 7.00 7.60 7.60 7.60 7.50 7.50 7.50 7.40	165 166 165 166 166 167 193 194 193 193 193 193
92/05/12 92/05/12 92/05/12 92/05/12 92/05/12 92/05/12 92/05/12 92/06/15 92/06/15 92/06/15 92/06/15	1118 1120 1122 1124 1126 1132 1134 1131 1133 1133 1137 1137 1137 1137 1137 1137 1137	WATER WATER WATER JATER JATER JATER JATER JATER JATER JATER JATER JATER	4 8 10 12 13.6 14 16 0.3 1 1.5 4 8	41238	18.6 18.5 18.4 18.4 18.0 17.8 23.0 23.0 22.8 22.8 22.8 22.7	7.0 6.8 6.7 6.5 6.4 5.6 6.4 5.6 6.4 5.6 6.4 5.6 7.0 6.7 6.7 6.2 6.1	74.55 70.55 71.65 70.55 68.45 67.45 58.95 80.55 79.35 77.05 73.65 71.35 70.15	7.60 7.50 7.40 7.40 7.30 7.40 7.50 7.50 7.50 7.50 7.50 7.40	166 166 166 166 166 165 165 165 174 174 174 173 174	92/08/18 92/08/18 92/08/18 92/09/22 92/09/22 92/09/22 92/09/22 92/09/22 92/09/22 92/09/22 92/09/22 92/09/22 92/09/22	1137 L 1140 L 1034 L 1035 L 1036 L 1037 L 1040 L 1040 L 1042 L 1043 L 1044 L	JATER JATER JATER JATER JATER JATER JATER JATER JATER JATER JATER	13.5 14 16 0.5 1 1.5 5 7 9 11 13 15 17	40142	26.7 26.5 26.0 26.0 26.0 26.0 26.0 26.0 26.0 26.0	5.11444 5.45555110 5.555555555555555555555555555555	63.05 63.05 50.05 65.95 64.65 64.65 64.65 64.65 64.65 62.25 62.25 62.25 61.05 59.85	7.30 7.30 7.50 7.50 7.50 7.40 7.40 7.40 7.40 7.30	191 190 190 176 177 176 177 176 175 173 174 177 177

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/TYPA/AMBNT/STREAM/SOLIDS

475358	1017	
35 06 26.0	085 12 20.0 2	
CHICKAMAUGA	RES. AT LIGHT	ED BUOY
47065 TEN		HAMILTON
TENNESSEE R	IVER BASIN	040801
TENNESSEE R	IVER 472.3	
131TVAC		060200
0000 METERS	DEPTH	

5020001021 0000.710 ON

DATE FROM TO	TIME OF DAY	MEDIUM	SMK OR DEPTH (M)	84002 CODE GENERAL REMARKS	00078 TRANSP SECCHI METERS	31616 FEC COLI MFM-FCBR /100ML	00080 Color Pt-co Units	00530 RESIDUE TOT NFLT MG/L	00605 ORG N N Mg/L	00610 NH3+NH4- N TOTAL MG/L	00630 N02&N03 N-TOTAL MG/L
92/01/24			0.5		1.50	10K					
92/01/24 92/01/24	1003	VERT	4				10	3	.180	.040	.51
92/04/21	1035	WAIER	13 0.5				10	2	.180	.040	.52
92/04/21	1043	VERT	4			10K					
92/04/21			13				10 10	4	.180 .190	- 040	.28
92/05/12 92/05/12			0.5		1.47	10K			. 190	.090	.34
92/05/12	1112	VERI	4	D 1 D 2			15	5 5	.100	.030	.10
92/05/12	1116	VERT	2	D3			15 10	5	.370	.020	.11
92/05/12	1126	WATER	13.6	D 1			10	5 3	.520	.030	. 11
92/05/12	1128	WATER	13.6	D 2			iŏ	4	.230	.090 .090	.15 .14
92/05/12 92/06/15	1130	WAIER	13.6	D 3	4 47		10	4	.180	.100	. 15
92/06/15	1139	VERT	4		1.13	10	10				
92/06/15	1151	WATER	14.2				10	4	.080 .360	.060	.27
92/07/21 92/07/21	1108	WATER	0.5		1.37	10K	10	4	.300	.070	.27
92/07/21	1128	VEKI	4 14				10	6	.330	.030	.20
92/08/18			0.5		1.36	10K	10	6	.250	.060	.24
92/08/18			4		1.50	TUK	10	5	.280	010	
92/08/18			13.5				10	3	.150	.010 .050	.19 .22
92/09/22 92/09/22			0.5		1.26	10K				.050	
92/09/22	1044	WATER	15				15	5	.180	.060	.11
							15	7	.240	.070	.11
FROM	TIME OF DAY	MEDIUM	SMK OR DEPTH (M)	84002 CODE GENERAL REMARKS	00665 Phos-tot Mg/l p	00671 PHOS-DIS ORTHO MG/L P	00680 T ORG C C MG/L	32211 Chlrphyl A Ug/L Correctd	32212 CHLRPHYL B	32214 CHLRPHYL C	32218 PHEOPHIN A
92/01/24	1003	VEPT	4					CORRECTO	UG/L	UG/L	UG/L
92/01/24	1008	WATER	13		.030 .030	.020	1.8	1.00	1.00K	1.00K	1.00
92/04/21 92/04/21 92/05/12 92/05/12	1053	WATER Vert	4 13 4 4	D 1	.020 .020 .020	.020 .009 .010 .003	1.8 2.0 2.0 2.3	10.00	1.00k	1.00	
92/05/12	1116 \	VFRT	4	D 2 D 3	.020 .020	.003	2.3	12.00	1.00	1.00	1.00K 1.00K
92/05/12	1126	WATER	13.6	Ď Ī	.020	.003	2.3 2.2				
92/05/12 92/05/12	1128 1		13.6	D 2	.020	.005	2.1				•
92/06/15	1139 \	VERT	13.6	D 3	.020	.005	2.1				
92/06/15	1151 \	√ATER	14.2		.030	.010	2.3	5.00	1.00K	1.00K	1.00K
92/07/21	1116	VERT	4		.020	.005	2.2	6.00	4		
92/07/21 92/08/18	1120 1	VATER VEDT	14		.020	.009	2.1	0.00	1.00	1.00	3.00
92/08/18 1	1134 6	JATER	4 13.5		.030	.002	2.3	4.00	1.00K	1.00K	1.00
92/09/22 1	1038 v	/ERT	4		.030	.006	2.2				1.00
92/09/22 1	1044 6	ATER	15		.060	.010	2.1 2.3	4.00	1.00	1.00	2.00

PGM=RET

475265 1053	5
35 18 00.0 085 04 33.	.02
CHICKAMAUGA RESERVOIR	2
47065 TENNESSEE	HAMILTON
TENNESSEE RIVER BASIN	040801
TENNESSEE RIVER 490.4	7
131TVAC	06020001025 0005.740 0
0000 METERS DEPTH	

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/TYPA/AMBNT/STREAM/SOLIDS

DATE FROM TO	T I ME OF DAT	NEDIUN	SMK OR DEPTH (M)	00060 STREAM FLOW CFS	00010 WATER TEMP CENT	00300 D0 Hg/L	00301 Do Satur Percent	00400 Рн Su	00094 CNDUCTVY FIELD HICROMHO	DATE FROM TO	TIME OF DAY	HEDIUM	SMK OR DEPTH (M)	00060 STREAN FLOW CFS	OGO 10 WATER TEMP CENT	00300 D0 Mg/L	00301 DO Satur Percent	00400 PH \$U	00094 CNDUCTVY Field Micromho
92/01/2/ 92/01/2/ 92/01/2/ 92/01/2/ 92/01/2/ 92/01/2/ 92/04/2/ 92/04/2/ 92/04/2/ 92/04/2/ 92/04/2/ 92/04/2/ 92/04/2/ 92/04/2/ 92/04/2/ 92/04/2/ 92/04/2/ 92/05/11 92/05/11 92/05/11 92/05/11 92/05/11 92/05/11 92/05/11 92/05/11 92/05/11 92/05/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/06/11 92/07/2 92/07/2 92/07/2		UAATTEERR RRRRRRRRRRRRRRRRRRRRRRRRRRRRRRR	0.5 1.5 46 80 1.5 46 7.8 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 46 82 9.5 1.5 2.5 1.5 2.5 1.5 2.5 1.5 2.5 46 82 9.5 1.5 2.5 1.5 2.5 1.5 2.5 1.5 2.5 2.5 1.5 2.5 2.5 2.5 1.5 2.5 2.5 2.5 1.5 2.5 2.5 2.5 1.5 2.5 2.5 1.5 2.5 2.5 1.5 2.5 2.5 2.5 2.5 2.5 2.5 2.5 2	· · ·	$\begin{array}{c} 6 . 1 \\ 6 . 1 \\ 6 . 1 \\ 6 . 1 \\ 8 . 1 \\ 8 . 1 \\ 8 . 8 . \\ 1 8 . 8 . \\ 1 8 . 8 . \\ 1 8 . 8 . \\ 1 8 . 8 . \\ 1 8 . 8 . \\ 1 1 8 . 8 . \\ 1 1 8 . 8 . \\ 1 1 1 . 1 . \\ 1 6 . . 1 . \\ 1 1 . 1 . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . \\ 1 . . 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TENNESSEE RIVER 490.		
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/TYPA/AMBNT/STREAM/SOLIDS

DATE FROM TO	TIME OF DAY	MEDIUM	SMK OR DEPTH (M)	84002 CODE GENERAL REMARKS	00078 TRANSP SECCHI METERS	31616 FEC COLI MFM-FCBR /100ML	00080 COLOR PT-CO UNITS	00530 RESIDUE TOT NFLT MG/L	00605 ORG N N MG/L	00610 NH3+NH4- N TOTAL MG/L	00630 N02&N03 N-TOTAL MG/L
92/01/24			0.5		1.25	10K					-
92/01/24			4			IUK	10	-			
92/01/24	0902	WATER	8				10 5	3	.180	.040	.51
92/04/22	0925	WATER	0.5		1.27	10K	2	5	.190	.030	.52
92/04/22			4		1.21	IUK	4.0	-			
92/04/22	0934	WATER	ġ				10	5	.210	.050	.26
92/05/12	0930	WATER	0.5		1.23	10K	10	6	.210	.090	.35
92/05/12	1000	VERT	4		1.25	TUK	4.0	_			
92/05/12			. 9.5				10	5 5	.210	.040	.26
92/06/15			0.3		1.21		2	5	.220	.080	.27
92/06/15			0.5		1.21	10					
92/06/15			9.4				15 10	7	.430	.060	.27
92/07/21			0.5		1 00		10	5	.400	.070	.28
92/07/21			0.5		1.80	10 K					
92/07/21			9				10 10	4	.260	.030	.28
		WATER	0.5		4 27		10	4	.170	.060	.30
92/08/18			0.5		1.27	10K					
92/08/18			8.5				10	4	.400	.020	.20
92/09/22			0.5				10	7	.380	.080	.20
92/09/22	0034	VEDT			1.47	10K					.20
92/09/22	0038	UATED	4 10				15	5	.250	.080	. 15
, _, 0, , LL	0,10		10				15	8	.100	.080	.15

DAT FRO TO	M OF	MEDIUM	SMK OR DEPTH (M)	84002 CODE GENERAL REMARKS	00665 Phos-tot Mg/l P	00671 PHOS-DIS ORTHO MG/L P	00680 T ORG C C MG/L	32211 CHLRPHYL A UG/L CORRECTD	32212 Chirphyl B Ug/l	32214 CHLRPHYL C UG/L	32218 Pheophtn A Ug/L
92/01 92/04	/24 0900 /24 0902 /22 0929 /22 0934	WATER	4 8 4 9		.030 .030 .030 .030	.010 .010 .010	1.9 1.8 2.2	2.00	1.00K	1.00K	1.00
92/05 92/05	/12 1000 /12 1020 /15 1030	VERT WATER	4 9.5		.020	.020 .008 .010	2.0 2.1 2.1	7.00	1.00	1.00	1.00K
92/06	/15 1032	WATER	9.4		.050	.010 .020	2.5 2.3	4.00	1.00	1.00K	1.00
92/07	/21 0951 /21 0957	WATER	4 9		.020 .020	.007 .010	2.1 2.0	5.00	1.00	1.00K	1.00
92/08	/18 1023 /18 1031	WATER	4 8,5		.040 .040	.002	2.2	5.00	1.00K	1.00K	1.00K
	/22 0934 /22 0938		4 10		.040	.020	2.2	2.00	1.00K	. 1.00	2.00

A-26



PGM=RET

475317 1089 35 38 10.0 084 47 06.0 2 OPP. LOWE BR. WAITS BAR RES. 47121 TENNESSEE MEIGS TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 531.0 1311VAC 06010201002 0002.040 ON 0300 METERS DEPTN

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/TYPA/AMBNT/FISH/STREAM/SOLIDS

	DATE TIME FROM OF TO DAY NEDIUM	SMK OR DEPTH (M)	00060 STREAM FLOW CFS	00010 Water Temp Cent	00300 D0 Mg/L	DO301 DO Satur Percent	00400 Ph Su	00094 CNDUCTVY Field Micronho	DATE TIME FROM OF TO DAY	MEDIUM	SMK OR DEPTH (M)	00060 STREAM FLOW CFS	00010 WATER TEMP CENT	00300 D0 HG/L 2.2	00301 D0 SATUR PERCENT 25.95	00400 PH SU 6.90	00094 CNDUCTVY FIELD Micronho 175
	92/01/22 1211 WATER 92/01/22 1212 WATER 92/01/22 1213 WATER 92/01/22 1214 WATER 92/01/22 1214 WATER 92/01/22 1214 WATER 92/01/22 1217 WATER 92/01/22 1219 WATER 92/01/22 1220 WATER 92/01/22 1221 WATER 92/01/22 1222 WATER 92/01/22 1222 WATER	0.5 1.5 6 10 12 14 16 18	34538	6.0 6.0 6.0 6.0 6.0 6.0 6.0 6.0 6.0 6.0	11.8 11.6 11.6 11.6 11.6 11.6 11.5 11.5 11.5	94.45 92.85 92.85 92.85 92.85 92.85 92.85 92.85 92.05 92.05 91.25 92.05	7.60 7.60 7.50 7.50 7.50 7.50 7.50 7.50 7.50 7.5	174 176 176 175 176 176 176 176 175 174 176 176	92/07/23 1426 k 92/07/23 1427 k 92/07/23 1428 k 92/07/23 1428 k 92/07/23 1429 k 92/07/23 1431 k 92/08/19 1130 k 92/08/19 1136 k 92/08/19 1136 k 92/08/19 1136 k 92/08/19 1136 k 92/08/19 1136 k 92/08/19 1136 k 92/08/19 1142 k 92/08/19 1144	IATER JATER JATER JATER JATER JATER JATER JATER JATER JATER	20 22 23 23.5 24.5 0.5 1.5 3 4.5 5.5	25233	23.8 23.5 23.5 23.4 23.2 26.0 26.0 25.8 25.4 25.4	2.1 1.4 1.1 .7 .6 10.7 10.6 9.9 9.4 7.9 7.7	24.75 16.15 12.65 8.05 130.55 130.55 129.35 120.75 114.65 94.05 91.75	6.90 6.80 6.70 6.70 8.90 8.90 8.90 8.80 8.70 8.30 8.30 8.20 8.00	176 166 162 157 192 192 192 192 193 196
	92/01/22 1223 WATER 92/01/22 1224 WATER 92/01/22 1225 WATER 92/01/22 1225 WATER 92/04/23 1210 WATER 92/04/23 1212 WATER 92/04/23 1214 WATER 92/04/23 1224 WATER 92/04/23 1224 WATER 92/04/23 1224 WATER 92/04/23 1224 WATER 92/04/23 1226 WATER	20 22 22.4 24 0.5 1.5 4 6 7 8 10 12	4067	6.0 6.0 18.9 18.2 17.5 16.2 15.2 14.8 13.8 13.2	11.4 11.3 11.4 10.9 10.9 10.9 10.9 10.9 8.8 8.4 8.0 7.9	91.28 90.48 91.24 121.38 114.78 114.78 112.48 102.08 91.28 86.38 80.88 75.58	7.50 7.50 8.70 8.80 8.80 8.70 8.80 8.70 8.10 7.90 7.80 7.80		92/08/19 1146 92/08/19 1146 92/08/19 1150 92/08/19 1152 92/08/19 1154 92/08/19 1156 92/08/19 1156 92/08/19 1200 92/08/19 1200 92/08/19 1206 92/08/19 1206 92/08/19 1206	WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER	6.5 8 10 12 14 16 18 20 22 22.5 24 25		25.2 24.6 24.6 24.5 24.1 24.1 23.8 23.8 23.8 23.8 23.8 23.8	6.8 5.1 5.1 4.8 4.3 4.0 3.7 3.7 3.6	81.05 70.25 60.75 60.05 51.85 50.65 47.15 43.55 43.55 43.55 43.55 43.55	7.70 7.40 7.30 7.30 7.20 7.20 7.20 7.20 7.20 7.20 7.20 7.2	197 198 199 200 200 200 200 200 200 200 200 200
B-27	02/04/23 1230 WATER 92/04/23 1232 WATER 92/04/23 1234 WATER 92/04/23 1236 WATER 92/04/23 1236 WATER 92/04/23 1238 WATER 92/05/14 1225 WATER 92/05/14 1227 WATER 92/05/14 1233 WATER 92/05/14 1235 WATER 92/05/14 1245 WATER 92/05/14 1245 WATER	14 16 20 22 24 0.5 1 1.5 3 4 5 6 8	17154	12.9 12.4 12.1 12.0 11.9 20.6 20.2 19.8 19.5 18.8 17.9 17.8 17.8	7.9 7.8 7.5 7.3 10.6 10.7 9.2 8.3 8.3 8.3	74.5\$ 72.2\$ 70.4\$ 67.6\$ 117.8\$ 115.2\$ 116.3\$ 108.5\$ 97.9\$ 87.4\$ 85.3\$ 77.3\$	7.60 7.60 7.50 8.70 8.70 8.40 8.40 8.40 8.40 7.90 7.90 7.90	143 139 137 137 142 161 161 161 161 161 161 159	92/09/23 1143 92/09/23 1143 92/09/23 1144 92/09/23 1145 92/09/23 1146 92/09/23 1146 92/09/23 1149 92/09/23 1150 92/09/23 1151 92/09/23 1152 92/09/23 1155 92/09/23 1155 92/09/23 1155	WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER WATER	0.5 1.5 5 7 9 11 13 15 17 21	26163	24.2 24.2 24.2 24.2 24.1 24.1 24.1 24.1	6.3 6.3 6.3 6.3 6.1 6.1 6.0 9 5.8 5.8 5.2	74.15 74.15 74.15 72.95 71.85 71.85 70.65 69.45 68.25 61.23	7.80 7.80 7.80 7.80 7.80 7.80 7.80 7.80	187 185 186 185 185 185 187 188
	92/05/14 1249 WATER 92/05/14 1255 WATER 92/05/14 1255 WATER 92/05/14 1258 WATER 92/05/14 1301 WATER 92/05/14 1301 WATER 92/05/14 1310 WATER 92/05/14 1310 WATER 92/06/17 1402 WATER 92/06/17 1408 WATER 92/06/17 1408 WATER 92/06/17 1411 WATER 92/06/17 1414 WATER 92/06/17 1414 WATER 92/06/17 1414 WATER 92/06/17 1414 WATER 92/06/17 1423 WATER 92/06/17 1423 WATER	10 12 14 16 20 25 0.5 1.5 3.4 4.3 7	37046	17.1 16.8 16.7 16.4 15.6 14.3 14.3 25.8 25.7 24.9 24.8 22.9 24.8 22.9 22.6 21.2 20.8	7.2 6.8 6.2 5.8 4.4 4.4 11.6 11.6 11.6 11.6 11.6 11.6 5.8 5.3	70.18 68.08 62.08 47.18 43.18 39.48 34.68 141.58 141.58 125.08 97.78 93.18	7,7(7,6(7,5(7,4(7,4(7,4) 9,1) 9,09 8,32 8,32 8,32 7,9 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,6(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,6) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7(7,7) 7,7() 7,7) 7,7) 173) 175) 172) 166) 162 0 159 0 150 0 150 0 155 0 173 0 173 0 173 0 173 0 172 0 172 0 172 0 172 0 172 0 172 0 172 0 172 0 172 0 172 0 172 0 172	92/09/23 1157 92/09/23 1158 92/09/23 1159	WATER	23 24 25		24.0 23.9	4.8 3.7	56.5 8 43.5 8	7.40 7.30	
	92/06/17 1425 WATER 92/06/17 1432 WATER 92/06/17 1432 WATER 92/06/17 1438 WATER 92/06/17 1438 WATER 92/06/17 1444 WATER 92/06/17 1450 WATER 92/06/17 1450 WATER 92/06/17 1450 WATER 92/06/17 1450 WATER 92/06/17 1450 WATER 92/06/17 1502 WATER 92/07/23 1411 WATER 92/07/23 1413 WATER 92/07/23 1413 WATER 92/07/23 1415 WATER 92/07/23 1415 WATER 92/07/23 1415 WATER 92/07/23 1415 WATER	6 8 10 12 14 16 18 20 22 22 4 25 0.5 1 1.5 4 4.55 5 5	19229	20.3 20.2 20.1 20.0 19.9 19.6 19.6 19.5 27.3 27.3 27.2 27.0 27.1 26.1 27.1 26.1 27.1	5 5 4 3 3 7 6	2 56.58 1 55.48 5 54.38 8 53.228 8 53.228 8 53.238 8 53.238 8 53.238 8 53.238 8 53.238 8 53.238 8 53.238 8 53.238 1 18 8 7 51.18 8 7 51.28 8 7 7 95.18 8 7 7 95.18 8 7 7 95.18 8 7 7 7 95.18 8 7 7 7 8 7 7 7 8 7 7 7 8 7 7 7 7 7 7 7	7.3 7.2 7.1 7.1 7.1 7.1 8.8 8.8 8.8 8.8 8.8 8.8	0 170 0 169 0 170 0 170 0 170 0 170 0 170 0 157 0 157 0 157 0 157 0 157 0 157 0 177 50 177 50 177 50 177									
	92/07/23 1416 WATER 92/07/23 1418 WATER 92/07/23 1418 WATER 92/07/23 1420 WATER 92/07/23 1420 WATER 92/07/23 1422 WATER 92/07/23 1422 WATER 92/07/23 1424 WATER 92/07/23 1424 WATER	5.5 6 7 8 10 12 14 16		25.9 25.6 24.9 24.5 24.3 24.2 24.3 24.2 24.1 23.5	4.3.3.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2.	2 51.2 6 42.9 1 36.5 0 35.3 9 34.1 8 32.9 5 29.4	7. 7. 7. 7. 7. 7. 7. 5. 7. 5. 7.	40 17 10 17 00 17 00 17 00 17 00 17					·				

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06010201002 0002.040 ON

92/01/22 1225 WATER 22.4 92/04/23 1210 WATER 22.4 120 10K 120 10K 120 10K	.56
92/04/23 1218 VERT 4 1.77 10K .100 .040	
92/05/14 1225 WATER 0.5 92/05/14 1239 VERT 0.5 1.40 10K 10 5 .240 .060	.21 .31
92/06/17 1420 VERT 0.5 92/06/17 1420 VERT 4 D1 92/06/17 1421 VERT 4 D1 92/06/17 1421 VERT 4 D2 15 3 (70)	- 13 - 41
92/06/17 1426 VATER 22.4 D1 15 4 .630 .010 92/06/17 1457 WATER 22.4 D1 15 4 .490 .020 92/06/17 1457 WATER 22.4 D1 15 4 .450 .020 92/06/17 1458 WATER 22.4 D2 10 10 .400 .020	.12 .13 .12
92/07/23 1410 WATER 0.5 92/07/23 1414 VERT 4 1.38 10K 15 10 .300 .040 92/07/23 1427 WATER 22 15 3 100	.36 .43 .38
92/08/19 1140 VERT 4 1.43 10K 10 6 250 .020 92/08/19 1206 WATER 22.5 10 10 3 (00	.10 .40
92/09/23 1147 VERT 4 1.47 10K 10 9 .390 .010 92/09/23 1157 WATER 23 15 3 .310 .040	.11 .38 .12
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92/04/23 1238 VERT 4 .050 .010 1.8 0.00 1.00K 1.00 1 92/04/23 1238 WATER 22 .020 .002K 2.0 6.00 1.00K 1.00 1 92/05/14 1239 VERT 4 .030 .004 1.7 6.00 1.00 1.00 3	.00K .00
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476041 1114C 35 49 50.0 084 36 33.0 2 WATTS BAR RESERVOIR 47145 TENNESSEE ROANE TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 560.80 1317VAC NG 06010201002 0043.170 OFF 0000 METERS DEPTH

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r	DATE FROM TO 92/01/22	TIME OF DAY 1104	MEDIUM	SMK OR DEPTH (M)	84002 CODE GENERAL REMARKS	00078 TRANSP Secchi Meters	31616 FEC COLI MFM-FCBR /100ML	00080 COLOR PT - CO UNITS	00530 RESIDUE TOT NFL1 MG/L	OO605 Org n N Mg/L	00610 NH3+NH4- N TOTAL	00630 N02&N03 N-TOTAL
	92/01/22	1108	VERT	0.5		1.10	10K			MG/L	MG/L	MG/L
	92/04/23 92/04/23 92/04/23 92/05/14	1105 1113 1123	WATER VERT	11.3 0.5 4 11		1.23	10K	10		.170 .190		. 72 . 71
	92/05/14 92/05/14 92/06/17 92/06/17 92/06/17	1054 1059 1101 1113	VERT WATER WATER VERT	12 4 0.5 0.5 4		1.20 .80	10K 10k	10 5 10	10	.430 .250 .290 .360		. 21 . 37 . 27 . 18
, ,	92/07/23 92/07/23 92/07/23	1240	VERT	12.3 0.5 4 12.4		- 98	10	10 10 15	17	.250.180	.020 .050	- 38 - 40
,	92/08/19 1 92/08/19 1 92/08/19 1 92/08/19 1	11 4 4	VENT	0.5		1.27	10K	10	7 . 14	-600 -250	.010 .040	.27 .34
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			.2 [°]	••				15 15	4 6	-240 -550	.020 .020	- 30 - 31
		108 1	MEDIUM	SMK OR DEPTH (M) 4	84002 CODE GENERAL REMARKS	00665 Phos-tot Mg/l P .040	00671 PHOS-DIS 1 ORTHO MG/L P	00680 ORG C C MG/L	32211 Chlrphyl A Ug/l Correctd	32212 Chlrphyl B Ug/l	32214 Chlrphyl C Ug/l	32218 PHEOPHIN A UG/L
	92/04/23 11 92/04/23 11	13 V	EDT .	11.3 4		.040	.020	1.9 1.9	4.00	1.00K	1.00	1.00
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	92/08/19 10: 92/08/19 10:	33 VE	ERT	12.4		.020	.007 .008 .003	2.3	8.00	1.00	1.00	2.00
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TENNESSEE VALLEY AUTHORITY

Resource Group Water Management Clean Water Initiative

TENNESSEE VALLEY RESERVOIR AND STREAM QUALITY, 1993 PHYSICAL AND CHEMICAL CHARACTERISTICS OF WATER RESERVOIR VITAL SIGNS MONITORING

Prepared by

Tammy L. Carroll, Joseph P. Fehring, and Dennis L. Meinert

Chattanooga, Tennessee June 1994

INTRODUCTION

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The Tennessee Valley Authority (TVA) initiated a Reservoir Monitoring Program in 1990 as part of its Water Resources and Ecological Monitoring Activities. In these first four years (1990-1993), the Reservoir Monitoring Program has undergone continual change and improvement. Initially, in 1990, only 12 TVA reservoirs were examined, the nine mainstream Tennessee River reservoirs (Kentucky through Fort Loudoun), and three major tributary storage reservoirs: Cherokee, Douglas, and Norris (Dycus and Meinert, 1991). In 1991, the Reservoir Monitoring Program was expanded to 24 reservoirs, to include the system's only two tributary reservoirs with navigation locks (Tellico and Melton Hill) and ten other smaller tributary reservoirs (Dycus and Meinert, 1992). In 1993, six additional tributary reservoirs were added (Dycus and Meinert, 1994). No further expansion of reservoir monitoring is planned.

The two objectives of the Reservoir Monitoring Program are to provide basic information on the "health" or integrity of the aquatic ecosystem in TVA reservoirs (referred to as Vital Signs Monitoring) and to provide screening level information for describing how well each reservoir meets the "fishable" and "swimmable" goals of the Clean Water Act (referred to as Use Suitability Monitoring).

The basis of Vital Signs Monitoring is examination of appropriate physical, chemical, and biological indicators monitoring tools - at one or more strategic locations in each

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reservoir, i.e. the forebay immediately upstream of the dam; the transition zone (the mid-reservoir region where the water changes from free flowing to more quiescent, impounded water; and the inflow or headwater region of the reservoir. The monitoring tools comprised: basic physical/chemical water quality sampling; sediment quality and toxicity testing; benthic macroinvertebrate community evaluations; and fish community evaluations. A summary report (Dycus and Meinert, 1994) presents results of TVA's 1993 Reservoir Monitoring Program, to include all Vital Signs (physical/chemical, sediment, benthic and fish communities) and Use Suitability Monitoring.

This technical data summary presents the 1993 Vital Signs physical/chemical water quality sampling information only. The basic group of 30 reservoirs were monitored. Embayment monitoring sites were added in 1993 on four reservoirs: the Big Sandy River embayment of Kentucky Reservoir; the Bear Creek embayment of Pickwick Reservoir; the Elk River embayment of Wheeler Reservoir; and the Hiwassee River embayment of Chickamauga Reservoir. Transition zone sites were moved somewhat downstream on Tellico and Douglas Reservoirs, to capture a less riverine environment; and the transition zone site was moved slightly upstream on Watauga Reservoir, to differentiate that site more from the forebay site. Further, the forebay site on Nottely Reservoir was moved upstream; and on Chatuge, a site was added on Shooting Creek.

MATERIALS AND METHODS

Reservoir Characteristics - The physical characteristics of a reservoir (volume, surface area, depth, hydraulic residence time, etc.) have a great effect on its intrinsic physical and chemical processes and water quality characteristics. The Vital Signs reservoirs are very broadly categorized as either run-of-the-river or tributary reservoirs, with generally large differences in their morphologic and hydraulic characteristics (Table 1). Primary differences include the greater depths and longer hydraulic residence times of tributary reservoirs, resulting in the development of strong thermal stratification and its associated physical/chemical processes. Short average residence times found on most of the main stream, run-of-the-river reservoirs (usually less than 30 days) results in well-mixed riverine conditions. Thermal stratification of the run-of-the-river reservoirs rarely exists and then for only short periods of time under conditions of low flow and intense solar heating. However, tributary reservoirs with their longer residence times (typically greater than 100 days) develop strong thermal stratification that begins in the spring and ends with seasonal cooling and mixing in the late autumn. During this summer stratification, inflowing water may be cooler than the epilimnetic surface water and may enter the reservoir as a cold underflow to the hypolimnion rather than mixing with the epilimnion. This results in an even longer residence time of the epilimnetic water than is indicated by the calculated average

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CHARACTERISTICS OF VITAL SIGNS RESERVOIRS

Average

Reservoir Name	Drainage Area <u>(sq. milcs)</u>	Reservoir Length [*] (miles)	Surface Area ^a (acres) 1000's	Depth at Dam [*] _(ft)	Volume [*] <u>(ac-ft)</u> 1000's	Average Annual Drawdown ^b _(ft)_	Average Reservoir Flow-POR _(cfs)_	Hydraulic Residence Time-1993	CY 1993 Reservoir Flow <u>(cfs)</u>		
			1000 3	Run-of-the-F	liver Reserve	oirs					
	40.000	184.3	160.3	88	2,839	5	66,600	27.5	52,097		
Kentucky	40,200	52.7	43.1	84	924	6	54,900	9.6	48,566		
Pickwick	32,820	15.5	15.5	108	634	3	51,500	6.8	47,236		
Wilson	30,750	13.3 74.1	67.1	66	1,050	6	49,400	11.4	46,264		
Wheeler	29,590	75.7	67.9	65	1,018	2	40,700	12.9	39,691		
Guntersville	24,450	46.3	10.4	60	241	0	35,900	3.6	34,092		
Nickajack	21,870	58.9	35.4	83	628	7	34,200	9.6	32,887		
Chickamauga	20,790	72.0/24.0°	39.0	105	1,010	6	27,100	19.5	26,145		
Watts Bar	17,300	50.0	14.6	94	363	6	18,400	9.7	18,897		
Fort Loudoun	9,550	44.0	5.7	69	120	0	4,920	12.7	4,764		
Melton Hill	3,343	33.2	16.5	80	415	6	6,300 ^ª	34.0	6,159 ^d		
Tellico	2,627 55.2 Tolly Tributary, Storage Reservoirs										
	0.010	73.0/53.0°	34.2	202	2,040	32	4,190	249.4	4,124		
Norris	2,912	54.0	30.3	163	1,481	28	4,460	162.2	4,604		
Cherokee	3,428		30.4	127	1,408	48	6,780	109.4	6,490		
Douglas	4,541	43.1	0.9	81	27	0	2,650	5.6	2,423		
Ft Patrick Henry	1,903	10.4	4.3	129	189	25	2,550	38.5	2,477		
Boone	1,840	17.4/15.3°	4.3 7.6	239	658	33	976	341.3	972		
South Holston	703	23.7	7.0 6.4	239	569	26	714	403.5	711		
Watauga	468	16.3		460	1,420	64	3,840	173.5	4,126		
Fontana	1,571	29.0	10.6	255	422	45	2,020	98.8	2,154		
Hiwassee	968	22.2	6.1	124	. 234	10	459	291.3	405		
Chatuge	189	13.0	7.0	124	170	24	416	228.0	376		
Nottely	214	20.2	4.2		85	24 7	1,420	33.1	1,296		
Ocoee #1 (Parksville)	595	7.5	1.9	115	193	36	614	156.2	623		
Blue Ridge	232	11.0	3.3	156	530	12	940	328.7	813		
Tims Ford	529	34.2	10.6	143	10	12 11°	380	14.4	337		
Bear Creek	232	16.0	0.7	74	94	14°	282	185.7	255		
Cedar Creek	- 179	9.0	4.2	79		14 12°	101	253.9	90		
Little Bear Creek	61	7.1	1.6	82	45	12 1°	14	616.2	9		
Beech	16	5.3	0.9	32	11		320		275		
Normandy	195	17.0	3.2	83	110	11	520	201.7			

Measurements based on normal maximum pool.
 Tennessee River and Reservoir System Operation and Planning Review, Final EIS, TVA/RDG/EQS-91/1, 1990.
 Major/minor arms of reservoir.
 Estimated flow based on releases from Chilhowee Dam (POR avg. = 4770cfs), and adjusted based on the additional drainage area between Chilhowee Dam (1977 sq miles) and Tellico Dam (2627 sq miles).
 Estimated based on difference between normal maximum summer pool and average minimum winter pool elevations.

Data Source: Hydrologic Data Management (Knoxville, TN), Systems Engineering, TVA, 1994. Same and the

reservoir volume/flow quotient, particularly if the turbine power intakes draw water exclusively from below the thermocline.

A major effect of this thermal stratification in tributary reservoirs is the depletion of hypolimnetic dissolved oxygen. The pattern of hypolimnetic dissolved oxygen depletion is variable both spatially and temporally for any one reservoir and among reservoirs. The general pattern, however, is that after the onset of thermal stratification, an anoxic zone first begins to develop in the transition zone of the reservoir. The large input and settling out of allochthonous organic matter (leaves and detritus washed in by the tributary streams) causes high rates of oxygen uptake in the transition zone and results in the initial depletion of dissolved oxygen in this area of the reservoir. The zone of anoxic water gradually extends both upstream and downstream, until it reaches the free-flowing river upstream and the dam downstream. At the same time the anoxic zone develops vertically and laterally. In the worst case, the entire hypolimnion may become anoxic and remain so until reaeration during autumn overturn. The hypolimnetic anoxia promotes the dissolution and release of metals, minerals, and nutrients from the reservoir sediment and bottom water, resulting in elevated concentrations of reduced forms of iron, manganese, sulfate, nitrate, and phosphate. When reservoir destratification occurs in autumn, the hypolimnion is mixed with surface water and replenished with oxygen.

Epilimnetic processes are also affected by thermal stratification, particularly in deeper tributary reservoirs. With

the onset of spring stratification, primary production can be quite high, particularly in the transition zone. The phytoplankton community frequently receives ample supplies of nutrients from periodic spring rainfall/runoff events, and with lower water velocities and increased rates of sedimentation, improved water clarity and light penetration often results in episodes of high algal productivity as evidenced by high chlorophyll-a concentrations, high pH values and supersaturation The longer residence times of tributary of dissolved oxygen. reservoirs results in water clarity usually being more a function of algal cell abundance and chlorophyll pigment and less a function of inorganic suspended material as is the case in mainstream reservoirs with shorter residence times and higher velocities. In mainstream reservoirs, the shorter detention times and higher velocities inhibit the deposition of suspended material, resulting in generally lower water clarity and light availability and often limiting algal productivity.

Epilimnetic water clarity often increases moving downstream into the forebay area of the reservoir and into the late summer as rainfall/runoff events occur less frequently. Even though water clarity increases, nutrients become depleted and primary production often is reduced as much of the organic matter produced during photosynthesis decomposes and settles below the photic zone. Bacterial decomposition consumes available dissolved oxygen (respiration) and releases carbon dioxide, thereby lowering the pH. Unlike mainstream reservoirs, this decomposition process in tributary reservoirs often results in two oxygen minima in the

water column - one in the epilimnion at or near the thermocline and one in the hypolimnion.

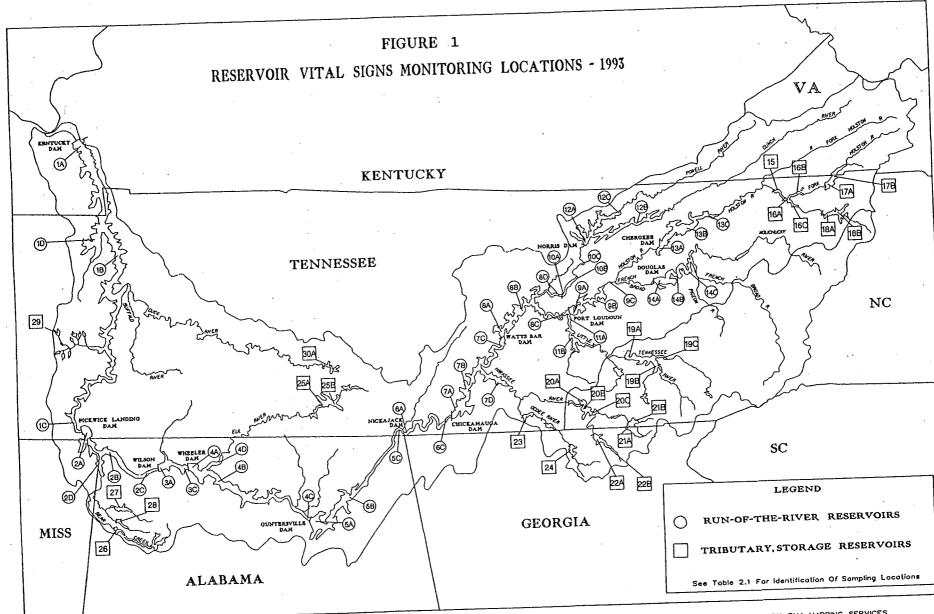
Consequently, physical/chemical differences in water quality between run-of-the-river and tributary reservoirs are expected in examining the 1993 Vital Signs data. In the Results and Discussion section of this report, water quality characteristics are discussed and comparisons are made among the Vital Signs reservoirs to point out these differences. Because physical/chemical water quality conditions are different, this does not imply that one reservoir is good and another is poor, it merely is a reflection of the unique physical and chemical differences between each. For example, one particular reservoir, Fort Loudoun, exhibits both run-of-the-river and tributary reservoir characteristics. Although it is located on the main stream of the Tennessee River, it is the most upstream reservoir and is formed by the confluence of the French Broad and Holston Rivers. Major tributary reservoir impoundments on both these river systems, Douglas Dam at mile 32.3 of the French Broad and Cherokee Dam at mile 52.3 on the Holston, release cool, hypolimnetic water low in dissolved oxygen during the summer. Depending upon the quantity and duration of water released from Douglas and Cherokee dams and the temperature of the impounded Fort Loudoun water, this cool water may flow under the warmer water impounded in Fort Loudoun Reservoir resulting in thermally stratified conditions. Consequently, Fort Loudoun can intermittently exhibit the characteristics of either a run-of-theriver reservoir or a tributary reservoir.

Overview - Physical/chemical variables were measured at a total of fifty seven locations on the thirty Vital Signs reservoirs (Figure 1, Table 2). The Vital Signs water quality monitoring activities on these reservoirs, followed either a "basic" or "limited" sampling strategy (Tables 2 and 3). The basic sampling strategy included monthly water quality surveys (April through September) at the forebay and transition zone (and on four reservoirs, an embayment--see Table 2) locations on eleven TVA reservoirs: the nine mainstem Tennessee River reservoirs; and Melton Hill and Tellico reservoirs. The limited sampling strategy included monthly water sampling (April through October) for a smaller list of parameters at the forebay locations (and at midreservoir locations on larger reservoirs) on nineteen tributary impoundments.

Water quality measurements, sample collections and sample handling followed standard practices accepted by the Environmental Protection Agency (EPA, 1979 and 40 CFR 136), US Geological Survey (USGS, 1977), and American Public Health Association (AWWA, WPCF, APHA, 1989) as specified in TVA quality assurance/quality control (QA/QC) procedures (TVA, 1990 and TVA, 1987). TVA laboratory analyses conformed to established EPA, USGS, and APHA QA/QC procedures (TVA, 1989b).

Details on the physical/chemical analyses and measurements on the water samples are summarized in Tables 3 and 4. The 1993 data are tabulated in the Appendix A - Physical/Chemical Characteristics of Water. All the data are stored and are

available on EPA's water quality data storage and retrieval (STORET) computer system.



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PRODUCED BY TVA MAPPING SERVICES

Table 2

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WATER QUALITY MONITORING LOCATIONS RESERVOIR VITAL SIGNS MONITORING, 1993

Basic Water Quality Monitoring Locations

Reservoir	Sampling <u>Locations</u> a	STORET ID No.	<u>Description</u> b
Kentucky	TRM 23.0 TRM 85.0 TRM 200-206	202832 477403 	1A-FB 1B-TZ 1C-I
Pickwick	Big Sandy 7.4 TRM 207.3 TRM 230.0	477210 476799 016923	1D-E 2A-FB 2B-TZ
Wilson	TRM 253-259 Bear Cr. 8.4 TRM 260.8 TRM 273-274	 017849 016912	2C-I 2D-E 3A-FB 3C-I
Wheeler	TRM 277.0 TRM 295.9 TRM 347-348	016900 017009	4A-FB 4B-TZ 4C-I
Guntersville	Elk River 6.0 TRM 350.0 TRM 375.2 TRM 420-424	017850 017261 017522	4D-E 5A-FB 5B-TZ
Nickajack	TRM 425.5 TRM 469-470	476344	5C-I 6A-FB 6C-I
Chickamauga	TRM 472.3 TRM 490.5 TRM 518-529	475358 475265	7A-FB 7B-TZ 7C-I
Watts Bar	Hiwassee 8.5 TRM 531.0 TRM 560.8 TRM 600-601	477512 475317 476041	7D-E 8A-FB 8B-TZ 8C-I
Fort Loudoun	CRM 19-22 TRM 605.5 TRM 624.6 TRM 652	477404 475603	8D-I 9A-FB 9B-TZ 9C-I
Melton Hill	CRM 24.0 CRM 45.0 CRM 59-66	477064 476194	10A-FB 10B-TZ 10C-I
Tellico	LTRM 1.0 LTRM 15.0 LTRM 21.0	476260 476456 476295	11A-FB 11B-TZ
Limited	Water Quality Moni	toring Locatio	ns
Norris	CRM 80.0 CRM 125.0	476009 477186	12A-FB 12B-MR
Cherokee	PRM 30.0 HRM 53.0 HRM 76.0 HRM 91	477187 475025 475028	12C-MR 13A-FB 13B-MR
Douglas	FBRM 33.0 FBRM 51.0 FBRM 61	475081 477510	13C-I 14A-FB 14B-MR 14C-I

Table 2 (Continued)

WATER QUALITY MONITORING LOCATIONS RESERVOIR VITAL SIGNS MONITORING, 1993

Limited Water Quality Monitoring Locations

Reservoir	Sampling <u>Locations</u> a	STORET ID No.	Descriptionb
Ft. Patrick Henry Boone	SFHR 8.7 SFHR 19.0 SFHR 27.0 WRM 6.5	477509 475858 476221 477511	15-FB 16A-FB 16B-MR 16C-MR
South Holston	SFHR 51.0 SFHR 62.5	475859 475573	17A-FB 17B-MR/I
Watauga	WRM 37.4 WRM 45.5	475576 477513	18A-FB 18B-MR
Fontana	LTRM 62.0 LTRM 81.5	370004 370177	19A-FB 19B-MR
Hiwassee	TkRM 3.0 HiRM 77.0 HiRM 85.0	370162 370001 370154	19C-MR 20A-FB 20B-MR 20C-I
Chatuge	HiRM 90 HiRM 122.0 Shooting Cr. 1	370003 1.5 370178	21A-FB 21B-FB
Nottely	NRM 23.5 NRM 31.0	120883 120806	22A-FB 22B-MR
Ocoee No. 1	ORM 12.5 ORM 16.5	475684	23-FB
Blue Ridge Tims Ford	TORM 54.1 ERM 135.0 ERM 150.0	130032 477072 475768	24-FB 25A-FB 25B-MR
Bear Creek Cedar Creek Little Bear Creek Beech Normandy	BCM 75.0 CCM 25.2 LBCM 12.5 BRM 36.0 DRM 249.5	017041 017233 017474 475876 477453	26-FB 27-FB 28-FB 29-FB 30-FB

- BRM Beech River Mile BCM - Bear Creek Mile а. CRM - Clinch River Mile CCM - Cedar Creek Mile ERM - Elk River Mile DRM - Duck River Mile HiRM - Hiwassee River Mile FBRM - French Broad River LBCM - Little Bear Creek Mile HRM - Holston River Mile LTRM - Little Tennessee River NRM - Nottely River Mile PRM - Powell River Mile ORM - Ocoee River Mile TRM - Tennessee River Mile SFHR - S. Fork Holston River TkRM - Tuckasegee River Mile TORM - TOCCOA River Mile PRM - Powell River Mile WRM - Watauga River Mile
- b. Numbers are keyed to Figure 1. FB forebay; TZ transitic zone; MR mid-reservoir; I Inflow; and E embayment. MR means that sampling location was referred to as an inflow location in the fish community evaluation (sampling done in autumn at lower reservoir water level elevations), and as a mid-reservoir location in the evaluation of the water qualit data (sampling done in summer at higher water level elevations).

Table 3 RESERVOIR VITAL SIGNS - 1993 WATER QUALITY MONITORING STRATEGY

Description	Sample Collection <u>Depths</u>	Monitor <u>Basic</u>	ing Strategy ^a <u>Limited</u>
Field Measurements Hydrolab/ (temperature, pH, dissolved oxygen, and conductivity)	0.3,1.5,4,6,8 ^b etc.	monthly	monthly
Secchi Depth		monthly	monthly
Fecal Coliform	0.3 (surface grab)	monthly	NA
Turbidity	composite (& bottom) ^C	monthly	monthly
Hydrogen Sulfide	bottom ^C	as needed ^c	i
Laboratory Measurements			
Chlorophyll-a	composite ^e	monthly	monthly
Nutrients			
Organic Nitrogen	composite (& bottom) ^C	monthly	April & August
Ammonia Nitrogen	composite (& bottom)	monthly	April & August
Nitrite & Nitrate Nitrogen	composite (& bottom)	monthly	April & August
Total Phosphorus	composite (& bottom)	monthly	April & August
Dissolved Ortho phosphorus	composite (& bottom)	monthly	April & August
Organic Carbon			
Total Organic Carbon	composite (& bottom)	monthly	April & August
Color and Solids			
Color	composite (& bottom)	monthly	NA
Suspended Solids	composite (& bottom)	monthly	NA

^a Basic monthly and April through September. Limited monthly is April through October, with some variables measured only in April and August.

^b <u>In situ</u> Hydrolab measurements were made at the depths indicated and at regularly spaced intervals (2-4 meters) from the water surface to the bottom of the water column. These measurements were also made at intermediate depths any time the temperature changed by more than 2°C or the dissolved oxygen changed by more than 1 mg/l.

- ^c Bottom grab samples were only collected as part of the basic sampling strategy, except for hydrogen sulfide. Bottom indicates a grab sample collected three meters above the bottom at forebay locations and one meter above the bottom at transition zone locations.
- d When low dissolved oxygen was measured near the bottom of the reservoir (i.e., less than 1 mg/l), the bottom water grab sample was smelled to check for the presence of hydrogen sulfide (rotten egg odor). If detected, the concentration was measured.
- ^e Composite indicates a photic zone composite sample with the photic zone defined as four meters or twice the Secchi depth, whichever is greater.

Table 4

RESERVOIR VITAL SIGNS MONITORING - WATER PHYSICAL/CHEMICAL MEASUREMENTS, 1993

EPA Storet <u>Code</u>	Description	Units	Detection <u>Limits</u>
	Field Measurer	nents	
00010 00300 00400 00094	Hydrolab Temperature Dissolved Oxygen pH Conductivity	°C mg/l Std. units micromhos	
00078	Secchi Depth	meters	0.1 meters
31616	Fecal Coliform	colonies/100mL	1/100 mL
82078	Turbidity	NTU	1 NTU
00745	Hydrogen Sulfide	mg/l	0.00025
	Laboratory Meas	urements	
32211	Chlorophyll-a	ug/l	1 ug/l
00605 00610 00630 00665 00671	Nutrients Organic Nitrogen Ammonia Nitrogen Nitrite & Nitrate Nitrogen Total Phosphorus Dissolved Ortho phosphorus	mg/l mg/l mg/l mg/l mg/l	0.02 mg/l 0.01 mg/l 0.01 mg/l 0.002 mg/l 0.002 mg/l
00680	Organic Carbon Total Organic Carbon	mg/l	0.2 mg/l
00080 00530	Color and Solids Color Suspended Solids	PCU mg/l	1 PCU 1 mg/l

HYDROLOGIC OVERVIEW OF WATER YEAR 1993

Many water quality characteristics (e.g., temperature, dissolved oxygen conductivity, water clarity, suspended solids, etc.) exhibit changes due to seasonal variations in atmospheric temperature and rainfall. During those times of year when runoff is minimal (normally August-October), streamflow is largely derived from the base flow of groundwater. Because of greater contact between the water and the soil/rock and the longer groundwater residence times, groundwater contains more dissolved minerals (i.e., higher concentrations of hardness and alkalinity, higher pHs and conductivities, etc.) than does surface water. During those times of the year when runoff is higher (normally January-March), streamflow is principally derived from rapid overland runoff that allows little time for mineral dissolution.

Consequently, during those times of the year with higher rainfall and subsequent higher flows, base flow accounts for a smaller proportion of the total streamflow, resulting in lower concentrations of most dissolved constituents. In addition, periods of intense rainfall and high overland flows wash off or "flush" a watershed and transport soil particles to streams, often carrying large loads of nonpoint source pollutants (nutrients, suspended solids, fecal bacteria, etc.) to streams and rivers.

In addition to flood control, electric power generation, and navigation, an important benefit of the TVA's system of dams and reservoirs is its ability to maintain adequate streamflow during extended periods of low rainfall and low runoff by the controlled

release of water from tributary storage impoundments. However, this alteration of natural streamflow (diminishing high flows during floods and augmenting low flows during droughts) by storing and then slowly releasing water from tributary storage impoundments creates conditions of strong thermal stratification and low dissolved oxygen in the bottom waters of these tributary storage impoundments.

From a water quality perspective, the lower streamflows occurring during the warmer summer months, combined with naturally occurring higher water temperatures and lower dissolved oxygen concentrations, result not only in lakes becoming thermally stratified but also having less water and less oxygen available to dilute and assimilate the wastes discharged to them. In addition, the warmer water temperatures increase aquatic biological processes (respiration, bacteriological decomposition, etc.). This results in oxygen being used at a faster rate, which can further lower oxygen concentrations. In combination, these factors (low streamflows and diminished assimilative capacity, warmer temperatures and higher biological oxygen consumption rates, and the inhibition of mixing and reaeration caused by thermal stratification) result in low dissolved oxygen concentrations and adversely impact the health of aquatic life. The summer of 1993 was a case in point. July 1993 was the hottest month on record (since 1890s) in the Tennessee Valley. Valley-wide temperatures averaged almost 83°F (28.3°C), about 5°F (2.8°C) above normal for For example, in Chattanooga, all 31 days in July had July. temperatures above 90°F (32.2°C), with temperatures up to 104°F

(40.0°C) and 15 days with temperatures 98°F (36.7°C) or higher. This record-breaking heat (and low streamflows) resulted in high water temperatures in the Tennessee River. In fact, all nine mainstem Tennessee River reservoirs had surface water temperatures that exceeded 86°F (30.0°C), some with highs up to 90°F (32.2°C).

In addition, Tennessee Valley rainfall and runoff were well below normal in the summer of 1993. In July, Valley-wide rainfall averaged only 1.76 inches (45mm), a deficit of 3 inches (76mm) below the long-term July mean of 4.77 inches (121mm). As a result, rainfall runoff was only 0.66 inches (17mm), compared to the long-term July mean of 1.03 inches (26mm). Further, runoff was significantly lower in the western half of the Tennessee Valley than in the eastern half. In July, runoff above Chattanooga was 90 percent of the long-term mean, while runoff was only 64 percent of the long-term mean above Kentucky Dam. For the period of January through July, runoff above Chattanooga was 80 percent of the long-term mean, while runoff was 72 percent of the long-term mean above Kentucky Dam. Consequently, flows in the Tennessee River in 1993 increasingly fell below the long-term average as the river flowed downstream from Fort Loudoun Dam to Kentucky Dam.

The high temperatures and low flows of July 1993 adversely impacted dissolved oxygen concentrations in the Tennessee River, particularly in the downstream reservoirs. In mid-July, hypolimnetic anoxia (DOs equal to 0 mg/L) was found in the forebays of Kentucky, Pickwick, Wilson, Wheeler, and Chickamauga Reservoirs. All-time low concentrations of DO were recorded in

the releases from Chickamauga Dam on July 16 (2.2 mg/L) and Nickajack Dam on July 19 (1.8 mg/L) when flows from both dams were only 9000 cfs. During the first two weeks of July (July 1 to 15), daily flows averaged only about 17,250-17,500 cfs at Chickamauga and Nickajack Dams, or about 55 percent of the normal flow for this period of time. Once the effects of the high temperatures and low flows on DOs in the Tennessee River were recognized, flows were immediately increased (by drawing water from tributary storage reservoirs) and DO concentrations improved. For example, at Chickamauga Dam, from July 16-31, average daily flows were increased to an average of about 24,500 cfs (about 80 percent of the normal flow for July) and DOs in the releases increased to an average of about 4.3 mg/L, ranging from 3.2 to 6.3 mg/L. Compounding this whole situation were the record-setting rains and flooding occurring in the mid-West along the Mississippi and Missouri Rivers during the "flood of the century." During this period, TVA minimized discharge from the Tennessee River through Kentucky Dam so as to not increase flood crests on the lower Ohio and Mississippi Rivers and worsen the already catastrophic flooding in those areas.

Obviously, examining atmospheric temperature, rainfall, and runoff patterns during 1993 aids in interpretation of the Vital Signs monitoring data and the ecological health assessments of the streams and reservoirs.

Atmospheric Temperature

Average annual temperature in the TVA region is approximately 60 degrees Fahrenheit, °F (15.6 degrees Celsius, °C), with January usually being the coldest month and July the hottest. According to U.S. Department of Commerce (USDOC) climatic data, atmospheric temperatures in the TVA region averaged only about 0.3°F (0.2°C) warmer than normal in 1993; however, 1993 was a year of extremes (USDOC, 1993). January and July were unusually warm with 5.0°F (2.8°C) and 4.7°F (2.6°C) above normal, respectively; while, March and April were below normal with departures greater than -2.0°F (-1.1°C) (Figure 2a).

In review, 1993 began with an unusually warm January but cooled to below normal in February. As has often occurred in the last 15 years, another cold spring with late freezes was experienced. A record-breaking late season blizzard struck the Valley in mid-March and hit hardest in the eastern half. Summer was hotter than normal, with Tennessee, Alabama, Georgia, North Carolina, and Virginia all having the hottest July on record since the 1890s. The persistent heat and high humidity created great stress on livestock and people. The daily records for Chattanooga Airport provide an indication of the unusual conditions. All 31 days had maximums above 90°F (32.2°C), with the observed maximums ranging from 92°F (33.3°C) to 104°F (40°C) and 15 days of 98°F (36.7°C) or higher. The last four months had near or below normal temperatures, and the annual average temperature was only slightly above normal.

<u>Rainfall</u>

The Tennessee River basin averages about 51-52 inches (1295-1320 millimeters (mm) of precipitation annually. However, there are large variations in the spatial distribution of precipitation. The range is from a high of about 93 inches (2360mm) in the mountains of southwestern North Carolina near Highlands, North Carolina, to a low of about 37 inches (940mm) in the shielded valleys of these same mountains near Asheville, North Carolina. Elsewhere in the Valley, precipitation usually ranges within five to ten inches (127mm to 254mm) of the basin average. March is usually the wettest month and October the driest.

Rainfall across the Tennessee Valley in 1993 averaged only 39.8 inches (1011mm), almost 12 inches (about 300mm) or 23 percent less than the long-term 100-year average. The diminished rainfall in 1993 followed another dry year, 1992, when annual rainfall was about 8 inches (204mm) or about 15 percent below the long-term average. The period January-May 1992 ranked as one of the ten driest on record in the Tennessee Valley. During 1993, only the month of December had rainfall greater than normal (6.1 inches [155mm]) compared to normal December rainfall of 4.8 inches (122mm); the greatest rainfall deficit occurred in July (1.8 inches [45mm] compared to the normal July rainfall of 4.8 inches [122mm]). In addition to the extremes of December and July, March and September precipitation was close to average while February, April, June, and October were more than an inch (254mm) below

average (figure 2b). During March 1993, the Tennessee Valley received the equivalent of 5.4 inches (137mm) of rain, much of this during the "Winter Snow Storm of the Century" when many areas received record amounts (greater than 20 inches [about 500mm]) of snowfall.

The unusually persistent hot weather and below average rainfall in the summer was related to an unusual upper air pattern, which kept the storm track well west and north of the region and allowed very few cold fronts to reach the Tennessee This nearly stationary position of a strong upper air Valley. trough over the Rocky Mountains was associated with the record flooding in the middle of the country and kept the Southeast hot and dry. This general pattern was most persistent in the summer, but frequently alternated with a pattern having an upper trough over or to the east of the Valley in the other seasons. This latter trough kept most storms associated with it to the south of the TVA region. These two upper air patterns dominated the weather during 1993, so significant rainfall events tended to occur only when there was a transition period between one and the other.

<u>Streamflow</u>

Streamflow varies seasonally with rainfall, although during the spring and summer evaporation and transpiration also significantly reduce the amount of runoff. Watersheds that receive 50 to 60 inches (1270 to 1524mm) of precipitation annually

average about 20 to 30 inches (508 to 762mm) of runoff. In a normal year, the discharge of the Tennessee River (approximately 66,000 cfs [1868 meters³/second]) corresponds to about 22 inches (about 560 mm) of runoff distributed over the 40,900 square mile (105,930 square kilometer drainage basin. A larger amount of runoff occurs during the wet winter and spring months (January-April) when precipitation events are frequent, temperatures are low, and there are no leaves or deciduous vegetation. Consequently, soil absorption, evaporation, and plant transpiration losses are low at that time of year, and both runoff and streamflow are higher than during the summer and autumn months. Average rainfall in the eastern and western portions of the Tennessee Valley (above and below Chattanooga) is about equal. However, topographic differences (viz. the largely steep and mountainous terrain in the eastern portion of the Valley, compared with the mostly flat and rolling terrain in the western portion of the Valley) and generally shallow soils result in higher amounts of runoff above Chattanooga.

In 1993, runoff for the Tennessee River basin was well below normal, particularly from February through July and particularly in the western half of the Valley. Runoff above Chattanooga was only slightly below normal in 1993, 21.4 inches, or 92 percent of the long-term mean of 23.4 inches. However, runoff above Kentucky Dam was only 17.6 inches, a deficit of almost 5 inches and only 78 percent of the long-term mean of 22.5 inches (figure 2c). The 1993 releases from tributary reservoirs in the western part of the Valley (e.g., Normandy, Tims Ford, etc.) were below their long-

term means, while the releases from the tributary reservoirs in the eastern part of the Valley (e.g., South Holston, Watauga, etc.) were close to normal. Consequently, flows in the Tennessee River in 1993 increasingly fell below the long-term average as the river flowed downstream from Fort Loudoun Dam to Kentucky Dam.

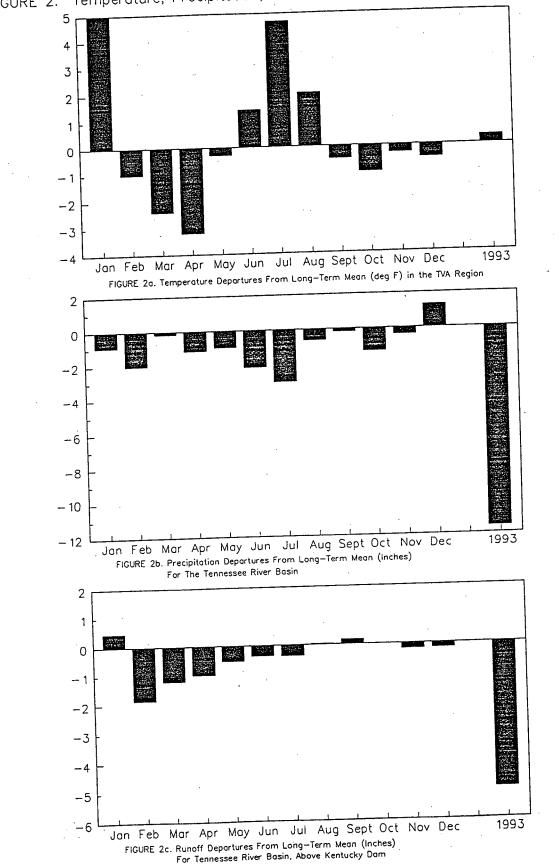


FIGURE 2. Temperature, Precipitation, and Runoff - Tennessee River Basin, 1993

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aquatic life in the tailrace of Chickamauga dam (i.e. inflow sampling site on Nickajack Reservoir), the inflow sampling site's DO rating used in the overall ecological health evaluation of Nickajack Reservoir was "poor." Based on no DO's actually being measured in the hypolimnion of the forebay of Nickajack Reservoir below 2 mg/l, the forebay sampling site's DO rating was "excellent".

Values of pH varied over a rather narrow range, from 7.0-8.0 during the summer of 1993. At the forebay, the highest chlorophyll-a concentration of about 10 ug/l was measured in May and averaged about 6 ug/l in the summer of 1993. Consequently, the chlorophyll-a rating used in the 1993 ecological health evaluation for Nickajack Reservoir was "good" (i.e. average concentration between 3 and 10 ug/l).

Chickamauga Reservoir

Water--During the summer of 1993 (April-September), coolest surface water temperatures in Chickamauga Reservoir were in April and the warmest in July. Surface temperatures ranged from a minimum of 17.0°C to a maximum of 31.7°C at the forebay; from 16.2°C to 30.1°C at the transition zone; and from 19.1°C to 28.8°C in the Hiwassee River embayment. The State of Tennessee's maximum water temperature criteria for the protection of fish and aquatic life is 30.5°C.

Dissolved oxygen (DO) concentrations at the 1.5-meter depth ranged from a low of 6.9 mg/l in September to a high of 11.4 mg/l in April at the forebay; from 5.7 mg/l in September to 10.3 mg/l in April at the transition zone; and from 7.3 mg/l in August to

9.9 mg/l in April at the sampling location in the Hiwassee River embayment. At the inflow sampling site (i.e. the tailrace of Watts Bar dam) a minimum DQ of 3.7 mg/l was recorded in August. The State of Tennessee's minimum dissolved oxygen criteria for the protection of fish and aquatic life is 5.0 mg/l, measured at the 1.5 meter depth.

Temperature data depict seasonal warming and weak thermal stratification in Chickamauga Reservoir from May through July. The maximum observed surface to bottom temperature differentials occurred in July. Temperature differentials were 5.5°C at the forebay, 3.2°C at the transition zone, and 4.1°C in the Hiwassee River embayment. There was also an oxycline at the forebay and transition zone in June and July when differences between surface and bottom DO's were about 6 to 9 mg/l at the forebay and transition zone. In July 1993, a minimum DO of less than 0.1 mg/l was measured on the bottom at the forebay and a minimum of 1.6 mg/l was measured on the bottom at the transition zone. Better DO conditions were observed in the Hiwassee River embayment portion of Chickamauga Reservoir, where maximum DO differentials were only 1.7 mg/l and near-bottom DO's only slightly below 6 mg/l.

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DO ratings used in the overall reservoir ecological health evaluation for Chickamauga Reservoir were "good" at the forebay; "good to excellent" at the transition zone; "excellent" in Hiwassee River embayment; and "fair" at the inflow. The forebay would have rated higher had it not been for the low near-bottom oxygen concentrations which existed in July. The fair rating at

the inflow sampling site on Chickamauga Reservoir was a result of oxygen levels being measured about 1.5 mg/l below the Tennessee criteria (5 mg/l, at the 1.5 meter depth) in the releases from Watts Bar dam.

Values of pH ranged from 6.8 to 8.8 on Chickamauga Reservoir in 1993. Near-surface pH values exceeding 8.5 (and DO saturation values exceeding 100 percent) were observed on only two occasions (April and July), both at the forebay. Both of these periods of high pH and high oxygen saturations were also coincident with high chlorophyll-a concentrations, indicative of periods of high photosynthetic activity. The State of Tennessee's maximum pH criteria for the protection of fish and aquatic life is 8.5.

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Total nitrogen (TN), total phosphorus (TP), and dissolved ortho phosphorus (DOP) were low in the Tennessee River portion of Chickamauga Reservoir in 1993. TN averaged only 0.37 mg/l at the forebay, the lowest TN concentration measured at any of the Tennessee River sampling sites in 1993. At both the forebay and the transition zone, TP and DOP concentrations averaged only about 0.026 mg/l and 0.005 mg/l, respectively, and were among the lowest TP and DOP concentrations measured at any of the Tennessee River sampling sites in 1993. Because of these low concentrations (and because TN/TP ratios often exceeded 20), periods of phosphorus limitation on algal productivity were likely to have occurred.

In 1993, Chickamauga Reservoir chlorophyll-a concentrations averaged 8.5 ug/l, 7.8 ug/l, and 5.5 ug/l, respectively, at the forebay, transition zone, and Hiwassee River embayment. Consequently, the chlorophyll-a ratings used in the 1993

ecological health evaluation for Chickamauga Reservoir were "good" (i.e. falling in the 3 to 10 ug/l range) at all three locations.

HIWASSEE RIVER WATERSHED

<u>Hiwassee Reservoir</u>

<u>Water</u>--The average flow through Hiwassee Reservoir was about 107 percent of normal; the average residence time was about 99 days. The reservoir is strongly stratified, with a maximum temperature difference in the water column at the forebay of 20.9°C in July. The maximum surface temperature was 28.7°C in July, both at the forebay and mid-reservoir. North Carolina's standard for maximum temperature of Class C waters is 29°C. Low D0 water (D0 <5.0 mg/l) first appeared at mid-reservoir in June and at the forebay in July, at the bottom of the water column at both locations. Depleted D0 water (D0 <2.0 mg/l) occurred at both locations at the bottom of the water column in August and September. The limited area of D0 depletion provided ratings for the reservoir ecological health index of fair at the forebay and good at mid-reservoir.

Conductivities averaged about 30 umhos/cm in April, and increased slightly in the DO-depleted area to a maximum of 40 and 38 umhos/cm at the forebay and mid-reservoir, respectively. The average conductivity in Hiwassee Reservoir was the fourth-lowest of the 19 tributary reservoirs. Only in June, July, and August did pH reach or exceed 8.4 SU, and only in the four- to eightmeter depth. Summer DO concentrations were normally higher at these depths. the 33 tributary reservoir stations. Organic and nitrate nitrogen concentrations were 0.03 and 0.09 mg/l in April, and 0.06 and 0.04 mg/l in August. Total and dissolved ortho phosphorus concentrations were 0.005 and 0.003 mg/l in April, and 0.002 and <0.002 in August. Total organic carbon concentrations were very low--0.8 and 1.4 mg/l in April and August, respectively. Chlorophyll-a concentrations averaged 2.5 ug/l. This chlorophyll concentration is considered fair in the reservoir ecological health index. Secchi depths varied from 1.6 meters in April to 3.6 meters in July, September, and October.

WATTS BAR RESERVOIR, FORT LOUDOUN RESERVOIR, AND MELTON HILL RESERVOIR WATERSHED

<u>Watts Bar Reservoir</u>

<u>Water</u>--During the summer of 1993 (April-September), surface water temperatures ranged from a minimum of 18.3°C in April to a maximum of 30.2°C in July in the forebay; and from 16.7°C to 29.8°C (for the same months) at the transition zone. The State of Tennessee's maximum water temperature criteria for the protection of fish and aquatic life is 30.5°C.

Values for DO at the 1.5-meter depth ranged from a low of 6.5 mg/l in September to a high of 12.6 mg/l in April at the forebay; and from 7.1 mg/l to 11.3 mg/l (for the same months) at the transition zone. At the inflow sampling site on the Tennessee River arm of Watts Bar Reservoir (i.e. the tailrace of Fort Loudoun dam) a minimum DO of 3.9 mg/l was recorded in September.

At the inflow sampling site on the Clinch River arm of Watts Bar Reservoir (i.e. the tailrace of Melton Hill dam) a minimum DO of 6.3 mg/l was recorded in March. The State of Tennessee's minimum dissolved oxygen criteria for the protection of fish and aquatic life is 5.0 mg/l, measured at the 1.5 meter depth.

Temperature and dissolved oxygen data show that Watts Bar Reservoir developed a moderate degree of both thermal and oxygen stratification throughout most of the summer of 1993. For the period April through August, monthly surface to bottom temperatur differentials were: 5.2°C, 5.5°C, 7.4°C, 7.3°C, and 4.0°C at the forebay; and 2.3°C, 2.6°C, 3.9°C, 6.2°C, and 2.2°C at the transition zone.

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DO versus depth data show that a rather strong oxycline also developed in Watts Bar Reservoir, particularly from June through August. During these three months, surface to bottom differences in DO were: 9.2 mg/l, 9.2 mg/l, and 5.8 mg/l at the forebay; and 7.2 mg/l, 5.8 mg/l, and 3.1 mg/l at the transition zone. At the forebay, near-bottom DO concentrations in the hypolimnion were less than 2 mg/l in June and July. In addition, the proportion of the hypolimnion with low DO's (i.e., less than 2 mg/l) averaged about 13% of the total cross sectional area, higher than in any other Tennessee River reservoir. The minimum observed DO concentration in Watts Bar Reservoir in 1993 was 0.6 mg/l at the bottom of the forebay in July; but DO's were never less than 4 mg/l at the transition zone.

DO ratings used in the overall reservoir ecological health evaluation for Watts Bar Reservoir were "poor" at the forebay; "excellent" at the transition zone and at the inflow sampling site on the Clinch River; and "fair" at the inflow site on the Tennessee River. The low forebay rating was due to the large proportion of the forebay hypolimnion with low DO's (i.e. less than 2 mg/l). The fair rating at the inflow sampling site on the Tennessee River arm of Watts Bar Reservoir was a result of oxygen levels being measured about 1 mg/l below the Tennessee criteria (5 mg/l, at the 1.5 meter depth) in the releases from Fort Loudoun dam.

Historically, pH's measured in Watts Bar Reservoir have been higher than other Tennessee River sampling sites. This is due to the addition of the cool, clear, well-oxygenated, nitrate rich, and hard water of the Clinch River which combines with the Tennessee River (and Watts Bar Reservoir) at Tennessee River Mile (TRM) 567.9, about 7 miles upstream from the transition zone sampling site. In the summer of 1993, values of pH ranged from 6.8 to 9.0 on Watts Bar Reservoir. Throughout the summer (April-September), near-surface values of pH frequently exceeded 8.5 at both the forebay and the transition zone, with DO saturation values commonly exceeding 100 percent, indicating high rates of photosynthesis. The State of Tennessee's maximum pH criteria for the protection of fish and aquatic life is 8.5.

The average total phosphorus concentrations observed in Watts Bar Reservoir (0.029 mg/l at the forebay and 0.035 mg/l at the transition zone) were among the lowest of the Tennessee River Vital Signs Monitoring locations in 1993. In addition, the average dissolved ortho phosphorus concentrations of 0.007 mg/l

and 0.004 mg/l, respectively, at the forebay and transition zones were also among the lowest observed at any of the Tennessee River Vital Signs Monitoring locations in 1993. TN/TP ratios on Watts Bar Reservoir are higher than on any other Tennessee River reservoir. The low phosphorus concentrations in combination with the relatively high nitrogen concentrations (supplied by both the Clinch and Tennessee River inflows) results in the high TN/TP ratios in Watts Bar (particularly at the transition zone) and suggest periods of phosphorus limitation on primary productivity.

The highest chlorophyll-a concentrations were measured in August at the forebay (10 ug/l) and in May at the transition zone (11 ug/l). Surface concentrations of chlorophyll-a averaged about 7 ug/l at the forebay and about 8 ug/l at the transition zone in 1993. Consequently, the chlorophyll-a ratings used in the 1993 ecological health evaluation for Watts Bar Reservoir were "good" (i.e., falling in the 3 to 10 ug/l range) at both locations.

Forebay Secchi depth and suspended solids measurements average 1.5 meters and 6.3 mg/l, respectively. These values indicate the light transparency of Watts Bar Reservoir forebay to be relatively high compared with other main stem Tennessee River reservoirs in 1993.

Fort Loudoun Reservoir

<u>Water</u>--Temperature and dissolved oxygen (DO) data show the establishment of stratification (both a thermocline and oxycline) in Fort Loudoun reservoir which persisted throughout most of the summer (April through September) of 1993. Summer surface water temperatures were warmest in July and coolest in April. They

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	N	Mean	Min	Max	N	Mean	Min	Мах	N	Mean		
Temperature (deg C) Dissolved Oxygen (mg/l) Percent Saturation (%) pH (s.u.) Conductivity (umhos/cm) Organic N (mg/l) Ammonia - N (mg/l) Nitrate+nitrite - N (mg/l) Total Nitrogen (mg/l) Total Phosphorus (mg/l) Dissolved Ortho - P (mg/l) TN/TP Ratio Total Organic Carbon (mg/l Chlorophyll-a (ug/l)	88 88 88 88 16 16 16 16 16 16	24.539 6.502 76.069 7.598 177.602 0.144 0.043 0.183 0.370 0.026 0.005 15.097 1.908 8.500 1.167	$15.100 \\ 0.020 \\ 0.200 \\ 6.800 \\ 162.000 \\ 0.020 \\ 0.010 \\ 0.010 \\ 0.230 \\ 0.020 \\ 0.002 \\ 8.000 \\ 1.700 \\ 7.000 \\ 1.000 \\ 1.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0.000 \\ 0$	$\begin{array}{c} 31.700 \\ 11.500 \\ 129.300 \\ 8.800 \\ 192.000 \\ 0.310 \\ 0.110 \\ 0.320 \\ 0.540 \\ 0.040 \\ 0.010 \\ 22.000 \\ 2.300 \\ 10.000 \\ 1.300 \end{array}$	60 60 60 12 12 12 12 12 12 12 12 12 12 12 5	$\begin{array}{c} 23.973 \\ 6.678 \\ 77.172 \\ 7.575 \\ 177.567 \\ 0.197 \\ 0.046 \\ 0.219 \\ 0.462 \\ 0.026 \\ 0.006 \\ 20.393 \\ 1.933 \\ 7.833 \\ 1.196 \end{array}$	15.300 1.200 14.800 6.900 161.000 0.010 0.010 0.010 0.010 0.002 7.750 1.800 3.000 1.000 2.000	30.100 10.700 126.800 8.600 190.000 0.900 0.340 1.140 0.050 0.010 50.000 2.200 13.000 1.300 21.000	53 53 53 53 12 12 12 12 12 12 12 12 12 6 6 12	21.928 7.868 88.640 7.200 139.585 0.172 0.079 0.157 0.409 0.043 0.007 9.829 2.342 5.500 0.933 11.833	$16.700 \\ 5.800 \\ 69.000 \\ 6.800 \\ 100.000 \\ 0.040 \\ 0.010 \\ 0.110 \\ 0.260 \\ 0.030 \\ 0.003 \\ 6.500 \\ 1.800 \\ 3.000 \\ 0.770 \\ 7.000 \\ \end{array}$	12.000 1.250 20.000
Secchi Depth (m) Suspended Solids (mg/l) True Color (PCU) Fecal Coliform (#/100ml)	16 16 8	6.167 6.333 10.000	3.000 1.000 10.000	17.000 10.000 10.000	12 12 6	7.000 7.917 10.000	5.000 10.000	10.000	12 5	14.583 68.000	10.000 10.000	

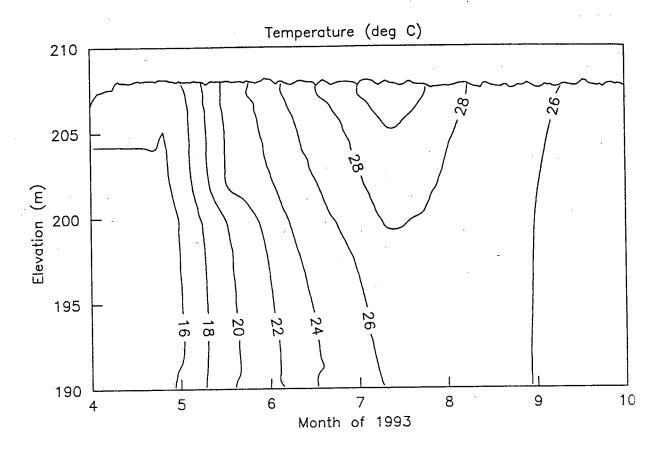
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	Watts	Bar Fore	bay (TRM	531.0)	Watts	Bar Tran	sition (T	RM 560.8)
	N	Mean	Min	Max	N	Mean	Min	Max
Temperature (deg C) Dissolved Oxygen (mg/l) Percent Saturation (%) pH (s.u.) Conductivity (umhos/cm) Organic N (mg/l) Ammonia - N (mg/l) Nitrate+nitrite - N (mg/l) Total Nitrogen (mg/l) Total Phosphorus (mg/l) Dissolved Ortho - P (mg/l) TN/TP Ratio Total Organic Carbon (mg/l) Chlorophyll-a (ug/l) Secchi Depth (m) Suspended Solids (mg/l) True Color (PCU) Fecal Coliform (#/100ml)	16 17 .) 17 16 (1) 17 8 6 17 17	22.981 6.458 73.773 7.787 184.975 0.187 0.024 0.244 0.461 0.029 0.007 15.990 2.008 7.167 1.545 6.308 7.308 10.000	5.000		69 69 69 12 12 12 12 12 12 12 12 12 12 12 12 6 6 12 12 6	$\begin{array}{c} 23.067\\ 7.933\\ 91.233\\ 7.832\\ 191.768\\ 0.218\\ 0.033\\ 0.277\\ 0.527\\ 0.035\\ 0.004\\ 16.531\\ 1.892\\ 7.833\\ 1.233\\ 8.667\\ 7.167\\ 10.000 \end{array}$	$14.400 \\ 4.000 \\ 4.000 \\ 6.900 \\ 162.000 \\ 0.040 \\ 0.010 \\ 0.300 \\ 0.020 \\ 0.002 \\ 7.500 \\ 1.700 \\ 3.000 \\ 1.000 \\ 4.000 \\ 1.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.000 \\ 10.00$	29.800 11.500 139.500 8.900 218.000 0.470 0.060 0.420 0.750 0.050 0.008 37.500 2.200 11.000 1.520 15.000 10.000

Watts Bar Transition (TRM 560.8)

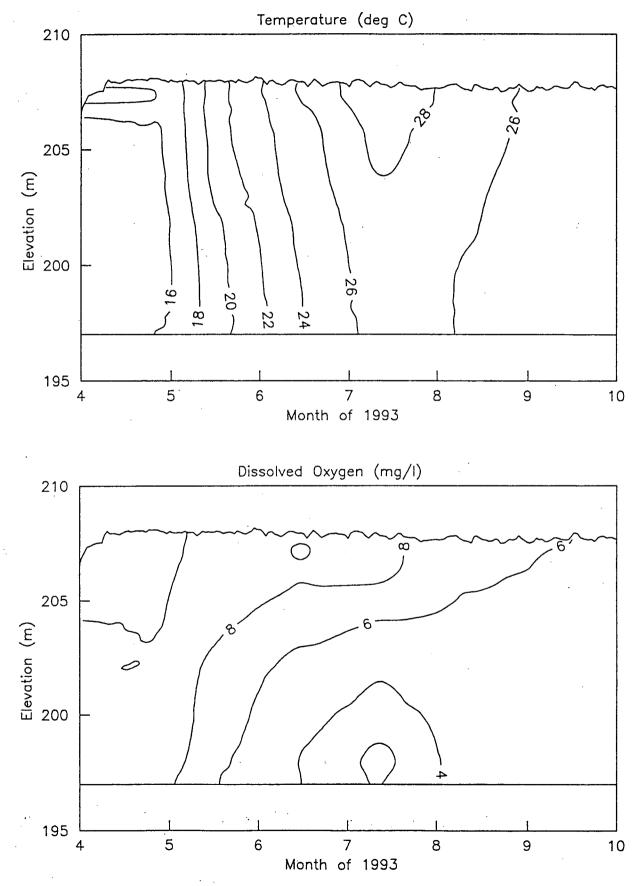
The applaint operation of

Chickamauga Reservoir – TRM 472.3

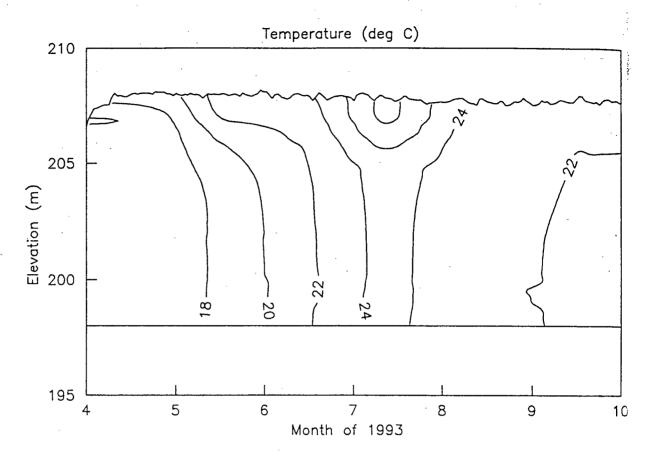


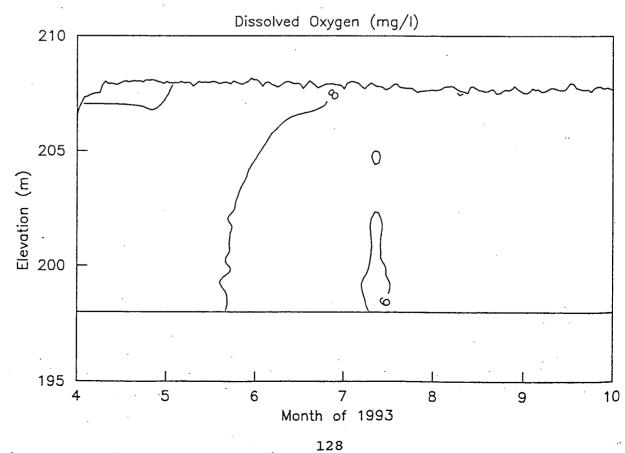
Dissolved Oxygen (mg/l) 210 <u>0</u> ଚ 205 Elevation (m) 007 195 190 8 9 6 7 10 5 Δ Month of 1993

Chickamauga Reservoir – TRM 490.5

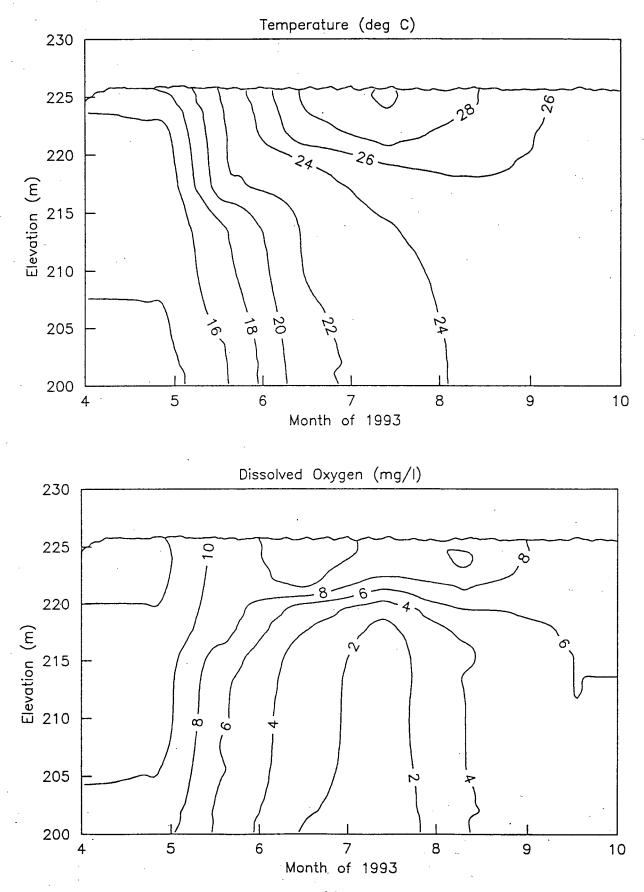


Chickamauga Reservoir – Hiwassee River Mile 8.5





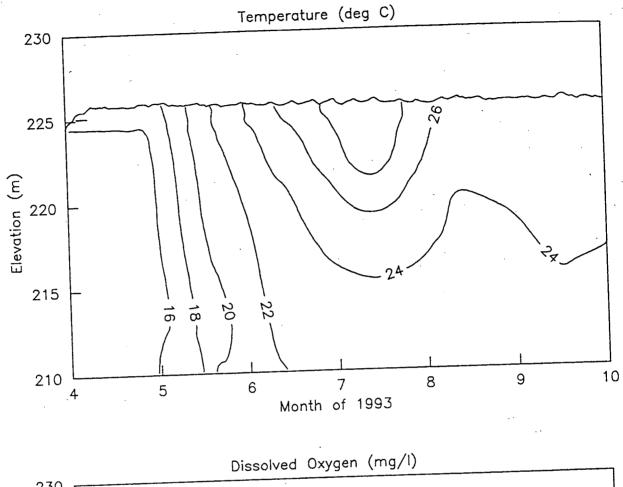
Watts Bar Reservoir - TRM 531.0

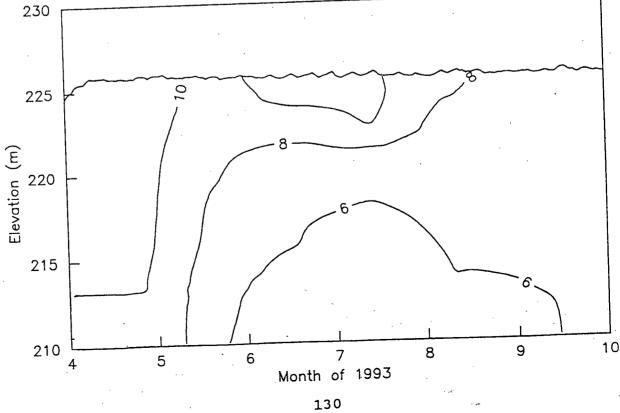


Watts Bar Reservoir - TRM 560.8

Figure 20

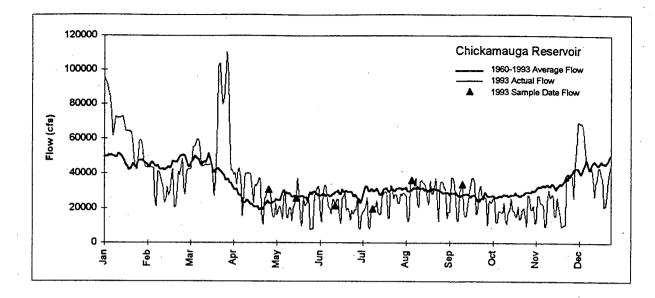
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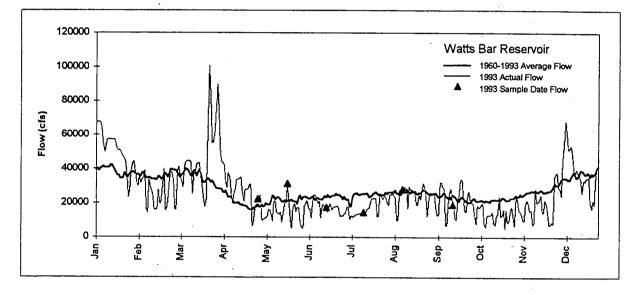


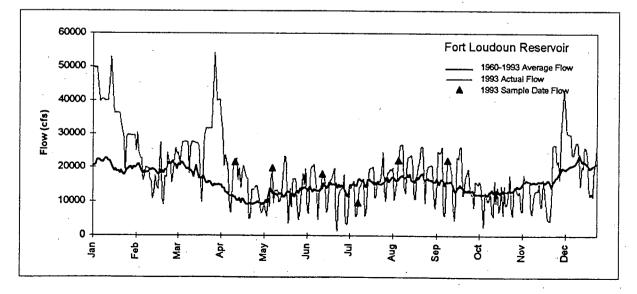


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STORET RETRIEVAL DATE 94/05/26 475358 35 06 26.0 085 12 20.0 2 CMICKAMAUGA RES. AT LIGHTED BUOY 47065 TENNESSEE ATVER BASIN TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 472.3 1311VAC 0000 METERS DEPTH 06020001021 0000.710 ON

/TYPA/AMBNT/STREAM/SOLIDS

INDEX 1021500 007720 00920 Miles 0953.80 0046.50 472.30

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يدحدن لالمتحسبة

		472.30						
FROM TO	TINE OF DAY HEDIUN	SMK Or Depth (M)	00060 STREAN FLOW CFS	OOD10 WATER TEMP CENT	00300 D0 Hg/L	00301 Do Satur Percent	00400 РН SU	00094 CHDUCTVY FIELD
93/04/29 93/04/29 93/04/29 93/04/29 93/04/29 93/04/29 93/04/29 93/04/29 93/04/29 93/04/29 93/04/29 93/04/29 93/04/29	1050 WATER 1052 WATER 1054 WATER 1056 WATER 1058 WATER 102 WATER 104 WATER 108 WATER 1108 WATER 115 WATER 115 WATER 115 WATER 350 WATER	0.3 1 1.5 2 4 6 8 10 12 14 16 14.6 0.3	25463	17.0 16.4 16.2 15.9 15.4 15.4 15.3 15.2 15.2 22.9	11.4 11.5 11.4 11.2 11.0 10.8 10.8 10.0 9.7 9.7 9.7 9.7 9.2 9.0	117.55 115.03 114.03 112.05 110.05 108.03 98.05 97.15 95.15 85.35 90.25 103.45	8.50 8.50 8.40 8.30 7.90 7.90 7.80 7.70 7.60 7.70	HICRONHO 171 170 170 169 173 175 175 169 173 169
93/05/19 1 93/05/19 1 93/06/16 1 93/06/16 1 93/06/16 1	352 WATER 355 WATER 355 WATER 357 WATER 357 WATER 358 WATER 359 WATER 400 WATER 400 WATER 400 WATER 400 WATER 10 WATER 10 WATER 12 WATER	1 1.5 6 7.4 10 12 14 14.5 17 0.1 1.5 3	21188	22.9 22.8 22.5 20.8 20.3 19.9 19.7 19.7 19.7 19.7 27.8 27.6 27.6 27.3 26.5	9.0 9.5 7.1 6.3 6.3 9.1 9.1 9.1 9.1 9.1 8.3	103.4\$ 96.6\$ 87.5\$ 78.9\$ 72.8\$ 68.5\$ 68.5\$ 66.3\$ 64.1\$ 65.0\$ 65.0\$ 61.5\$ 115.2\$ 115.2\$ 112.3\$	7.60 7.70 7.50 7.20 7.00 6.90 6.80 6.80 6.80 6.80 6.80 8.40 8.40 8.40 8.40	177 178 178 180 180 181 182 183 183 183 183 183 183 183 183 183 183
93/06/16 14 93/06/16 14 93/06/16 14 93/06/16 14 93/06/16 14 93/06/16 14 93/06/16 14 93/06/16 14 93/06/16 14	16 WATER 22 WATER 24 WATER 26 WATER 28 WATER 30 WATER 32 WATER 34 WATER 36 WATER 36 WATER	3.5 4.5 5 8 9 10 12		26.5 26.4 26.1 25.8 25.3 25.0 24.6 24.2 23.9	8.0 7.8 6.8 6.4 5.6 5.0 4.4 3.4 2.8	101.25 97.65 95.15 82.95 81.75 78.05 66.75 59.55 51.25 40.05	8.10 8.00 7.60 7.50 7.30 7.20 7.10 7.00	167 166 166 168 171 166 162 169 171
93/06/16 14 93/07/13 14 93/07/13 14 93/07/13 14 93/07/13 14 93/07/13 140 93/07/13 140 93/07/13 140 93/07/13 141 93/07/13 141 93/07/13 141 92/07/13 141	22 WATER 22 WATER 23 WATER 23 WATER 24 WATER 25 WATER 26 WATER 29 WATER 21 WATER 21 WATER 21 WATER 21 WATER 21 WATER 21 WATER	14.5 16 0.3 1.5 2.5 3.5 5.5 5.5 6 7.5 8	19467	23.8 23.7 31.3 30.6 30.2 29.9 29.8 29.8 29.6 29.2 29.2 29.2 29.2 29.2 29.2 29.2	2.03 99.4 99.1 8.125 8.125 5.00	32.9\$ 31.8\$ 23.5\$ 125.7\$ 125.3\$ 126.3\$ 126.3\$ 126.3\$ 126.3\$ 106.6\$ 94.7\$ 85.5\$ 71.8\$ 64.1\$ 5.0.6\$	7.00 7.00 8.80 8.80 8.80 8.80 8.80 8.80	169 165 172 172 171 172 172 172 173 174 173 178 175 177
93/07/13 141 93/07/13 141 93/07/13 141 93/07/13 141 93/07/13 142 93/07/13 142 93/07/13 142 93/07/13 142 93/07/13 142 93/08/10 1346 93/08/10 1357 93/08/10 1357 93/08/10 1357 93/08/10 1357 93/08/10 1357	6 WATER 7 WATER 9 WATER 9 WATER 1 WATER 2 WATER 3 WATER 9 WATER WATER WATER WATER WATER WATER WATER WATER	8.5 910 12 14.7 16.7 17.5 0.3 1.5 2 6 8 10	35983	28.3 27.6 27.6 27.0 26.7 26.6 26.4 26.2 27.8 26.4 26.2 27.7 27.7 27.7 27.7 27.3 27.3 27.1	3.8 3.6 2.1 1.2 .02 7.3 7.2 6.1 5.6	48.15 45.65 32.93 25.93 21.05 14.85 .25 92.45 92.45 91.13 87.35 71.65 69.15	7.20 7.10 6.90 6.80 6.80 6.80 6.80 6.80 7.90 7.90 7.90 7.90 7.90 7.60 7.50	180 182 187 181 182 180 181 182 180 183 184 187 187 187 187 187 189 189 189
93/08/10 1405 93/08/10 1407	WATER WATER WATER 1 WATER	12 14 16 6.5 17 0.3 1	33892	27.1 27.1 27.0 27.0 27.0 27.0 27.0 27.0 25.7 25.6	5.5 5.1 4.5 4.5 4.4 7.2 7.1	67.9\$ 63.0\$ 58.0\$ 55.6\$ 55.6\$ 54.3\$ 87.8\$ 86.6\$	7.50 7.40 7.30 7.30 7.30 7.70 7.70	188 188 188 188 187 188 187 188 182 183

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93/09/15 93/09/15 93/09/15 93/09/15 93/09/15 93/09/15 93/09/15	1310 WATER 1312 WATER 1313 WATER 1314 WATER	SHK OR DEPTH (H) 1.5 4 6 8 10 12 14 16 17	00060 Stream Flow CFS	00010 WATER TEMP CENT 25.6 25.5 25.4 25.2 25.2 25.2 25.2 25.2 25.2	00300 D0 NG/L 6.9 6.7 6.3 5.8 5.8 5.8 5.8 5.8 5.8 5.8 5.8 5.8	00301 D0 SATUR PERCENT 86.15 79.85 69.05 69.05 69.05 69.05 69.05 69.05	00400 PH SU 7.60 7.60 7.40 7.40 7.40 7.40 7.40 7.40	00094 CNOUCTYY FIELD MICRONHO 184 185 185 185 185 185 183 181 192 189
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STORET RETRIEVAL DATE 94 475358 1017 35 06 26.0 085 12 20.0 2 Chickamauga Res. At Ligh	TED BUOY	
47065 TENNESSEE Tennessee River Basin	NAMILTON 040801	/TYPA/ANBNT/STREAM/SOLIDS
TENNESSEE RIVER 472.3 131TVAC 0000 Neters Depth	06020001021 0000.710 OM	

INDEX 1021500 007720 00920 Miles 0953.80 0046.50 472.30 00530 RESIDUE TOT NFLT MG/L 00605 00610 NH3+NH4-N TOTAL 31616 FEC COLI HFM-FCBR /100HL 00080 84002 CODE GENERAL 00078 Transp SHK ORG H N NG/L COLOR PT-CO UNITS OR DEPTH (H) DATE FROM TO TIME OF DAY <u>M</u>edium SECCHI HG/L REMARKS TO DAY MEDIU 93/04/29 1050 WATER 93/04/29 1100 VERT 93/04/29 1120 WATER 93/05/19 1350 WATER 93/05/19 1354 VERT 93/05/19 1401 WATER 93/06/16 1402 WATER 93/06/16 1402 WATER 93/06/16 1418 VERT 93/06/16 1419 VERT 93/06/16 1420 VERT 93/06/16 1420 VERT 93/06/16 1439 WATER 93/06/16 1439 WATER 93/06/16 1439 VATER 93/06/16 1439 VATER 93/06/16 1439 VATER 93/06/16 1430 WATER 93/06/10 1356 VERT 93/08/10 1356 VERT 93/08/10 1356 VERT 93/09/15 1311 VERT 93/09/15 1316 WATER 10K 0.3 1.30 .130 5 q 5 q 6 4 14.6 1.00 10K .090 .070 5 10 36 14.5 0.3 0.3 0.3 4 10K D1 D2 D3 D1 D2 D1 D2 D1 D2 D3 D1 D2 D3 10K .120 .160 .170 .130 .130 .090 4 ş 4 5 47 555 14.5 14.5 0.3 14.7 0.3 14.7 0.3 18.5 0.3 18.5 14 8

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DATE FROM TO	T I NE OF DAY	MEDIUM	SMK OR DEPTH (M)	84002 CODE GENERAL REMARKS	00665 Phos-tot Ng/L P	00671 PHOS-DIS Ortho Mg/L P	T	00680 Org C C Hg/L	32211 CHLRPHYL A UG/L Correcto	32212 Chlrphyl B Ug/L	32214 CHLRPHYL C UG/L	32218 Pheophtn A Ug/L
			4		.020	.002K		1.9	9.00	1.00	2.00	2.00
93/04/29 93/04/29 93/05/19 93/05/19	1120 1354 1401	WATER VERT WATER	14.6		.020 .030 .030	.007 .002 .006		1.7 2.0 1.7	9.00	1.00K	1.00	1.00
93/06/16 93/06/16 93/06/16	1403	WATER	0.3 0.3 0.3	01 02 03					7.00	1.00K	1.00	2.00
93/06/16 93/06/16 93/06/16	1418 1419 1420	VERT VERT VERT	44	D 1 D 2 D 3 D 1	.030 .030 .030 .030	.002 .002K .002 .020		2.3 2.3 2.3 1.9	5.00	1.00K	1.00	1.00 2.00
93/06/16 93/06/16 93/06/16 93/06/16 93/07/13	1439	WATER WATER	14.5 14.5 14.5 4	D 2 0 3	.030 .030 .020	.010 .010 .003		1.8 1.8 2.2 1.8	10.00	1.00	1.00	2.00
93/07/13	1420	WATER	14.7		.020	.005 .0021		2.0	9.00	1.00K	1.00	2.00
93/08/10 93/08/10 93/09/15 93/09/15	1409	VERT WATER VERT WATER	18.5 14		.040 .020 .030	.006 .007 .009		1.9 1.8 1.8	7.00	1.006	1.00	1.00

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STORET RETRIEVAL DATE 94/05/26 475265 1053 35 18 00.0 085 04 33.0 2

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CHICKAMAUGA RESERVOIR 47065 TENNESSEE TENNESSEE RIVER BASIN TENNESSEE RIVER 490.47

TINE

93/04/29 0915 WATER

93/04/29 0920 WATER 93/04/29 0925 WATER

93/04/29 0930 WATER 93/04/29 0935 WATER

93/04/29 0945 WATER 93/04/29 0950 WATER

93/04/29 0955 WATER 93/05/19 1038 WATER

93/05/19 1039 WATER

93/06/16 1308 WATER 93/06/16 1311 WATER

93/06/16 1314 WATER 93/06/16 1314 WATER 93/06/16 1317 WATER 93/06/16 1323 WATER 93/06/16 1323 WATER 93/07/13 1337 WATER

93/07/13 1333 WATER 93/07/13 1338 WATER 93/07/13 1338 WATER 93/07/13 1340 WATER 93/07/13 1341 WATER 93/07/13 1341 WATER 93/07/13 1343 WATER 93/07/13 1345 WATER

93/07/13 1346 WATER 93/07/13 1347 WATER

93/07/13 1348 WATER 93/07/13 1349 WATER

93/07/13 1350 WATER 93/08/10 1234 WATER

93/08/10 1234 WATER 93/08/10 1236 WATER 93/08/10 1238 WATER 93/08/10 1240 WATER

93/08/10 1242 WATER

93/08/10 1244 WATER 93/08/10 1246 WATER

93/08/10 1248 WATER

93/08/10 1250 WATER

93/09/15 1205 WATER 93/09/15 1205 WATER 93/09/15 1206 WATER 93/09/15 1207 WATER

93/09/15 1208 WATER 93/09/15 1209 WATER 93/09/15 1211 WATER 93/09/15 1212 WATER

93/09/15 1213 WATER 93/09/15 1214 WATER

D.F.

131TVAC 0000 METERS DEPTH

DATE

FROM

t:

STORET RETRIEVAL DATE 94 475265 1053 35 18 00.0 085 04 33.0 2 Chickanauga Reservoir 47065 Tennessee Tennessee River Basin Tennessee River 490.47	HAMILTON 040801	/TYPA/AWBNT/STREAM/SOLIDS
131TVAC 0000 Meters Depth	06020001025 0005.740 ON	

INDEX 1021500 007720 Miles 0953.80 0046.50	00920 490.47				,		00605	00610	00630
DATE TIME FROM OF TO DAY MEDIUM	SHK OR DEPTR (H)	84002 CODE GENERAL REHARKS	00078 TRANSP SECCHI METERS	31616 FEC COLI MFN-FCBR /100HL	00080 COLOR PT-CO UNITS	00530 RESIDUE TOT NFLT KG/L	ORG N NG/L	NH3+NH4- H TOTAL ' MG/L	NOZANOS N-TOTAL MG/L
93/04/29 0915 WATER 93/04/29 0940 VERT	0.3 4 9.6000	•	1.25	10K	59 59	· 5 6	.030	.020 .040	. 33 . 34
93/04/29 0955 WATER 93/05/19 1038 WATER 93/05/19 1042 VERT	9.8000 0.3 4 10		1.00	10K	5	6 5	.130 .080	.010K .070	. 17 . 26
93/05/19 1046 WATER 93/06/16 1247 WATER 93/06/16 1302 VERT 93/06/16 1320 WATER	0.3 9.5			10K	10 5	6 9	.140	.010 .090	. 16
93/07/13 1337 WATER 93/07/13 1344 VERT 93/07/13 1349 WATER	0.3 4 9.6000		1.18	10K	10 10	4	.210 .170	.030	.11
93/08/10 1234 WATER 93/08/10 1243 VERT 93/08/10 1248 WATER	0.3 4 9.6000		1.30	10K	10 10	2 7	.280	.010K .040	.20
93/09/15 1205 WATER 93/09/15 1210 VERT 93/09/15 1213 WATER	0.3 4 9		1,25	. 104	10 10	6 21	.130 .900	.070	.18 .17

DATE FROM TO	TIME OF DAY	HEDIUN	SHK OR DEPTH (H)	84002 CODE GENERAL REMARKS	00665 PHDS-TOT NG/L P	00671 PHOS-DIS DRTHD MG/L P	т	00680 Org C C Mg/L	32211 CHLRPHYL A UG/L CORRECTO	32212 CHLRPHYL B UG/L	32214 CHLRPHYL C UG/L	32218 PHEOPHTN A UG/L
93/04/29	0940	VERT	4	•	.020	.002		1.8	7.00	1.00	1.00	2.00
93/04/29 93/05/19	0955	WATER	9.6000		.020	.004		1.8	13.00	1.00K	2.00	2.00
93/05/19	1046	WATER	10		.020	.007		1.8	10.00	1.00	1.00	3.00
93/06/16 93/06/16	1320	WATER	9.5		.040	.003		1.9	9.00	1.00	1.00	3.00
93/07/13 93/07/13	1349	WATER	9.6000		.020	.010		1.9	5.00	1.00K	1.00	1.00
93/08/10 93/08/10 93/09/15 93/09/15	1248	VERT WATER VERT WATER	9.6000		.020 .030 .050	.008 .010 .010		1.9 1.9 1.8	3.00	1.00K	1.00	1.00

ARTRIEVAL DATE 94/05/26 ARTRIEVAL DATE 94/05/26 ARTRIEVAL DATE 94/05/26 AUGA RESERVOIR - 2 HILES ABOVE HIGHWAY 58 ATTE TEMPESSEE RIVER BASIN HIWASSEE RIVER BASIN OG02002 0000 HETERS DEPTH

INDEX 1021500 007720 00920 6824 Miles 0953.80 0046.50 499.70 008.50

	TINE OF DAY MEDIUN	••	00060 Stream Flow CFS	COCIO WATER TEMP CENT	00300 Do Hg/L	00301 DO SATUR PERCENT	00400 PH SU	00094 CNDUCTVY Field Hicronho
FROM TO 93/04/28 93/04/28 93/04/28 93/04/28 93/04/28 93/04/28 93/04/28 93/04/28 93/05/19 93/05/19 93/05/19 93/05/19 93/05/19	047 MEDIUM DAY MEDIUM 1605 WATER 1607 WATER 1607 WATER 1611 WATER 1611 WATER 1611 WATER 1617 WATER 1617 WATER 1531 WATER 1532 WATER 1533 WATER 1534 WATER 1535 WATER 1535 WATER 1535 WATER 1535 WATER 000 WATER 0015 WATER 030 WATER 030 WATER 040 WATER 045 WATER	OR DEPTH (H) 0.3 1.5 2.2 6 8 8.4 0.3 1.5 2.2 6 8 8.8 9.3 0.3 1.5 2.5 4 6 8 8.5 0.3 1.5 2.5 4 6 8 8.5 9.3 0.3	STREAM Flow CFS	WATER TEMP CEWY 19.1 17.7 17.3 17.3 16.7 16.7 16.7 16.7 23.4 22.6 21.7 21.3 22.6 21.7 21.3 23.5 23.5 25.0 22.6 22.6 21.7 21.6 22.6 22.6 22.6 22.6 22.6 23.5 23.6 23.6 23.6 22.6 22.6 22.6 22.6 22.6	DO	DO SATUR	PH	CHDUCTVY FIELD HICRONHO 160 164 164 164 163 161 163 152 151 163 152 151 148 148 147 157 137 137 137 137 137 137 137 137 137 13
93/08/10 11 93/08/10 10 93/08/10 10 93/08/10 10 93/08/10 10 93/08/10 10 93/09/15 09 93/09/15 09 93/09/15 09 93/09/15 09 93/09/15 09 93/09/15 09	021 WATER 023 WATER 025 WATER 027 WATER 027 WATER 029 WATER 031 WATER 047 WATER 040 WATER 050 WATER 051 WATER 051 WATER 051 WATER	1 1.5 4 6 8 9 0.3 1 1.5 4 6 8 9	·	23.3 23.2 22.6 22.5 22.5 22.5 22.5 22.5 22.1 22.0 21.9 21.8 21.8 21.8	7.6 7.7.4 7.4 7.4 7.9 7.8 7.8 7.8 7.8 7.8 7.8 7.8 7.8 7.8	87.45 87.45 88.51 84.15 84.15 83.05 89.85 88.65 88.65 88.65 88.65 88.65 88.65 88.65 88.65 88.65 88.65 88.65	7.30 7.30 7.20 7.10 7.10 7.10 7.10 7.10 7.10 7.10 7.1	119 114 117 122 123 122 122 122 100 100 100 100 103 103 103 101

/TYPA/ANBHT/STREAM

STORET RETRIEVAL DATE 94/05/26 477512 HRM 008.50 35 21 38.0 084 53 30.0 2 CHICKANAUGA RESERVOIR - 2 HILES ABOVE HIGHWAY 58 47121 TENHESSEE RIVER BASIN 44050 47125 CHICK 950501 HIVASSEE RIVER 8.5 1311VAC 930501 06020002 /TYPA/AMBHT/STREAM 0000 RETERS DEPTH

INDEX 1021500 007720 00920 6824 MILES 0953.80 0046.50 499.70 008.50

DATE From To	TIME OF DAY	MEDIUN	SHK OR DEPTH (M)	84002 CODE GENERAL REMARKS	00078 TRANSP SECCHI METERS	31616 FEC COLI MFM-FCBR /100ML	OOOBO Color Pt-co Units	00530 RESIDUE TOT NFLT MG/L	00605 Org n N Ng/l	00610 NH3+NH4- N Total Mg/L	00630 H02&H03 N-TOTAL MG/L
93/04/28			0.3		1.25	300					NU/L
93/04/28 93/04/28	1621	WATER	8.4				100	7	.170	.090	. 18
93/05/19 93/05/19	1530	WATER	0.3		1.13	10K	159	12	.110	.100	. 18
93/05/19	1539	WATER	8.8				15 15	8 10	.120	.030	. 11
93/06/16 93/06/16	1000	WATER	0.3		.85			10	.040	.130	.11 .21
93/06/16	1035	WATER	8				10 15	10 20	.070	.100	. 18
93/07/13 93/07/13	1010	VERT	0.3		. 77	10K			.100	.100	. 20
93/07/13 93/08/10	1025	WATER	8.5				15 20	10 18	.190	.090	. 14
93/08/10	1026	VERT	0.3		.80	10K				.110	. 17
93/08/10 93/09/15	0947	WATER	0.3		. 80	10K	15 15	8 16	.420	.010	. 12
93/09/15 93/09/15	0952 0954	VERT WATER	8			TOK	15 15	14	.110	.070	.12

DATE FRON TO	TIME OF DAY	MEDIUN	SMK OR DEPTH (M)	84002 CODE GENERAL REMARKS	00665 Phos-tot Mg/l P	D0671 PHOS-DIS Ortho Mg/L P	00680 T ORG C C MG/L	32211 CHLRPHYL A UG/L CORRECTD	32212 Chlrphyl B Ug/L	32214 CHLRPHYL C UG/L	32218 PHEOPHTN A UG/L
93/04/28 93/04/28 93/05/19	1621	WATER	8.4		.050	.005	2.5	5.00	1.00K	1.00K	1.00K
93/05/19 93/06/16	1539	WATER	8.8		.040 .050 .040	.003	2.5	12.00	1.00K	1.00	1.00
93/06/16 93/07/13 93/07/13	1010	VERT	8		.050	.010	1.9	3.00	1.00K 1.00K	1.00%	1.00
93/08/10 93/08/10	1026	VERT	8.5 4 8		.040 .040 .060	.005 .010	2.6	4.00	1.00K	1.00	2.00
93/09/15 93/09/15	0952	VERT	4 8		.030	.020 .004 .005	2.7 1.9 1.8	4.00	1.00K	1.00	1.00K

بيعفى المتحديك بعدر والتدويس

	94/03/24 ******
475317 JS JB 10.0 084 WATTS BAR RESERV 47121 TENNESSEE TENNESSEE RIVER BASIN TENNESSEE RIVER 531.0	0 2 OPPOSITE LOWER BRIDGE NEIGS 040801 06010201002 0002.040 ON
0000 HETERS DEPTH	

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/TYPA/AMBHT/FISH/STREAM/SOLIDS

STORET RETRIEVAL DATE 94/05/26 473317 1089 35 38 10.0 084 47 06.0 2 WATTS BAR RESERVOIR - OPPOSITE LOWER BRIDGE 47121 TENNESSEE MEIGS TENNESSEE RIVER BASIN 040801 TENNESSEE RIVER 531.0 131TVAC 06010201002 0002.040 ON

/TYPA/AHBHT/FISH/STREAH/SOLIDS

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INDEX 1021500 007720 00920 Hiles 0953.80 0046.50 531.00

DATE TINE FROM OF TO DAY MEDIUM 93/04/28 1405 WATER 93/04/28 1415 VERT 93/04/28 1439 WATER	SHK OR DEPTH (H) 0.3	84002 CODE GENERAL REMARKS	00078 TRANSP SECCHI METERS 2.00	31616 FEC COLI MFM-FCBR /100HL 10K	00080 Color Pt-Co UNITS	00530 RESIDUE Tot NFLT Kg/L	00605 Org N H Mg/L	00610 NH3+NH4- N TOTAL NG/L	00630 NO2£NO3 N-TOTAL MG/L
93/05/20 1313 WATER 93/05/20 1317 VERT 93/05/20 1329 WATER 93/06/17 1255 WATER 93/06/17 1355 VERT 93/06/17 1340 WATER 93/07/14 1325 WATER 93/07/14 1327 WATER 93/07/14 1327 WATER	23.4 0.3 4 23.6 0.3 4 23 0.3 0.3 0.3 0.3	D 1 D 2	1.50 1.60 1.67	10K 10K	100 50 5 5 10	6 8 7 5 8	.200 .160 .030 .030 .020K .060	.010 .040 .010 .080 .020 .030	.32 .36 .18 .30 .21 .41
93/07/14 1343 VERT 93/07/14 1345 VERT 93/07/14 1347 VERT 93/07/14 1415 WATER 93/07/14 1415 WATER 93/07/14 1410 WATER 93/08/11 1313 VERT 93/08/11 1313 VERT 93/09/15 1152 VERT 93/09/16 1151 VERT 93/09/16 1150 VERT 93/09/16 1202 WATER	23.5 23.5 23.5 23.5 23.5 23.5 23.5 24 0.3 4 0.3 23.5	D3 D1 D2 D3 D1 D2 D3	1.30 1.20	10K 10K 10K 10K	10 5 10 5 10 10 5 5 5 5 5 5	4 3 12 9 10 4 11 3 3 11	.290 .290 .310 .230 .190 .160 .250 .230 .230 .250	.010x .010x .010x .010 .010 .020 .010x .010x .010x	.02 .02 .02 .46 .45 .44 .10 .38 .13 .13

2.3. 時間の日本市においても思いないできた。ことは、ことをついても、

93/05/20 93/05/20 93/06/17 93/06/17 93/07/14 93/07/14 93/07/14 93/07/14	1329 WATER 1305 VERT 1340 WATER 1325 WATER 1327 WATER 1329 WATER	23.4 23.6 23.6	84002 CODE GENERAL REMARKS D1 D2 D3 D1	00665 PHOS-TOT MG/L P .020 .030 .020 .030 .020 .040	00671 PHOS-DIS ORTHO MG/L P .004 .003 .005 .002x .020	00680 T ORG C C MG/L 1.9 1.8 2.2 1.6 2.2 1.7	32211 CHLRPHYL A UG/L CORRECTD 10.00 5.00 4.00	32212 CHERPHYL 9 0G/L 1.00 1.00K 1.00	32214 CHLRPHYL UG/L 2.00 1.00	32218 PHEOPHTN UG/L 1.00 1.00K 2.00
93/07/14 1 93/07/14 1 93/07/14 1 93/08/11 1	347 VERT 415 WATER 417 WATER 419 WATER 313 VERT 327 WATER	23.5 23.5 23.5 23.5	02 03 01 02 03	.020 .020 .010 .040 .040 .030 .030	.002K .002 .002 .020 .020 .020	2.3 2.4 2.5 1.7 1.7 1.7	8.00 9.00 9.00	1.00K 1.00K 1.00K	1.00 1.00 1.00	1.00 1.00 1.00
93/09/16 11 93/09/16 11 93/09/16 12	51 VERT	23.5		.050 .020 .020	.003 .010 .004 .004	2.5 1.6 2.2 2.2	10.00	1.00K	1.00	3.00
				.040	.010	2.1	5.00	1.00	1.00	4.00

STORET RETRIEVAL DATE 94/05/26 476041 1114C 35 49 50.0 084 36 33.0 2 Watts Bar Reservoir 47145 Tennessee Roare Roane Tennessee River Basin 040801 Tennessee River 560.80 131174C Ro 06 0000 Heters Depth ROANE 040801 HQ 06010201002 0043.170 OFF

/TYPA/AMBNT/STREAM/SOLIDS

INDEX 1021500 007720 00920 Miles 0953.80 0046.50 560.80

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WILE? UN	55.00	0040.00	500.00						
DATE	TIME		SMK OR	00060 STREAN FLOW	00010 WATER TENP	00300 D0	00301 DD Satur	00400 PH	00094 CNDUCTVY FIELD
FROM TO	OF DAY	NEDIUN	DEPTH (N)	CFS	CENT	MG/L	PERCENT	S U	HICRONHO
					16.7	11.5	118.6\$	8.40	165
93/04/28	1210	WATER	0.3		16.3	11.4	114.05	8.40	165
93/04/28	1215	WATER	1.5		16.0	11.3	113.0\$	8.30	164
93/04/28		WATER	1.3		15.7	11.2	112.0\$	8.30	164
93/04/28		WATER	2		15.3	10.7	104.95	8.20	171
93/04/28		WATER	6		15.2	10.5	102.95	8.10	177
93/04/28		WATER	8		15.1	10.4	102.0\$	8,10	179
93/04/28		WATER	10		. 14.8	10.3	101.05	8.10	186
93/04/28		WATER	12		14.4	10.0	96.25	8.00	206
93/04/28		WATER	12.5		14.4	10.0	96.2\$	8.00	195
93/04/28		WATER WATER	0.3		21.7	9.Z	104.55	7.70	179
93/05/20			1		21.7	9.1	103.4\$	7.70	179
93/05/20		WATER	1.5		21.7	9.1	103.4\$	7.70	180
93/05/20		NATER			21.0	8.3	92.25	7.40	179
93/05/20		UATER	6		20.7	8.2	91.15	7.40	174
93/05/20		WATER	ě		20.2	7.8	84.8\$	7.20	181
93/05/20			10		19.7	7.6	82.6\$	7.20	183
93/05/20	5 1150	UATER	12		19.3	6.9	73.45	7.10	183
93/05/20	5 1151	WATER	12.6		19.2	7.0	74.55	6.90	183 185
93/05/20	5 1152	WATER	13.1		19.1	6.9	73.45	7.00	195
93/06/1	7 1047	WATER	0.3		27.1	11.1	137.05	8.70 8.60	195
93/06/1	7 1050	WATER	1		26.2	10.9	132.95	8.60	194
93/06/1	7 1055	WATER	1.5		25.9	10.3	125.6\$ 120.7\$	8.50	192
93/06/1	7 1100	WATER	2		25.7	9.9	106.0\$	8.20	185
93/06/1	7 1105	WATER	2.5		25.4	8.9 8.3	98.8\$	8.00	181
93/06/1	7 1110	WATER .	4		25.0	7.1	83.55	7.60	175
93/06/1	7 1120) WATER	4.5		24.3	6.7	78.8\$	7.50	172
93/06/1	7 1125	WATER	6		23.6	6.1	71.85	7.40	174
93/06/1	7 1130	WATER	8		23.6	6.1	71.85	7.40	170
93/06/1	7 1135	WATER	10		23.3	5.1	58.6\$	7.30	163
93/06/1	7 1140	WATER	12		23.2	4.6	52.95	7.20	166
93/06/1	7 1145	WATER	12.5		23.2	4.0	46.0\$	7.20	162
93/06/1		WATER	13		29.8	10.3	135.5\$	8.80	205
93/07/1	4 1125	WATER	0.3		29.7	10.6	139.5\$	8.90	206
93/07/1	4 1129	Y WATER	1.5		29.5	10.5	134.6\$	8.90	204
93/07/1		S WAIER	1.3		29.5	10.4	133.3\$	8.70	205
93/07/1		7 WATER	í		29.4	10.1	129.5\$	8.80	205
93/07/1	9 119	I WALER	3.5		29.2	9.5	121.8\$	8.70	207
93/07/1	4 114	S WATER	3.5						

DATE From	TINE		SMK OR DEPTH	00060 Stream Flow	00010 WATER TEMP	00300 D0	00301 00 Satur	00400 PH	00094 CNDUCTVY field
TO	DAY	NEDIUM	(H)	CFS	CENT	MG/L	PERCENT	SU	NICRONHO
93/07/14			,		28.6	9.2	117.9\$	8.60	208
			4.5		27.7	7.3	92.4\$	8.00	214
93/07/14		WATER	4.5		27.0	7.1	87.75	7.90	210
93/07/14		WATER	6.5		25.8	6.5	79.35	7.60	201
93/07/14		WATER	0.5		25.0	5.8	69.05	7.40	200
93/07/14			10		24.1	5.2	61.25	7.30	192
93/07/14			12		23.6	4.6	54.15	7.20	194
93/07/14			12.5		23.6	4.5	52.95	7.20	191
93/07/14			0.3		25.5	8.3	98.85	8.10	202
93/08/11		WATER WATER	0.3		25.2	8.1	96.45	8.10	201
93/08/11			1.5		25.1	8.0	95.25	8.00	200
93/08/11		WATER	1.2		24.9	7.4	88.1\$	7.90	200
93/08/11			÷,		24.2	6.7	78.85	7.70	199
93/08/11		WATER	2		23.9	6.5	76.55	7.60	199
93/08/11		WATER	8		23.8	6.5	76.55	7.60	200
93/08/11		WATER	10		23.6	6.2	72.95	7.60	200
93/08/11		WATER	12		23.6	6.0	70.65	7.60	203
93/08/11		WATER	12.5		23.5	5.7	65.5\$	7.50	205
93/08/11		WATER	12.5		23.3	5.2	59.85	7.50	206
93/08/11		WATER	0.3		24.2	7.3	85.95	7.70	203
93/09/16	1050	WATER	0.3		24.1	7.1	83.5\$	7.70	203
93/09/16		WATER	1.5		24.1	7:1	83.55	7.70	203
93/09/16		WATER WATER	1.3		24.1	7.1	83.5\$	7.70	203
93/09/16		WATER	4		24.1	6.9	81.25	7.70	203
93/09/16		WATER	2		24.1	6.9	81.2\$	7.60	201
93/09/16		WATER	6 8		24.1	6.9	81.25	7.60	209
93/09/16			10		24.0	6.8	80.05	. 7.60	207
93/09/16		WATER	12		23.7	6.3	74.15	7.50	218
93/09/16		WATER			23.7	6.2	72.95	7.50	209
93/09/16		WATER	12.4			6.1	71.85	7.50	214
93/09/16	1101	WATER	13		23.7	0.1	11.03	. 1.30	214

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STORET RETRIEVAL DATE 94/05/26 476041 1114C 35 49 50.0 084 36 33.0 2 WATTS BAR RESERVOIR 47145 TENMESSEE ROANE TENMESSEE RIVER BASIN 060801 1811VAC HQ 061 0000 METERS DEPTH H9 06010201002 0043.170 OFF

/TYPA/AMBNT/STREAM/SOLIDS

INDEX 1021500 007720 00920 MILES 0953.80 0046.50 560.80

DATE From To	T I ME OF DAY	NEDIUM	SHK OR DEPTH (H)	84002 CODE GENERAL REMARKS	00078 TRANSP Secchi Meters	31616 FEC COLI MFH-FCBR /100HL	00080 Color PT·CO UHITS	00530 Residue Tot nflt Mg/l	00605 ORG N N Hg/L	00610 NH3+NH4- N Total Ng/l	00630 No24No3 N-Total	
93/04/28	1210	WATER	0.3		1.52	10K				No/C	MG/L	
93/04/28 93/04/28	1235	VERT				100	5 a	я	.150			
93/05/20 93/05/20	1142	VATER	12.5		1.00	10K	5 Q	8 9	.090	.010	.34 .42	
93/05/20	1151	VATER	12.6				10	8	.040	.020	.28	
93/06/17 93/06/17	1047	WATER	0.3	•	1.15	10x	5	10	.180	.060	.35	
93/06/17	1145	WATER	12.5				10 10	6 15	.090	.020	. 19	
93/07/14 93/07/14	1125	WATER	0.3		1.33	10 K	10	15	.040	.050	. 29	
93/07/14	1217	VATER	12				10	5 14	.320	.030	. 13	
93/08/11 93/08/11	1118	VATER	0.3		1.20	10K	5	14	.200	.040	. 3 9	
93/08/11	1129	VATER	13				10	5	.470	.010K	.27	
93/09/16	1055	VERT	0.3		1.20	10K	tĸ	6	.330	.020	.30	•
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							,	14	.250	.060	. 20	

DATE FROM TO	TIME OF Day	NEDIUN	SMC OR DEPTH (M)	84002 CODE GENERAL REMARKS	00665 Phos-tot Ng/l P	00/71 PHOS-DIS ORTHO MG/L P	00680 T ORG C C MG/L	32211 CHLRPHYL A UG/L Correctd	32212 CHLRPHYL 8 Ug/L	32214 CHLRPHYL C UG/L	32218 PKEOPHIN A Ug/L
93/04/28 93/04/28	1300	WATER	12.5		.030	.002	1.9	9.00	1.00	1.00	2.00
93/05/20 93/05/20 93/06/17	1151	WATER	12.6		.030 .040	.003	2.0 1.8	11.00	1.00K	2.00	2.00
93/06/17 93/07/14	1145	VATER	12.5		.040	.002K .004 .003	2.0 1.7 2.2	10.00	2.00 1.00K	2.00	5.00
93/07/14 93/08/11 93/08/11	1123	VERT	12		.050	.007 .002K	1.8	3.00	1.00	1.00	2.00
93/09/16 93/09/16	1055	VERT	12.4		.030 .030 .040	.005 .005 .008	1.7 2.0 1.8	5.00	1.00	1.00K	4.00

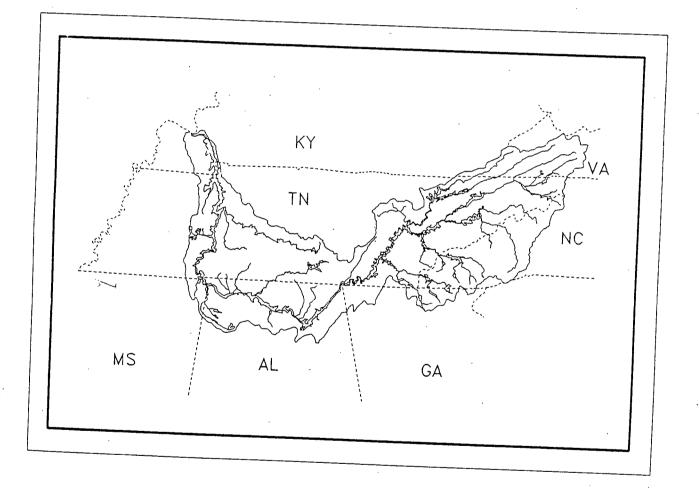
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Tennessee Valley Authority

Water Resources Division Chattanooga, Tennessee

TVA/WR/AB--91/4 May 1991

RESERVOIR VITAL SIGNS MONITORING - 1990 FISH COMMUNITY RESULTS



WATER RESOURCES & ECOLOGICAL MONITORING WATER RESOURCES MANAGEMENT

Introduction

The Tennessee Valley Authority (TVA) operates 9 reservoirs on the Tennessee River and 37 reservoirs on its tributaries. TVA is committed to maintaining the health of aquatic resources created when the reservoir system was built. To that end, TVA in cooperation with Valley states, operates a water resource monitoring program that includes physical, chemical, and biological data collection components. Biological monitoring will target the following selected elements within three zones of the reservoir (inflow, transition, and forebay):

- Sediment/Water-column Acute Toxicity Screening
- Benthic macroinvertebrates
- Fish

Reservoir fisheries monitoring is divided into the following activities:

- Fish Biomass
- Fish Tissue Contamination
- Fish Community Monitoring
- Fish Health Assessment

This report presents the results of fish community monitoring and fish health assessments. Reports on other components and activities are published in companion reports, and a summary report on the results of the monitoring program also is available.

Fish Community Monitoring

The basic ecological principle underlying the community monitoring program is that characteristics of fish populations, because of their trophic status, will reflect changes in the entire aquatic ecosystem. The program's objective is to provide the minimum information necessary to evaluate the status of the fish community at inflow, transition zone, and forebay areas of reservoirs throughout the Tennessee Valley. The information gathered is used to:

- Screen for significant differences from average conditions
- Detect long-term trends

Introduction

- Aid in establishing project priorities
- Trigger intensive sampling to determine causes and solutions where problems are identified

Fish Health Assessment Index

The general health of aguatic communities is a reflection of the guality of the water and habitat. The primary objective of fish health assessment was to develop baseline information on the health of populations of a top predator species represented in fish communities in reservoirs throughout the Tennessee Valley. Future samples will allow detection of year-to-year variations and long-term trends. The survey utilized a method developed by Goede (1991), to assess the general health and condition of a fish population. To better reflect existing conditions the method was modified by TVA into an index known as the Fish Health Assessment Index (FHAI). The largemouth bass was selected as the initial species to investigate in the Tennessee Valley because of ease of capture, widespread distribution, and position on the food chain (top predator). Results of ordered observations of both external and internal condition of individuals were entered into a portable computer and analyzed using a program (AUSUM430) developed by TVA. Abnormalities were tabulated and mathematically weighted in the calculation of the FHAI according to severity, such that as debilitating anomalies increase, the FHAI also increases. Thus better water quality is indicated by a lower FHAI. Comparisons can then be made among individual fish, size-groups of fish, sites, groups of sites, reservoirs, and/or groups of reservoirs.

The autopsy based FHAI is based on the following assumptions (Goede, 1991):

- If the appearance of the organ and tissue systems is "normal" the fish is normal.
- In response to environmental stresses some change in function of organs and tissues is necessary for homeostasis or maintenance of internal environment of the fish.
- If change in function persists because of continuing environmental stress(es), there will be change or modification of structure that is observable as a gross change in organs or tissues.
- Observable changes in structures of certain tissues and organs indicate s adaptive change in response to extrinsic environmental stresses.
- If appearance of the organ and tissue systems shows departure from normal, the fish is responding to intrinsic changes brought about by extrinsic environmental stresses.

Introduction

While these assumptions generally hold true for chronic environmental stresses, there are occasions where there is (or has been) environmental stresses that yield no observable changes in gross tissue structure or appearance. Two examples are as follows:

- Changes in function may be sufficiently acute and severe that mortality occurs before gross observable change in structure and/or appearance occurs.
- Microscopic, histological structural change may occur without gross manifestation.

Degrees of "normal" or nature of "normal" are subject to some interpretation. "Normal" must be considered relative to age, sex, season, species, etc. For example, "normal" level of fat storage of healthy, free-ranging fish entering the winter season is much higher than from the same fish coming out of the winter. Results must be compared with established norms. The autopsy-based system of fish health assessment index is *not* a diagnostic tool. If it indicates that a population of fishes in a given reservoir or location is unhealthy, further investigation (e.g. histological examination of the tissue by a pathologist) may be required to determine what is causing the fish to be unhealthy.

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Methods

Fish Community Monitoring

Fourteen reservoirs were studied during fall 1990 including: Kentucky, Pickwick, Wilson, Wheeler, Guntersville, Nickajack, Chickamauga, Watts Bar, Fort Loudoun, Tellico, Melton Hill, Norris, Cherokee, and Douglas. Shoreline electrofishing samples were collected during daylight hours from inflow, transition, and forebay zones of each reservoir from late September to December, 1990. The forebay area was defined as the main channel shoreline extending into the "mouths" of tributary streams within approximately three miles above the dam. The transition zone is the main reservoir reach where hydraulic and water chemistry conditions suggest a shift from a riverine to a reservoir environment. The inflow site was in the tailwater of the upstream dam or in the free-flowing stream(s) entering reservoirs farthest upstream on the tributaries. The September-December time frame was chosen as sampling all reservoirs requires an extended period of time, and distribution of fish populations is most stable in fall. A total of 10 electrofishing transects (10 minutes duration each) was sampled within each of the 42 sites (14 reservoirs). All habitats were sampled at each site, with dominant types receiving the most effort. Habitat distinctions were based on major changes in substrate (e.g., rock, riprap, or clay), and/or cover (e.g., brush or aquatic vegetation). Sample size/duration was selected based on sensitivity analysis of largemouth bass electrofishing samples in two Oklahoma reservoirs (Gilliland 1985).

Black bass species captured were measured and weighed, other major sport fish species and channel catfish were measured, and all other species were enumerated prior to release. All fish were examined for obvious external abnormalities, diseases, and parasites, and this information was noted. Fish observed but not captured were included in the records if positive identification and enumeration could be made. Estimates of young-of-year (YOY) numbers were made in instances where high densities were encountered (i.e., YOY shad and bluegill). However, estimates of these species were not made in 1989, so year to year comparisons involving shad, bluegill, total fish abundance, and species composition are not possible in this report.

Where conditions permitted, ten 100-ft-monofilament experimental gill nets with five 20-ft panels (mesh sizes of 1, 1 1/2, 2, 2 1/2, and 3 inch bar mesh) were set for one overnight period in each reservoir zone. In forebay and transition zones, nets were set in all habitat types, alternating mesh sizes toward the shoreline between sets. In inflow areas, nets were set wherever flow conditions allowed. Availability of adequate sample locations limited number of nets set to less than ten at some inflow stations.

Analyses of data included comparison of total catch rates with historical catch rates for the particular reservoir grouping and among zones within reservoirs, comparison of functional group composition, calculation of proportional stock density (PSD) and relative stock density (RSD) for principal species, and calculation of mean relative weight (Wr) for black bass species captured at each sample area. The PSD/RSD analyses compare the number of fish attaining various size categories with the total number of catchable-sized individuals of a given species (Anderson and Weithman 1978). Four size categories: quality, preferred, memorable, and trophy are based upon percentages of maximum attainable lengths (Gabelhouse 1984). Catchable or stock length includes all individuals measuring 25 percent or more of the maximum length. Quality, preferred, memorable, and trophy size fish are at least 37, 46, 59, and 74 percent of the maximum length, respectively (Appendix A). PSD, therefore is the percentage of the total catchable population of a given species represented by individuals of quality size. Relative stock densities (RSD1, RSD2, and RSD3) are percentages of preferred, memorable, and trophy sizes, respectively.

Relative weight analysis involves the development of standard weight tables from historical length-weight information, that identify "expected" weights of fish at different lengths (Anderson and Weithman 1978). Fish having "expected" weights will have Wr values of 100, while those heavier will have Wr values greater than 100. Standard weight tables were generated for five groupings of TVA reservoirs: mainstream Tennessee River, Tennessee storage, North Carolina-Georgia storage, Cherokee, and Douglas (Appendix B).

The 1990 observations (e.g., catch rates, PSD/RSD, Wr) from each reservoir were compared with averages from groupings of similar reservoirs just mentioned. The groupings were determined by principal component analysis of historical rotenone data from 25 TVA reservoirs based on the numerical abundance of the ten most frequently occurring fish species (Tom McDonough, pers. comm.).

Fish Health Assessment

A FHAI was calculated for the three locations and inflow) in each reservoir. Largemouth bass were collected as part of the electrofishing survey and were transported to a mobile laboratory for examination. Fifteen individuals greater than length were 250 mm total length were examined from each of the three locations in each reservoir. Attempts were made to minimize holding time to reduce handling stress which can alter blood chemistry results. Upon arrival at the mobile laboratory, fish were anesthetized with tricaine methanesulfonate (MS-222) in 50 mg/l (ppm) solution.

Fish were processed in the manner described in detail by Goede (1991). The body cavity of the fish was opened using sharp/blunt-ended surgical scissors by making a ventral incision from the vent forward to the pectoral girdle, cutting closely to one side of the pelvic girdle. Care was taken not to make the incision deep enough to damage internal organs. Blood was then collected by cardiac puncture and prepared for determination of using a hematocrit tube which was then sealed and centrifuged to allow measurement of hematocrit, leucocrit, and plasma protein levels. The liver was examined immediately for

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anomalies because of the tendency for rapid discoloration following death of the individual.

An external examination was initiated following an ordered sequence of eyes, skin, gills, fins, opercles, pseudobranchs, and thymus. Finally, an internal examination was conducted, following an ordered observation sequence of mesenteric fat, spleen, hindgut, kidney, bile, parasites, and determination of sex. The abnormalities sought are listed in table 6 for each reservoir. Abnormal conditions and other data were entered directly into a computer program developed by TVA entitled "AUSUM430". Results were calculated and printed for immediate use.

Summary Comparisons

Thirty-four selected measurements of the fish communities observed at each reservoir were compared to assess the relative health of the fisheries environment within each reservoir and each reservoir location (inflow, transition zone, and forebay). Table 7 in each of the following reservoir chapters summarizes these comparisons. Nine categories were selected for comparison: species richness, fish health, trophic composition, sensitive species, electrofishing and gill netting catch rates, relative weight of Micropterus spp., proportional stock densities of selected species, and overall fish abundance. The lower eight mainstream reservoirs (Kentucky to Watts Bar) were compared individually to the average values of the 34 measurements for that group of reservoirs. Similarly, Fort Loudoun and five Tennessee storage reservoirs were compared individually with the averages for that group. Results for 1990 were compared with 1989 results for each reservoir where 1989 data were available. Sampling stations within each reservoir were compared to average values of like stations for the two reservoir groups in 1990. In each comparison a condition considered more healthy than the group average was designated with a '+' sign. A condition less healthy than the average received a '-' sign. Values falling within + or - five percent of the average were given a '0.' Whenever sufficient observations for meaningful comparisons were lacking, blanks appear in the table. In most cases values higher than the group average were considered more healthy. However, this was not the case for the percentages of omnivores and tolerant species, the catch rates of gizzard shad, and the fish health assessment index. The columns of comparisons were summed to determine the overall condition of the fish community compared to the group average and the previous year. Columns with summary values greater than +2 were designated 'Better', those less than -2 were designated 'Worse,' and those within + or - 2 were designated 'Average.'

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Results - Chickamauga Reservoir

Fish Community Assessment

A total of 10,070 fish, including 41 species, were captured by electrofishing and gill net sampling in 1990. Electrofishing accounted for 34 species in 5.0 hours of electrofishing and gill netting resulted in capture of 27 species in 30 net nights (table 1). The forebay zone accounted for 4881 of the total fish collected by electrofishing and 219 of the fish obtained with gill nets (table 2). No endangered or threatened species were collected. Of the taxa collected, 1 was parasitic, 1 was planktivorous, none were herbivorous, 15 were insectivorous, 5 were benthic insectivorous specialists, 6 were omnivorous, and 13 species were piscivorous (table 3).

Catch Rates of Selected Species

Comparisons of fall 1990 electrofishing catch rates of selected species with those of fall 1989 and overall average catch rates for these species in Tennessee River mainstream reservoirs (table 1) indicate largemouth bass were more numerous during fall 1990 (52.6) than fall 1989 (31.8), and were also more abundant than the average for the reservoir grouping (27.8). Smallmouth bass were more abundant during fall 1990 (2.4) than fall 1989 (0.4), but are less abundant in Chickamauga Reservoir than the average for the mainstream reservoir grouping (7.9). Spotted bass were more numerous during fall 1990 (25.6) than fall 1989 (12.0), and were also more abundant than the average for the reservoir grouping (12.7). Bluegill were more numerous during fall 1990 (259) than fall 1989 (129), and were also more abundant than the average for the reservoir grouping (133). Redear sunfish abundance declined during fall 1990 (33.6) from fall 1989 (58.4), but were still more abundant than the average for the reservoir grouping (16.9). Channel catfish were relatively uncommon in this reservoir, as evidenced by a catch rate of 1.8 fish per hour during 1989 and 1990, which is slightly less than the reservoir group average (2.5). Gizzard shad, 260 per hour during fall 1990, were less abundant than the average for the reservoir grouping (757). Whereas, threadfin shad (980) were on par with the average for the reservoir grouping (955).

The abundance of three selected groups from experimental gill netting (channel catfish, *Morone* spp, and *Stizostedion* spp.) was compared to fall 1989 and the reservoir group average catch rates (table 1). The comparisons indicate channel catfish abundance (1.2) was similar to the reservoir group average (1.3), and had tripled since the 1989 sample. The catch rate of combined *Morone* spp., 4.8 per net night, was equal to the mainstream reservoir

Results - Chickamauga Reservoir

group average (4.9), and was approximately three times greater than the 1989 abundance at Chickamauga Reservoir (1.7). Stizostedion spp., however, were rare in the fall samples of both years, 0.1 per net night, and were below the average abundance in mainstream reservoirs (0.5).

Comparisons of electrofishing catch rates among the three reservoir stations (table 2) showed largemouth bass were most abundant in the forebay area of the reservoir with a catch rate of 75.0 fish per hour, compared to 55.8 and 27.0 in the transition and inflow areas, respectively. Smallmouth bass were found in equal abundance (1.2) at the inflow and transition areas of Chickamauga Reservoir, but were more abundant in the forebay area (4.8). Spotted bass were more abundant in the forebay area with a catch rate of 67.8 fish per hour compared to the transition, 9.0 per hour, and were absent from the inflow area. Bluegill were most abundant in the forebay area of the reservoir with a catch rate of 543 fish per hour, compared to 154 and 81.6 in the transition and inflow areas, respectively. Redear sunfish were most abundant in the inflow area of the reservoir with a catch rate of 55.8 fish per hour, compared to 30.0 and 15.0 in the forebay and transition areas, respectively. Channel catfish were found in equal abundance (1.2) at the inflow and forebay areas of Chickamauga Reservoir, but were more abundant in the transition area (3.0). Gizzard shad were most abundant in the forebay area of the reservoir with a catch rate of 359 fish per hour, compared to 326 and 94.2 in the transition and inflow areas, respectively. Threadfin shad were found in comparable abundance in the transition and forebay areas of the reservoir with catch rates of 1605 and 1335, respectively, but were absent from the inflow area.

Variations in the spatial abundance of channel catfish, *Morone* spp., and *Stizostedion* spp., gizzard shad, and threadfin shad based on gill net samples revealed found channel catfish were equally abundant in the inflow and forebay areas of the reservoir with a catch rate of 1.4 fish per net-night, compared to 0.8 in the transition area. Combined *Morone* spp. were found in greater abundance in the inflow (5.1) and forebay (6.1) areas, than the transition area (3.2). The small numbers of *Stizostedion* were found in the inflow and transition areas, which had catch rates of 0.1 fish per net night.

Relative Abundance of Selected Groups

Trophic composition of Chickamauga Reservoir based on the combined electrofishing and gill netting catch showed the following relationships. The dominant trophic group by number was planktivores, represented by 49 percent of the sample. Following them in abundance were insectivores, which comprised 27 percent of the sample, omnivores (16 percent), piscivores (8 percent), specialist (1 percent), and parasites (tr.).

Shad were the most abundant taxonomic grouping encountered, as 3 species represented 63 percent of the total sample by number. Sunfishes, excluding *Micropterus* spp., represented 18 percent of the sample with 8 species. The 5 species of suckers amounted to 0.4 percent of the sample. Catfish (3 species) were 0.8 percent of the sample. There were 3 species of black bass (*Micropterus* spp.) present, which represented 4.2 percent of the sample.

Results - Chickamauga Reservoir

Cyprinids (minnow family) comprised 1.0 percent of the sample with 1 species. There were 9 small cyprinid and darter species observed.

Species tolerant of degraded conditions, which included spotted gar, gizzard shad, golden shiner, and spotfin shiner, accounted for 14 percent of the sample. Species considered intolerant to pollution included spotted sucker, black redhorse, and longear sunfish accounted for 0.6 percent of the sample by number.

Relative Weights

Relative weight (Wr) indices for black bass species sampled at Chickamauga Reservoir during fall 1990 were compared with those obtained in fall 1989, whenever possible (table 4). There was little difference in relative weights of largemouth bass in fall 1990 (99) and fall 1989 (97). Smallmouth bass were not sampled in adequate numbers (15 or more) to draw useful conclusions about the relative weight of this species in this reservoir. There was little difference in relative weights of spotted bass in fall 1990 (88) and fall 1989 (86).

Largemouth bass had a higher relative weight in the forebay area (101) than the inflow area (94), but approximately the same as the transition area (98). Spotted bass had a higher relative weight in the inflow area (93) than the forebay area (87), but approximately the same as the transition area (90).

Proportional and Relative Stock Densities

The size distribution of important sportfish species is described by proportional (PSD) and relative (RSD1-3) stock densities (table 5). The percentage of quality-sized channel catfish sampled during fall 1990 at Chickamauga Reservoir was shown as a PSD value of 64. This value is essentially that that observed during fall 1989 (67). Channel catfish in the preferred size range or larger were slightly less abundant in fall 1990 (RSD1 = 8) than fall 1989 (10). Channel catfish in the memorable size range or larger were not found during 1989 or 1990.

The percentage of quality-sized bluegill sampled during fall 1990 at Chickamauga Reservoir was shown as a PSD value of 24. This value is greater than that observed during fall 1989 (15). Bluegill in the preferred and memorable size ranges were found in fall 1990 (RSD1 = 1, RSD2 = 1), but were not found in fall 1989. Bluegill in the trophy size range were not found during 1990.

The percentage of quality-sized redear sunfish sampled during fall 1990 at Chickamauga Reservoir was shown by a PSD value of 43. This value is approximately the same as that observed during fall 1989 (46). Redear sunfish in the preferred size range or larger were slightly more abundant in fall 1990 (RSD1 = 8) than fall 1989 (6). Redear sunfish in the memorable size range were found in fall 1990 (RSD2 = 1), but were absent in fall 1989. Redear sunfish in the trophy size range were not found during 1990.

Results - Chickamauga Reservoir

The percentage of quality-sized spotted bass sampled during fall 1990 at Chickamauga Reservoir was indicated by a PSD value of 35. This value is nearly identical to that found in 1989(36). Spotted bass in the preferred size range were more abundant in fall 1990 (RSD1 = 15) than fall 1989 (4). Spotted bass in the memorable size range or larger were not found during fall 1990 or fall 1989.

The percentage of quality-sized largemouth bass sampled during fall 1990 at Chickamauga Reservoir was indicated by a PSD value of 47. This value is very nearly the same as that observed during fall 1989 (45). Largemouth bass in the preferred size range or larger were slightly more abundant in fall 1990 (12) than fall 1989 (10). Largemouth bass in the memorable size range were similar in abundance in fall 1990 (RSD2 = 3) than fall 1989 (2). Largemouth bass in the trophy size range were not found during 1989 or 1990.

Variations in PSD and RSD1-3 values among the three sampling areas of Chickamauga Reservoir were observed. Channel catfish had the highest percentage (PSD = 80) of quality-sized individuals in the transition zone of the reservoir, compared to the inflow (53) and forebay (64) zones. There were more preferred-sized channel catfish in the transition (RSD1 = 20) zone than the forebay (9) zone, but none were found in the inflow zone. No memorable-sized or larger channel catfish were found in the fall 1990 sample. There were no appreciable differences in the percentages of quality-sized bluegill among the inflow (PSD=26), transition (24), and the forebay (22) zones. Preferred-sized bluegill were only found in the transition zone (RSD1 = 2). Memorable-sized bluegill were only found in the transition zone (RSD2= 2). No trophy-sized bluegill were found in the fall 1990 sample at any sampling station. Redear sunfish had the highest percentage (PSD = 81) of quality-sized individuals in the transition zone of the reservoir, compared to the inflow (20) and forebay (42) zones. Redear sunfish had the highest percentage (RSD1 = 19) of preferred-sized individuals in the transition zone of the reservoir, compared to the inflow (4) and forebay (5) zones. Memorable-sized redear sunfish were only found in the transition zone (RSD2 = 3). No trophy-sized redear sunfish were found in the fall 1990 sample. Largemouth bass had a higher percentage (RSD1 = 20) of preferred-sized individuals in the transition zone of the reservoir compared to the forebay zone (8) and the inflow zone (12). memorable-sized largemouth bass were greater at the transition zone (RSD2 = 7) than the inflow (4) and the forebay (1) zones. No trophy-sized or larger largemouth bass were found in the fall 1990 sample.

Fish Health Assessment Index (FHAI)

The FHAI for Chickamauga Reservoir averaging all areas sampled in 1990 was 54.6 (table 6). In comparison to the overall average FHAI for mainstream reservoirs, 48.1, the relative health of Chickamauga Reservoir appears to be worse. Conditions in 1990 appear to have diminished since 1989 when the reservoir average FHAI was 53.5. not appreciably changed since the previous year's value of 53.5.

The FHAI for the inflow on Chickamauga Reservoir was 28.6, and the three most frequently anomalies found involved parasites, spleen, and liver. Com-

Results - Chickamauga Reservoir

pared to the average FHAI for inflow zones of all the reservoirs within the mainstream reservoir group, 39.9, the apparent health of the inflow zone of this reservoir is better. The inflow FHAI for 1990 at Chickamauga Reservoir showed an improvement over the 1989 FHAI of 50.6, when the most frequently encountered anomalies were found in the hematocrit, spleen, and kidney.

The FHAI for the transition on Chickamauga Reservoir was 69.3, and the three most frequently anomalies found involved parasites, spleen, and kidney. Compared to the average FHAI for transition zones of all the reservoirs within the mainstream reservoir group, 52.8, the apparent health of the transition zone of this reservoir is worse. The transition FHAI for 1990 at Chickamauga Reservoir showed declining health of the transition zone since the 1989 FHAI of 54.0, when the most frequently encountered anomalies were found in the hematocrit, liver, and kidney.

The FHAI for the forebay on Chickamauga Reservoir was 66.0, and the three most frequently anomalies found involved parasites, plasma protein, and liver. Compared to the average FHAI for forebay zones of all the reservoirs within the mainstream reservoir group, 51.3, the apparent health of the forebay zone of this reservoir is worse. The forebay FHAI for 1990 at Chickamauga Reservoir showed declining health of the forebay zone since the 1989 FHAI of 56.0, when the most frequently encountered anomalies were found in the hematocrit, liver, and kidney.

Conclusion

In summary Chickamauga Reservoir was found to support a poorer fish community than the average Tennessee River mainstream reservoir during 1990 based on comparison of 34 measurements (table 7). The community has shown improvement over the previous year of sampling. Although the fish community in the inflow zone appears to be in worse condition than the average of inflow zones of mainstream reservoirs, the fish community in the forebay zone appears to be in better condition than the average of forebay zones of mainstream reservoirs. The transition zone supports a fish community approximately equivalent to the average transition community found in mainstream reservoirs.

Results - Chickamauga Reservoir

Table 1.	Species list and catch per unit effort of fishes sampled during fall electrofishing and gill netting on Chickamauga Reservoir, 1990, (stations combined) compared with previous year and averages for Mainstream reservoirs. Total efforts(*)
	shown in parentheses.

	E	lectrofis	shing Tainstream		Gill netting Mainstream			
	1990	1989	Average	1990	1989	Average		
Name	(5.0)	(5.0)	(42)	(30)	(30)	(237)		
Chestnut lamprev	8.8	0.0	1.2	0.0	0.0	0.0		
Spotted gar	0.4	0.8	1.8	0.1	Ť	0.1		
Skipjack herring	0.0	ŏ.ŏ	7.7	3.9	1.2	4.7		
Gizzard shad	259.6	67.0	756.8	2.3	3.2	6.7		
Threadfin shad	980.0	41.4	955.0	0.0	0.0	0.0		
Mooneye	0.0	0.0	Ţ	0.1	0.1	0.1		
Carp Golden shiner	20.2	6.2 1.6	8.3 3.0	0.1	0.0	. 0.4		
Emerald shiner	137.0	0.8	82.3	0.0	0.1 0.0	0.1		
Spotfin shiner	3.4	0.2	7.8	0.0	0.0	0.0		
Steelcolor shiner	1.2	0.0	0.1	0.0	ō.ō	ŏ.ŏ		
Pugnose minnow	0.2	0.0	0.1	0.0	0.0	0.0		
Bluntnose minnow	0.2	2.0	0.4	0.0	0.0	0.0		
Bullhead minnow Northern hogsucker	1.4	0.0	0.4	0.0	0.0	0.0		
#Smallmouth buffalo	0.0	0.0 0.0	0.1 0.6	, T	0,0	T		
Spotted sucker	3.2	4.6	4.0	0.0	T. 0.3	0.4 0.2		
Silver redhorse	ō.ō	o.ŏ	0.0	Ť	ŏ.ŏ	0.1		
Black redhorse	0.0	0.0	0.3	0.1	0.0	Ť		
Golden redhorse	0.8	1.0	1.4	Т	T	0.1		
Blue catfish	0.0	0.0	0.1	0.7	0.1	0.9		
#Yellow bullhead Channel catfish	0.0	0.2	0.0	0.0	0.0	0.0		
Flathead catfish	1.8 1.8	1.8 0.6	2.5	1.2	0.4	1.3		
White bass	0.4	0.0	3.2	1.6	0.1 0.3	0.2 1.8		
Yellow bass	2.0	2.8	9.6	2.5	1.2	2.3		
Striped bass	0.0	ō.ō	0.2	ō.6	0.2	0.4		
#Hybrid striped bass	0.0	0.0	0.0	0.0	T	0.4		
Narmouth Radionanal aum fint	9.2	14.4	2.7	0.0	0.2	<u>т</u> ,		
Redbreast sunfish Green sunfish	30.8 1.0	33.6 1.0	9.1	0.0	0.0	0.0		
Bluegill	259.4	129.2	2.6 132.8	0.0	0.0 0.2	0.0		
Longear sunfish	4.8	4.6	19.8	0.0	Ť	Ť		
Redear sunfish	33.6	58.4	16.9	1.0	0.5	0.8		
#Hybrid sunfish	0.0	0.2	0.1	0.0	0.0	0.0		
Smallmouth bass	2.4	0.4	7.9	_T_	0.0	0.2		
Spotted bass Largemouth bass	25.6	12.0	12.7	0.7	0.3	0.6		
White crappie	52.6 0.8	31.8 1.8	27.8 2.0	0.1 0.1	0.0 0.1	0.3		
Black crappie	7.2	4.8	2.6	0.1	0.1	0.1 0.4		
Yellow perch	7.6	19.4	1.8	0.1	0.1	Ť		
Logperch	1.6	0.8	3.7	0.0	0.0	0.0		
Sauger	0.2	0.0	0.2	0.1	0.1	0.4		
Freshwater drum Brook silverside	5.4	2.6	5.3	0.1	0.4	0.4		
Brook SIIVerside	43.2	16.4	15.4	0.0	0.0	0.0		
Total: CPUE No. collected	1912 9558	462	2112	17.1	9.3	23.6		
Species:		2312 and 2 hyb	87984	512	279	5593		
1990: No. collected	10070		1051					
		s, electr						

* Electrofishing effort units are hours Gill net effort units are net-nights # Species found only in 1989 T Catch per effort less than 0.05

Results - Chickamauga Reservoir

Table 2. Species list and catch per unit effort at inflow, transition, and forebay stations during fall electrofishing and gill netting on Chickamauga Reservoir, 1990. Total efforts(*) shown in parentheses.

			. ·			
	E	lectrofish	ning		ll nettir	
Common			n Forebay		ransition	
Name	(1.7)	(1.7)	(1.7)	(10)	(10)	(10)
		· · · · · · · · · · · · · · · · · · ·				
Chestnut lamprey	0.0	0.6	25.8	0.0	0.0	0.0
Spotted gar	0.0	0.0	1.2	0.0	0.0	0.3
Skipjack herring	0.0	0.0	0.0	3.4	3.0	5.2
Gizzard shad	0.0 94.2	325.8	358.8	1.1	2.6	3.1
Threadfin shad	0.0	1605.0	1335.0	0.0	0.0	0.0
Mooneye	0.0	0.0	0.0	0.1	0.1	0.0
Carp	3.6	27.6	29.4	0.0	0.0	0.2
Golden shiner	2.4	3.6	5.4	0.0	0.5	0.3
Emerald shiner	0.6	165.6	244.8 9.0	0.0	0.0	0.0
Spotfin shiner Steelcolor shiner	0.6	0.6	3.0	0.0 0.0	0.0 0.0	0.0
Steelcolor sniner Pugnose minnow	0.6 0.0	0.6	0.0	0.0	0.0	0.0
Bluntnose minnow	0.0	0.0	0.6	0.0	0.0	0.0
Bullhead minnow	1.8	1.8	0.6	0.0	0.0	0.0
Northern hogsucker	0.0	0.0	0.0	0.1	0.0	0.0
Spotted sucker	7.2		1.2	0.8	0.3	0.3
Silver redhorse	0.0	ō.ō	ō.ō	0.1	0.0	0.0
Black redhorse	7.2 0.0 0.0	0.0	0.0	0.3	0.0	0.0
Golden redhorse	2.4	0.0	0.0	0.1	0.0	0.0
Blue catfish	0.0	0.0	0.0	0.7	1.1	0.4
Channel catfish	1.2	3.0	1.2	1.4	0.8	1.4
Flathead catfish	0.0	0.0	5.4	0.2	0.3	0.1
White bass	1.2	0.0	0.0	0.9	2.2	1.8
Yellow bass	4.2	1.8	0.0	2.1	1.8	3.7
Striped bass	0.0	0.0	0.0	0.2	1.1	0.6
Harmouth	0.6	17.4	9.6	0.0	0.0	0.0
Redbreast sunfish	7.8	29.4	55.2	0.0	0.0 0.0	0.0
Green sunfish	0.6 81.6	1.8 153.6	0.6 543.0	0.3	0.2	0.5
Bluegill Longear sunfish	0.0	5.4	9.0	0.0	0.0	0.0
Redear sunfish	55.8	15.0	30.0	0.5	1.3	1.3
Smallmouth bass		1.2	4.8	0.0	ô.ó	ô.í
Spotted bass	1.2 0.0	9.0	67.8	0.5	0.5	1.0
Largemouth bass	27.0	55.8	75.0	0.0	0.0	ō.3
White crappie	0.0	1.2	1.2	0.0	0.1	0.3
Black crappie	0.0 7.8	3.0	10.8	0.0	0.5	0.9
Yellow perch	0.0	15.6	7.2	0.0	0.2	0.0
Logperch	0.0	0.0	4.8	0.0	0.0	0.0
Sauger	0.6	0.0	0.0	0.1	0.1	0.0
Freshwater drum	8.4	6.6	1.2	0.0	0.1	0.1
Brook silverside	1.2	41,4	87.0	0.0	0.0	0.0
· · · · · · · · · · · · · · · · · · ·						
	712 (2493.6	2928.6		16.9	21 0
Total: CPUE	312.6		2928.6 4881	12.9 129	16.8	21.9 219
No. collected Species: 41	521 (and 0	4156 budbrid	4001	163	168	217
opecies: 41		nyortui				

* Electrofishing effort units are hours; gill net units are net-nights.

Results - Chickamauga Reservoir

Common Name	Inflow	Transition	Forebay
1.00100			
Parasites			×
Chestnut lamprey		- -	$-\frac{2}{1}$
Group total	0	1	-
Planktivores		x	×
Threadfin shad	 0		
Group total	Ū		
Insectivores	×	×	
Mooneye	X	××	X
Emerald shiner Spotfin shiner	X	×	×××
Steelcolor shiner	×	×	~
Puanose minnow	v	~	
Silver redhorse	× × × × × × ×		
Black redhorse	Ŷ		
Golden redhorse	ÿ	X .	X
Warmouth Redbreast sunfish	X	X	Š
Bluegill	×	Š	÷
Longear sunfish	×	× × × × × × × × × ×	****
Redear sunfish	^	Ŷ	×
Yellow perch	x	X	×
Brook silverside	12	11	10
Group total	16		
Specialized benthic insectivores	x	×	×
Bullhead minnow	×××		
Northern hogsucker	X	X	X
Spotted sucker Logperch			÷
Freshwater drum	×	×	× × ×
Group total	4	3	•
Omnivores	J	v	×
Gizzard shad	X	×	÷ ×
Carp	X X	X	×
Golden shiner			X
Bluntnose minnow Blue catfish	×××	X	Š
Channel catfish			*****
Group total	5	5	6
Piscivores			v
Spotted gar		y y	*****
Skipjack herring	S	i î	×
Flathead catfish	5	K X	×
White bass	ý	(<u>X</u>	×
Yellow bass Striped bass	>	ć X	. <u>X</u>
Green sunfish	>	s s	- Ç
Smallmouth bass	× >> >> >>	××××××××××××××××××××××××××××××××××××××	Ŷ
Spotted bass		2	X
Largemouth bass	•	· ×	×
White crappie Black crappie		K X	×
Black crappie Sauger			
Jan Jan			12
Group total	1	1 12	12
	- 3	2 33	34

Table 3. Species list according to trophic designation of fish encountered during fall electrofishing and gill netting at Chickamauga Reservoir, 1990.

Results - Chickamauga Reservoir



Table 4. Relative weight (Wr) analysis of principal species at Chickamauga Reservoir, 1990, compared to standard weights established for Mainstream Reservoirs.

.....

	Inflow		Transition		Forebay		Overall		1989	
Common Name	Mean Wr	N	Mean Wr	N	Mean Wr	N	Mean Wr	N	Mean Wr	N
Channel catfish	-	-	110	5	108	2	109	7	-	-
Smallmouth bass	99	2	92	2	98	9	97	13	97	2
Spotted bass	93	5	90	18	87	120	88	143	86	45
Largemouth bass	. 94	42	9 8	90	101	123	99	255	97	159

Table 5. Proportional (PSD) and relative stock density (RSD) of principal species at Chickamauga Reservoir, 1990 . Available 1989 values included.

Common Name	N	Quality (PSD)	Preferred (RSD)	Memorable (RSD2)	Trophy (RSD3)
			(RSU)	(RSU2)	(KSUS)
Channel catfish					0
Inflow	15	53	0 20	0 0	ŏ
Transition	10	80 64	20	ŏ	ŏ
Forebay	11 36	64	8	Ŏ	0 0
Overal1-1990 1989	21	67	10	ŏ	0
1989	21	•			
Bluegill Inflow	27	26	0	0	0
Transition	42	24	2	2	0
Forebay	36	22	0 1	2 0 1	0
Overal1-1990	105	24	1	1	0
1989	400	15	U	U	Ū
Redear sunfish			,	0	0
Inflow	51	20 81	4 19	र	ŏ
Transition	32	42	17	3 0 1	0
Forebay Overall-1990	38 121	43	5	ĩ	0
1989	128	46	6	ō	ŏ
1,0,					
Spotted bass Inflow	5	100	60	0	0
Transition	14	21	14	0	0
Forebay	21	29	5 15	0	0
Overal1-1990	40	35	15	0	0
1989	45	36	4	0	U
Largemouth bass				,	•
Inflow	25	52	12	4	0
Transition	44	39	20	í	0
Forebay	79	49	.8 12	· 1	0 0 0
Overal1-1990 1989	148 128	47 45	12	7 1 3 2	ŏ

Results - Chickamauga Reservoir

Fish health assessment index (FHAI) results for Chickamauga Reservoir, 1990, compared to 1989. Table 6.

·	Inflo 1990 1		Transi 1990		Fóreb 1990 1	
Health Assessment Index (FHAI)	28.6 5	0.6	69.3	54.0	66.0 5	56.0
Standard Deviation	17.8 3	3.8	39.2	30.4	25.7 3	\$4.4
Coefficient of Variation	62.2 6	6.6	56.6	56.3	39.0 6	51.5
Sample Size	15	30	15	30	15	30
Number of Anomalies in:						
Eyes	0	0	0	0	0	0
Liver	2	11	6	11	5	12
Spleen	3	12	7	5	3	7
Kidney	0	11	6	10	4	9
Skin	1	0	1	0	2	0
Gills	Ċ	0	1	0	1	0
Pseudobranchs	0	0	0	0	0	. 0
Thymus	0	1	0	0	0	0
Hind Gut	0	0	0	0	0	0
Fins	0	0	0	0	0	0
Opercles	0	1	0		0	0
Parasites	14	0	15		12	0
Hematocrit	0	13	3		1	13 3
Leucocrit	1	0	2		2 ·	8
Plasma Protein	0	2	1	5	11	0
Mean FHAI for Chickamauga Reservo	ir: 199 198		54.6 53.5			
Mean FHAI for mainstream reservoi	rs: 199 198		48.1 45.8			
Mean FHAI for all sites sampled in each area category (i.e., inflow, transition, and forebay) for mainstream reservoirs: 1985		59.9 47.3		52.8 49.0		51.3 42.9
Mean FHAI for all sites sampled in each area category (i.e., inflow, transition, and forebay) for all reservoirs sampled: 1984	0	41.5 45.4	-	47.8 42.3		50.0 39.3
Overall FHAI for all sites sample in the Tennessee Valley: 199 198	0 47.1					

•

	nickamaug 1990 vs. ainstream Ave.	a Chickamauga 1990 vs. Chickamauga 1989	vs. Mair	ns within Ch average val nstream Rese Transition	ues at rvoirs	Reservoir
Species richness						
Total species	0	.+	0	+	+	
Sunfish species		0	-	•	+	
Sucker species Catfish species		+	-	ō	ō	
Small cyprinid		+	+	+	+	
Fish health						
FHAI	-		+	-	-	
Trophic compositio (pct)	on					
Omnivores	+		+	-	+	
Insectivores	+		0	+	+	
Planktivores	+		+	-	•	
Piscivores Specialists	-	•	-	* *	-	
Sensitive species				25		
Tolerant (pct)		<u>,</u>	+	-	•	
Intolerant spp Electrofishing ca rate (no./hr)		•	-	•	•	
Total catch ra	+o -		-	•	•	
Largemouth bas		+	+	+ .	•	
Smallmouth bas		+	-	-	-	
Spotted bass	+	+	-	+	+	
Bluegill	•	+	+	-	+	
Redear sunfish Gizzard shad	+	-	+	-	+	
Gill netting catc rate (no./net-nig						
Total catch ra	te -	+	-	-	-	
Channel catfis		+	+	-	-	
Morone spp.	0	+	-	+	+	
Stizostedion s	pp	0	-	-	-	
Relative weight						
Largemouth bas		0	-	0	-	
Smallmouth bas Spotted bass	s _	0		0	-	
Proportional stoc	L					
densitý	n .					
Largemouth bas		0	-	-	-	
Smallmouth bas						
Spotted bass	-	0		-	-	
Bluegill Redear sunfish	-	-	-	-	-	
Channel catfis	h 0	0	-	•	+	
Overall fish abundance						
Grand total fi	sh -	*	•	-	.+	
	-8	10	-4	-1	3	
	-		Mana	-	-	
	Horse	Better	Worse	Average	Better	

Table 7. Summary comparison of 34 selected measurements of the fish community sampled at Chickamauga Reservoir, 1990.*

Plus signs indicate a healthier condition. Minus signs indicate a less healthy condition. Zeroes indicate differences less than + or - 5 pct. Blanks indicate insufficient data for comparison.
 ** Small cyprinids also include darters, topminnows, and brook silversides.

Results - Chickamauga Reservoir



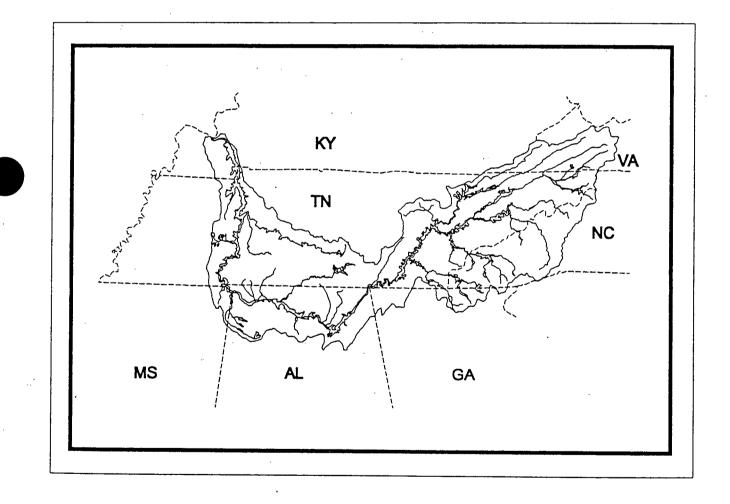


Tennessee Valley Authority

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Water Resources Division Chattanooga, Tennessee TVA/WR-92/5 July 1992

RESERVOIR VITAL SIGNS MONITORING - 1991 FISH COMMUNITY RESULTS



WATER RESOURCES & ECOLOGICAL MONITORING

Introduction

The Tennessee Valley Authority (TVA) operates 9 reservoirs on the Tennessee River and 37 reservoirs on its tributaries. TVA is committed to maintaining the health of aquatic resources created when the reservoir system was built. To that end, TVA conducts the Water Resources and Biological Monitoring Program that includes physical, chemical, and biological data collection components. Biological monitoring targets the following selected elements within three zones of the reservoir (inflow, transition, and forebay):

- Sediment/Water-column Acute Toxicity Screening (forebay and transition zone only)
- Benthic macroinvertebrates
- Fish

Reservoir fish monitoring is divided into the following activities:

- Fish Biomass
- Fish Tissue Contamination
- Fish Community Monitoring
- Fish Health Assessment

This report presents the results of fall 1991 fish community monitoring and fish health assessment data using a new analytical approach: Reservoir Index of Biotic Integrity (RIBI). Fish health assessment is included in this report as one of the RIBI metrics. Reports on other components and activities are published in companion reports, i.e., fish biomass (Wilson 1992) and fish tissue (Bates et al. 1992), and a summary report on the results of the monitoring program also is available (Dycus and Meinert 1992).

Philosophical Approach

The basic ecological principle underlying the community monitoring program is that characteristics of fish populations, because of their trophic status, will reflect changes in the aquatic ecosystem. The program's objective is to provide the minimum information necessary to evaluate the status of the fish community at inflow, transition zone, and forebay areas of reservoirs throughout the Tennessee Valley.

Introduction

The quality of an aquatic resource, in this case fish, is indicative of the quality of its physical and chemical environment. As relatively long-lived biological indicators, fish integrate conditions of the aquatic environment over long periods of time (i.e., seasons and years). Aquatic environments that produce healthy fish communities are expected to also provide favorable conditions for other aquatic organisms, and ultimately, man.

The use of an index to evaluate reservoir fish communities is based on methods developed for stream fish surveys (Karr et al. 1986). The author of the original stream IBI, Dr. James R. Karr, and research associates, Drs. Michelle Dionne and Martin Jennings, are co-developing the reservoir index of biotic integrity (RIBI) under contract with TVA.

Like the stream IBI, the RIBI evaluates fish communities based on a series of measurements, or metrics, derived from fish samples. Each metric describes a facet of fish community functioning or structure. Comparison of metric performance over time or between areas can be useful in detecting biological responses to one or more sources of degradation in the aquatic ecosystem. The categories of RIBI metrics include:

- Species richness and composition
- Trophic composition
- Reproductive composition
- Abundance and fish health

Healthy aquatic ecosystems are recognized by the presence of diverse fish communities. Physical, chemical, or bacteriological degradation will have negative effects on species diversity and/or abundance. Species richness metrics address total species observed plus key groups of species, such as sunfish species, sucker species, and species designated as being particularly sensitive (intolerant) of habitat degradation. Just as intolerant species indicate good community health, high proportions of tolerant individuals signify degraded health of the fish community.

Trophic composition metrics describe the proportions of omnivores and invertivores. Piscivores are not included in the present analysis, but are being examined for future use. Omnivores as a group are less sensitive to environmental stresses due to their ability to vary their diet.

Spawning requirements of some reservoir species make them more vulnerable than others. Migratory spawners shed their eggs in flowing headwater areas of reservoirs or tributary streams, where egg survival is subject to prevailing stream conditions (temperature, siltation, water level fluctuation, sediment quality, dissolved oxygen, and chemical water quality). Lithophilic broadcast spawners, many of which are also migratory, shed eggs that are subject to the availability of suitable rock substrates, i.e., conducive to egg survival. Species of both groups shed their eggs over a relatively short spawning season (2-3 weeks), emphasizing the importance of favorable conditions for successful reproduction to the continued existence of the species.

Introduction .

Other metrics address fish abundance and health. High quality communities support large numbers of individuals (excluding shad). Fish health assessment measures environmental stress on a top predator (largemouth bass) based on rigorous external and internal examinations. Another measure of fish community health under investigation for use in future RIBI analysis is incidence of diseases, lesions, tumors, external parasites, deformities, and blindness among all species sampled.

Each metric is compared to a set of reference conditions, or scoring criteria, and rated 1, 3, or 5 with values of 1 indicating "poor" conditions, and values of 5 indicating "good" conditions. Because "natural" reservoirs do not exist, reference conditions were empirically derived from previously collected fall fish community data. Scoring criteria are designed for each reservoir zone, and ideally for each type of reservoir, although the present criteria are more applicable to mainstream reservoirs than tributary reservoirs.

The scores of the 11 metrics are summed to create the RIBI, one number that describes the overall condition of the fish community of a given reservoir zone. The RIBI can then be used to compare between zones of various reservoirs and over time, and becomes a tool for monitoring the quality of fish communities.

Introduction

Methods

Fish Community Monitoring

Twenty-two reservoirs were studied during fall 1991 (table 1). Shoreline electrofishing samples were collected during daylight hours from inflow, transition, and forebay zones of each reservoir from middle September to Decem-The forebay area was defined as the main channel shoreline ber, 1991. extending into the "mouths" of tributary streams within approximately three miles above the dam. The transition zone was the main reservoir reach where hydraulic and water chemistry conditions and sediment particle size suggest a shift from a riverine to a reservoir environment. In mainstream reservoirs the inflow zone was the tailwater of the upstream dam, while in tributary reservoirs the inflow zone was near the mouth of the free-flowing headwater stream(s). The September-December time frame was chosen because an extended period of time is required to sample all reservoirs, and distribution of fish populations is most stable in fall. A total of 10 electrofishing runs (10 minutes duration each) was sampled within each of the 56 sites (22 reservoirs). All habitats were sampled at each site, with dominant types receiving the most effort. Habitat distinctions were based on major changes in substrate (e.g., rock, rip-rap, or clay), and/or cover (e.g., brush or aquatic vegetation). Sample size/duration was selected based on sensitivity analysis of largemouth bass electrofishing samples in two Oklahoma reservoirs (Gilliland 1985). A range finder was used to measure shoreline distance covered during each 10 minute electrofishing run during the 1991 season.

Black bass species captured were measured and weighed, other major sport fish species and channel catfish were measured, and all other species were enumerated prior to release. All fish captured were examined for obvious external abnormalities, diseases, and parasites, and this information was noted. Fish observed but not captured were included in the records if positive identification and enumeration could be made. Estimated counts were made in instances where high densities of fish were encountered (usually shad), and were recorded in the remarks portion of the field form. Young-of-year (YOY) fish of a given species were counted separately from adults and also recorded in the remarks section, except for YOY gizzard and threadfin shad. Shad YOY were included on the field form in the usual manner, but were assigned special codes to identify them as YOY. Designations of YOY fish were at the discretion of the crew leader according to the following general guidelines: sunfish <25mm, shad < 100 mm, and black bass < 100 mm. Hybrid individuals were listed on the field form as separate species. Fifteen largemouth bass from the electrofishing samples in each reservoir zone were selected for fish health assessment.

Methods

Voucher specimens of each small minnow or darter species (other than brook silversides and logperch) collected were preserved in 10 percent formalin and transported to the Norris Aquatic Biology Laboratory for verification of species identity. Any other individuals of questionable identity or hybrids were also taken to the Norris laboratory for positive identification.

Where conditions permitted, ten 100-ft-monofilament experimental gill nets with five 20-ft panels (mesh sizes of 1, 1 1/2, 2, 2 1/2, and 3 inch bar mesh) were set for one overnight period in each reservoir zone. In forebay and transition zones, nets were set in all habitat types, alternating mesh sizes toward the shoreline between sets. In inflow areas, nets were set wherever flow conditions allowed. Below dams nets were set in areas protected from river currents, such as the spilling basin, off lock and wing walls, and in pockets and side channels. Availability of adequate sample locations limited number of nets set to less than ten at some inflow stations.

Lengths and weights of all black bass species and channel catfish were recorded, as were lengths of other sport species. Counts of the remaining species were made prior to releasing. Any incidences of diseases, parasites, or anomalies were recorded in the remarks column of the field form.

Species lists and catch per effort data for both sampling gears are presented for each reservoir sampling zone in the Appendix.

Reservoir	Infl Location		Transition Location Date	Forebay Location Date
Mainstream				
Kentucky Pickwick Wilson Wheeler Guntersville Nickajack Chickamauga Watts Bar "" Fort Loudoun Tellico Melton Hill	TRM 652.	0 10/29 2 10/03 2 10/10 0 11/06 0 10/24 0 10/31 0 11/19 0 11/20 0 12/10	TRM 117.0 11/07 TRM 230.7 10/30 TRM 313.9 10/09 TRM 392.5 09/24 TRM 431.0 11/05 TRM 490.0 10/22 TRM 560.0 10/29 TRM 560.0 10/29 TRM 624.0 12/09 LTRM 20.6 12/04	TRM 22.4 11/05 TRM 209.1 10/31 TRM 260.6 10/02 TRM 278.2 10/08 TRM 351.0 09/11 TRM 425.0 11/07 TRM 472.0 10/23 TRM 531.0 10/30 TRM 607.8 12/05
Tributary	CRM 59.	0 12/18	CRM 44.5 12/17	CRH 25.0 12/12
Norris	-	-	CRM 124.0 10/08	CRM 80.5 10/09
Cherokee Boone " South Holston Watauga Douglas Hiwassee Chatuge	- WRM 44.0 FBRM 64.5	11/07 10/01 10/29 10/10	PRM 30.1 11/12 HRM 75.5 11/06 SHRM 27.0 10/16 WRM 8.3 10/17 SHRM 62.0 10/22 FBRM 56.9 10/30 HIRM 85.0 10/09	HRM 53.0 11/05 SHRM 19.8 10/15 SHRM 52.0 10/23 WRM 37.4 10/02 FBRM 33.0 10/31 HiRM 77.0 10/08 HiRM 122 10/01
Nottely Parksville (Ocoee #1) Blue Ridge	-	-		HIRM 122 10/01 NoRM 23.5 10/02 ORM 12.5 10/17 ToRM 54.1 10/03

Table 1. Fish community sampling locations and dates at selected reservoirs for the TVA Reservoir Monitoring Program, fall 1991 (FY 1992).

Fish Health Assessment

Fifteen largemouth bass (> 250 mm total length) were collected during electrofishing surveys at the reservoir monitoring zones and transported to a mobile laboratory for examination (Goede 1991). After the fish were anesthetized with tricaine methanesulfonate (MS-222) in 50 mg/l (ppm) solution, the body cavity of each fish was opened using sharp/blunt-ended surgical scissors by making a ventral incision from the vent forward to the pectoral girdle, cutting closely to one side of the pelvic girdle. Care was taken not to make the incision deep enough to damage internal organs. Blood was then collected by cardiac puncture using a hematocrit tube which was centrifuged to allow measurement of hematocrit, leucocrit, and plasma protein levels. The liver was examined immediately for anomalies because of the tendency for rapid discoloration following death of the individual.

An external examination was made on the eyes, skin, gills, fins, opercles, pseudobranchs, and thymus. Internally, the mesenteric fat, spleen, hindgut, kidney, bile, parasites, and sex were examined. Abnormal conditions and other data were entered directly into a computer program developed by TVA entitled "AUSUM430" which calculated a fish health assessment index (FHAI) for each reservoir zone. Abnormalities were weighted according to severity such that as debilitating anomalies increased, the resulting FHAI also increased. Thus better fish health was indicated by lower FHAI values. Results were used as one metric in the RIBI.

Methods

Reservoir Index of Biotic Integrity (RIBI)

Shoreline electrofishing data collected in the manner described above were input to the preliminary RIBI analysis program developed by Drs. Karr and Jennings at the University of Washington in Seattle (Jennings and Karr 1992). The preliminary analysis was based on eleven metrics (table 2). All species present, native or introduced, were considered in the total species counts, except hybrids and species that only were present as YOY. Gizzard shad, threadfin shad, and YOY counts were not included in the total fish abundance metric or any of the proportional metrics. For the sunfish species metric only species of the genus *Lepomis* were considered. At some locations electrofishing samples did not collect sufficient numbers of largemouth bass to perform fish health assessment. In these cases scores of "3" were arbitrarily assigned.

Scoring criteria for the eleven metrics were developed from ranges in fall electrofishing data collected from TVA mainstream reservoirs in 1989 and 1990. The observed data ranges were trisected (Fausch et al. 1984), such that values falling in the upper third were designated "good", and values falling in the middle and lower thirds were designated "fair" and "poor", respectively. This manner of reference value determination is unlike that of stream IBI methods, in which scoring criteria are based on pristine, undisturbed environments, because there are, by definition, no naturally occurring, undisturbed reservoirs upon which to draw comparisons. Scoring criteria used in this analysis, drawn from mainstream reservoir data, are less applicable to tributary reservoirs, but the RIBI's calculated for tributary reservoirs are valuable for comparison of fish communities within that group. Relative lack of fall electrofishing data from TVA tributary reservoirs precluded development of reasonable scoring criteria, however as more fall data become available from monitoring tributary reservoirs, this deficiency in RIBI analysis will be removed.

Results of 1991 sampling were compared to the scoring criteria, and each of the eleven metrics were rated "5" if the observed value fell within the "good" range, or "3" or "1" if the observed value fell in the "fair" or "poor" range, respectively.

Fish species common to the lower mainstream reservoirs of the Tennessee River were classified according to trophic guild, relative tolerances of environmental degradation, and reproductive guild (table 3) for use in RIBI analysis to calculate the values of the eleven metrics. These classifications are subject to future modification as our knowledge of reservoir fish community relationships increases.

The sum of the eleven metric ratings is the RIBI, the index value that summarizes the overall condition of a given reservoir fish community. The RIBI values of fall fish community surveys, 1989-1991 (Jennings and Karr 1992) were also trisected to designate "good", "fair", and "poor" fish communities at inflow, transition, and forebay zones of TVA reservoirs. Mainstream and tributary reservoirs were considered jointly because they were analyzed according to the same scoring criteria. However, future analyses will be based on at least two reservoir groupings.

7

Methods

The nine Tennessee River reservoirs plus Melton Hill and Tellico were designated mainstream reservoirs. The latter two were included because they are also "run-of-the-river" reservoirs, having in common navigation and relatively minor winter drawdown zones. The remaining tributary reservoirs have no navigation locks and experience major winter drawdowns.

Table 2. Preliminary RIBI metrics and scoring criteria developed for TVA mainstream reservoirs. Scoring reflects relative fish community quality, with a score of 5 representing highest quality, and a score of 1 the poorest (from Jennings and Karr 1992).

Metric	5	Inflow 3	1		Transiti	on		Foreba	
			1	5	3	1	5	3	, 1
Species richness and composition					、 、				
 Total species Sunfish species Sucker species Intolerant species Percent of individuals as tolerant species 	>27 >4 >5 >3 <7.5	20-27 3-4 3-5 2-3 7.5-15	<20 <3 <3 <2 >15	>26 >4 >3 >3 >3	21-26 3-4 2-3 2-3 7.5-15	<21 <3 <2 <2 >15	>23 >4 >2 >3 < 7.5	19-23 3-4 2 2-3	<19 <3 <2 <2
Trophic composition						~15	< 7.5	7.5-15	>15
 6. Percent of individuals as omnivores 7. Percent of individuals 	<2.5	- 2.5-5	>5	<5	5-10	>10	<5	-	
as invertivores eproductive composition	>70	55-70	<55	>80	70-80	<70	>80	5-10 70-80	>10 <70
 8. Migratory spawning species 9. Lithophilic spawning species 	>3 >6	2-3	<2	>2	1-2	0	>2	1-2	0
bundance and fish health	20	4-6	<4	>4	2-4	<2	>4	2-4	<2
0. Total number of	>600 <45	300-600 45-70	<300 >70	>800	400-800	<400	>600	300-600	<300
-			-/0	<45	45-70	>70	<45	45-70	>70

Methods

Table 3. Core fish species list with trophic, tolerance, and rep designations(*) for use in preliminary electrofishing f Index of Biotic Integrity (RIBI) for TVA reservoirs, 19	
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Species	Trophic Guild	Tolerance	Migratory Spawner	Lithophilic Spawner	
Chestnut lamprey	PS		м		
Spotted gar	PI		••		
Longnose gar Shortnose gar	PI	TOL			
Bowfin	PI PI	TOL			
American eel	PI				
Skipjack herring	ΡÎ	INT	н		
Gizzard shad	OM	TOL	11		
Threadfin shad Mooneye	PL	_			
Chain pickerel	IN		H -	L	
Central stoneroller	PI HB				
Goldfish	OM	TOL			
Common carp	OM -	TOL			
Silver chub	SP	INT			
Golden shiner Emerald shiner	OH	TOL		•	
Ghost shiner	IN				
Spotfin shiner	IN IN	TO			
Mimic shiner	· IN	TOL			
Steelcolor shiner	ÎN				
Pugnose minnow	IN				
Bluntnose minnow Fathead minnow	OH				
Bullhead minnow	OH				
River carpsucker	IN Om				
Quillback	OH		М		
Northern hog sucker	SP	INT	- M		
Smallmouth buffalo	OH		- H	L	
Bigmouth buffalo Black buffalo	PL		H		
Spotted sucker	OH		H		
Silver redhorse	IN In	INT	H	L	
Shorthead redhorse	IN		H	L	
River redhorse	ŠP	INT	M H	L	
Black redhorse	IN	INT	M	L ·	•
Golden redhorse Blue catfish	IN ·		Й		
Black bullhead	OH			-	
Yellow bullhead	OH. OH	TOL			
Brown bullhead	OH	TOL TOL			
Channel catfish	OM	IUL			
Flathead catfish	PI				
Blackstripe topminnow	IN				
Blackspotted topminnow Mosquitofish	IN				
Brook silverside	IN In	TOL			
White bass	PI			•	
fellow bass	PÎ		H	Ļ	
Rock bass	PI	INT	п	L	
Redbreast sunfish	IN	TOL			
Green sunfish Marmouth	IN	TOL			
rangespotted sunfish	IN				
Sluegill	IN In				
ongear sunfish	ÎN	INT		•	
Redear sunfish	IN	2011			
potted sunfish mallmouth bass	IN				
potted bass	PI				
argemouth bass	PI PI				
hite crappie	PI				
lack crappie	PI				
ellow perch	IN				
ogperch	SP			,	
auger alleye	PI PI		н	L	

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* Designations: trophic- herbivore (HB), parasitic (PS), planktivore (PL), omnivore (OH), insectivore (IN), piscivore (PI), specialized benthic insecti-vore (SP) tolerance- tolerant (TOL), intolerant (INT) migratory spawning species (H) lithophilic spawning species (L)

Table 7a. Preliminary scoring of electrofishing results for the eleven metrics and overall reservoir index of biotic integrity (RIBI) for Chickamauga Reservoir, 1991.

Metric	Inflow Obs. Score		Transition Obs. Score		Forebay Obs. Score	
A. Species richness and composition				_		
1. Total species 2. Sunfish species 3. Sucker species 4. Intolerant species	25 5 1 1	3 5 1 1	21 6 1 2	. 3 5 1 3	21 6 0 1	3 5 1 1
5. Percent of individuals as tolerant species	12	3	7	5	1	5
8. Trophic composition						
6. Percent of individuals as omnivores	6	1	5	5	1	5
 Percent of individuals as invertivores 	72	5	90	5	94	5
C. Reproductive composition						
8. Migratory spawning species	4	5	2	3 ·	1	3
9. Lithophilic spawning species	5	3	3	3	2	1
D. Abundance and fish health						
10. Total number of individuals	696	. 5	885	5	1744	5
11. Fish health assessment index (FHAI)	79	1	75	1	83	1
RIBI		,33 fair		39 good		35 900d

Appendix

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Table	7Ь.
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Species list and catch per unit effort at inflow, transition, and forebay stations during fall electrofishing and gill netting on Chickamauga Reservoir, 1991. Total efforts(*) shown in parentheses.

		E1	ectrofish	ing	Gill netting			
Common	Designation+			n Forebay		Transitio		
Name	-	(1.7)	(1.7)	(1.7)	(4)	(10)	(10)	
Longnose gar	PI,TOL	0.0	0.0	0.0	9.0	0.0	0.0	
Skipjack herring	PI,INT,M	0.0	0.0	0.0	0.8	4.6	6.5	
Gizzard shad	OM, TOL	190.2	127.8	34.2	1.0	3.4	2.8	
Threadfin shad	PL	1.8	1.2	0.0	0.0	0.0	0.0	
ov threadfin shad	• =	60.0	0.0	184.8	0.0	0.0	0.0	
Carp	ON.TOL	3.6	10.8	8.4	0.3	0.1	0.2	
Solden shiner	OM, TOL	0.0	15.0	0.6	0.0	0.0	0.1	
Emerald shiner	IN	10.8	263.4	676.8	0.0	0.0	0.0	
Spotfin shiner	ÎN,TOL	43.8	0.0	0.0	0.0	0.0	0.0	
Bluntnose minnow	OM	1.8	0.0	0.0	0.0	0.0	0.0	
Quillback carpsucker		0.0	0.0	0.0	1.0	0.0	0.0	
Smallmouth buffalo	OM,M	0.0	0.0	0.0	0.3	0.0	0.0	
Spotted sucker	IN, INT, M.L	4.2	0.6	0.0	0.0	0.0	0.0	
Blue catfish	· OM	0.6	0.0	0.0	2.8	0.5		
Channel catfish	OM	20.4	0.6				1.0	
lathead catfish	PI	7.8	1.2	0.0	5.8	0.5	1.2	
hite bass				2.4	0.5	0.1	0.2	
	PI,M,L	7.2	0.0	0.6	0.3	1.3	0.3	
ellow bass	PI,M,L	38.4	1.8	0.0	2.3	0.4	0.4	
Striped bass		0.0	0.0	0.0	0.0	0.2	. 0.1	
lybrid striped bass		0.0	0.0	0.0	0.0	0.3	0.2	
armouth	IN	2.4	7.2	1.8	0.0	0.0	. 0.0	
Redbreast sunfish	IN, TOL	4.2	11.4	3.6	0.0	0.0	0.0	
Freen sunfish	PI, TOL	4.2	1.8	0.6	0.0	0.0	8.0	
Bluegill	IN	159.6	142.2	252.0	0.3	0.0	0.0	
ongear sunfish	IN, INT	0.0	2.4	2.4	0.0	0.0	0.0	
ledear sunfish	IN	44.4	24.6	24.0	0.0	1.2	1.2	
lybrid sunfish		1.2	0.0	0.0	0.0	0.0	. 0.0	
mallmouth bass	PI	9.0	3.6	10.8	0.0	0.0	0.0	
potted bass	PI	15.6	5.4	22.8	0.3	1.6	1.1	
argemouth bass	PI	13.8	14.4	15.6	0.0	0.2	0.2	
hite crappie	PI	0.0	0.0	3.0	0.0	0.3	6.0	
lack crappie	PI	1.8	0.0	0.6	0.3	0.1	0.1	
ellow perch	IN	9.6		1.2	0.0	0.1	0.1	
ogperch	IN,L	1.8	18.6	7.2	0.0	0.0	0.0	
Sauger	PI,H,L	0.6	. 0.0	0.0	0.0	0.1	0.0	
alleye	PI,N,L	0.0	0.0	0.0	0.5	0.1	0.0	
reshwater drum	IN	0.0	1.8	0.0	1.0	0.4	0.8	
Brook silverside	IN	10.8	0.0	12.0	0.0	0.0	0.0	
								
Total: CPUE	•	669.6	660.0	1265.4	26.0	15.8	16.6	
No. collect	ted	1116	1100	2109	104	158	166	
Species:	35 (and 2 hyt			• •				

* Electrofishing effort units are hours; gill net units are net-nights. + Designations: trophic- herbivore (HB), parasitic (PS), planktivore (PL), omnivore (ON), invertivore (IN), specialist benthic invertivore (SP) tolerance- tolerant (TOL), intolerant (INT) migratory spawning species (M), lithophilic broadcast spawning species (L)

RESERVOIR VITAL SIGNS MONITORING - 1992 FISH COMMUNITY RESULTS TABLES

INTRODUCTION

Fish communities of 12 TVA reservoirs have been monitored under the "Vital Signs" program since fall 1989. An additional 11 reservoirs were added to the program in 1991. Reservoir fish monitoring is divided into fish biomass, tissue contamination, community monitoring, and health assessment. These tables represent results of fall 1992 fish community monitoring and fish health assessment.

METHODS

Fish Community Monitoring

Twenty-three reservoirs were studied during fall 1992. Shoreline electrofishing samples were collected during daylight hours from inflow, transition, and forebay zones of each reservoir (one zone may have been omitted from specific reservoirs for various reasons) from mid-September to December, 1992. A total of 10 electrofishing transects 1000 feet in length were collected from each of the reservoir zones. All habitats were sampled in proportion to their occurrence in the zone.

Black bass species captured were measured and weighed, other major sport species and channel catfish were measured, and all other species were enumerated prior to release. Fish observed but not captured were included in the records if positive identification and enumeration could be made. Estimated counts were made in instances where high densities of fish were encountered, and were recorded. Young-of-year (YOY) fish were counted separately from adults. Fifteen largemouth bass from the electrofishing samples in each reservoir zone were selected for fish health assessment.

Where conditions permitted, 12 100-ft-monofilament experimental gill nets with five 20-ft panels (mesh sizes of 1, 1 1/2, 2, 2 1/2, and 3 inch bar mesh) were set for one overnight period in each reservoir zone. In forebay and transition zones, nets were set in all habitat types, alternating mesh sizes toward the shoreline between sets. In inflow areas, nets were set wherever flow conditions allowed. Availability of adequate sample location limited number of nets set at some inflow stations.

Fish Health Assessment

Fifteen largemouth bass greater than 250 mm total length were collected during electrofishing surveys at the reservoir monitoring zones and transported to a mobile laboratory for examination. An external and internal examination to observe anomalies was conducted and data entered directly into a computer program developed by TVA entitled "AUSUM 430" which calculated a fish health assessment index (FHAI) for each reservoir zone. Results were used as one metric in the Reservoir Fish Assemblage Index (RFAI).

Reservoir Fish Community Index (RFAI)

Shoreline electrofishing data collected in fall 1992 were input in to a preliminary (RFAI) developed by TVA and Drs. Karr and Jennings at the University of Washington in Seattle. The RFAI employs 11 fish community metrics:

- (A) species richness and composition total number of species, sunfish species, sucker species, intolerant species, and percentage of tolerant individuals sampled;
- (B) trophic composition percentage of invertivorous individuals, and percentage of omnivorous individuals;
- (C) reproductive composition numbers of migratory spawning species, and numbers of lithophilic spawning species;
- (D) Overall fish abundance; and
- (C) fish health assessment index of largemouth bass.

Scoring criteria for the eleven metrics were developed from ranges in fall electrofishing data collected from TVA reservoirs in 1989 through 1991. The observed data ranges were trisected, such that values falling in the upper third were designated "good", and values falling in the middle and lower thirds were designated "fair" and "poor", respectively. Independent scoring criteria were developed for each area (inflow, transition, and forebay) of both run-of-the-river and tributary reservoirs. Results of 1992 sampling were compared to the scoring criteria, and each of the eleven metrics were rated "5" if the observed value fell within the "good" range, or "3" or "1" if the observed value fell in "fair" or "poor" range, respectively. The sum of the eleven metric ratings constituted the RFAI index value which summarizes the overall condition of a given reservoir fish community. Attainable RFAI values were divided into five categories (excellent, good, fair, poor, and very poor) to generally discribe the environmental condition of that section of a reservoir based on attributes of the resident littoral zone fish community. Table 5. Species list and catch per unit effort at the inflow, transition, and forebay during fall
electrofishing and gill netting on Chickamauga Reservoir, 1992. (Electrofishing
effort = 300 meters of shoreline and gill netting effort = net-nights.)

		Electrofishing		1	Gill Netting	
Common name	Inflow	Transition	Forebay	Inflow	Transition	Forebay
Longnose gar	•			4.5	•	0.2
Skipjack herring	•		•	2.5	4.0	2.5
Gizzard shad	17.6	0.8	3.7	6.3	5.4	5.9
Threadfin shad	2620.0	100.6	20.0		•	
Hybrid shad	•	•	•	0.3	0.1	0.1
Mooneye	•	•		0.3	0.2	0.1
Carp	0.1	2.8	0.1		0.1	0.6
Golden shiner	•	•	•		0.4	0.9
Emerald shiner	•	. •	43.3			•
Spotfin shiner	•	•	0.3		•	•
Northern hog sucker	•		•			0.1
Smallmouth buffalo	•	•	•	0.3		
Spotted sucker	0.3	•	•	.	1.0	0.1
Shorthead redhorse	•		•	· ·	0.1	•
Black redhorse	0.1	- ·	•	.		•
Blue catfish	•	•	•	0.5	0.2	0.4
Channel catfish	0.3	•		6.8	0.6	0.7
Flathead catfish	•	•	0.3	0.8	0.2	0.1
White bass	0.1		0.1	1.3	1.9	0.2
Yellow bass	1.4	•		9.5	3.9	1.9
Striped bass	0.4			0.8	0.1	
Hybrid striped x white bass	•			0.3		•
Warmouth	0.5	0.2	•		0.1	0.1
Redbreast sunfish	0.1	0.4	0.9		•	
Green sunfish	0.4	•	0.1			•
Bluegill	9.0	2.0	29.8		0.1	0.1
Longear sunfish	0.2	•	•		•	
Redear sunfish	2.8	0.9	1.4	0.8	1.8	0.6
Smallmouth bass	0.3	•	8.6	0.3	0.2	
Spotted bass	0.4	0.5	3.6	1.3	2.5	2.5
Largemouth bass	1.9	1.3	0.7	0.8	0.4	0.7
White crappie	•	•	0.2			•
Black crappie	•			0.5	0.4	1.1
Yellow perch	•	0.9	0.2	· ·	0.1	•
Logperch	•	•	0.3	· ·	•	•
Sauger	•	•	•	2.5	0.1	•
Walleye				1.3	0.1	
Freshwater drum	0.1	0.4	0.1	0.8	0.7	0.1
Brook silverside	•	4.4	7.1	· · ·	•	· ·
Total: CPUE	2656.0	115 0	100.0			
	2656.0	115.2	120.8	41.8	24.7	19.0
Number of samples	10	10	10	4	10	10
Number collected	26560	1152	1208	167.2	247	190
Species: 37				1		

 Table 5b.
 Scoring of electrofishing results for the eleven metrics and overall Reservoir Fish Association Index (RFAI) for Chickamauga Reservoir.

	Infl	ow	Trans	sition	Fore	bay
Metric	Obs.	Score	Obs.	Score	Obs.	Scor
A. Species richness and composition						
1. Number of species	19	1	12	1	19	
2. Sunfish species	6	5	4	3	. 4	
3. Sucker species	2	1	0	1	0	
4. Intolerant species	3 -	3	0	· 1	0	j
5. Percent tolerant species	3	5	23	1	1	-
B. Trophic composition						
6. Percent omnivores	2	5	20	1	0	4
7. Percent insectivores	73	5	67	3	86	5
C. Reproductive composition						
8. Migratory spawning species	4	3	0	1	1	1
9. Lithophylic spawning species	4	3	0	1	3	3
D. Abundance and fish health						
10. Number of individuals	184	1	137	1	971	5
11. Fish Health Assessment Index	52	3	36	3	54	3
RFAI		35		17		35
		Fair		Very		Fair
				poor		1.911

RESERVOIR VITAL SIGNS MONITORING - 1993

FISH COMMUNITY RESULTS TABLES

INTRODUCTION

Fish communities of TVA reservoirs have been sampled for littoral and bottom-dwelling pelagic species since fall 1989 to provide insight into the environmental quality of these reservoirs based on measurements of various parameters of the resident fish populations. Only 12 reservoirs were sampled in 1989-90, an additional 11 were added in 1991, and 7 more were added in 1993. The following tables detail results obtained during the 1993 fall fish community samples.

METHODS

Fish Community Monitoring

Shoreline electrofishing samples were collected during daylight hours from inflow, transition, and forebay zones of most reservoirs from September to mid-November, 1993. On relatively small reservoirs only one or two zones were sampled, with the forebay area always selected. A total of 15 electrofishing transects, each covering 300m of shoreline, were collected from each of the sampled zones. All habitats were sampled in proportion to their occurrence in the zone. Where conditions permitted, 12 experimental gill nets with five 6.1m panels (mesh sizes of 2.5, 5.1, 7.6, 10.2, and 12.7cm) were set for one overnight period in each reservoir zone. In forebay and transition zones, nets were set in all habitat types, alternating mesh sizes toward the shoreline between sets. Nets in inflow areas were set where flow conditions allowed which restricted the number of nets set in this zone.

Total length (mm) and weight (g) was obtained for all black bass species captured, length only was taken of other sport species and channel catfish, with other species being enumerated prior to release. Fish observed but not captured during electrofishing were included if positive identification could be made. Estimated counts were also used in electrofishing samples when high densities of fish were encountered, as long as identification was possible. Young-of-year fish were counted separately from adults and juveniles. All fish measured were inspected for external signs of disease, parasites, and anomalies. Natural hybrids were included as an anomaly.

Reservoir Fish Assemblage Index

Shoreline electrofishing and gill netting data collected during fall 1993 were input into a Reservoir Fish Assemblage Index (RFAI) developed by TVA and Dr. James Karr (University of Washington). A separate index used for electrofishing and gill netting. The RFAI uses 12 fish community metrics which can be broken into five general catagories:

- 1. Species richness and composition -- total number of species, piscivore species, sunfish species, sucker species, intolerant species, and percent tolerant individuals;
- 2. Trophic composition -- percent omnivores and percent insectivores;
- 3. Reproductive composition -- number of lithophilic spawners,
- 4. Abundance and health -- total catch per unit effort and percent individual with external diseases, parasites, and/or anomalies.

Scoring criteria for the 12 metrics were developed from ranges in fall electrofishing and gill netting data collected from TVA reservoirs from 1990 through 1993. The observed data for each gear type were trisected, such that values falling in the upper third were designated "good", and values falling in the middle and lower thirds were designated "fair" and "poor", respectively. In addition to gear type, independent scoring criteria were developed for each area (inflow, transition, and forebay) of both run-of-the-river and tributary reservoirs. Results of 1993 sampling were compared to the scoring criteria, and each of the 12 metrics were rated "5" if the observed value fell within the "good" range, or "3" or "1" if the observed value fell in "fair" or "poor" range, respectively.

The sum of the 12 metric ratrings constituted the RFAI value which summarizes the overall condition of a given reservoir fish community. Attainable RFAI values were divided into five categories (see below) to generally discribe the environmental condition of that section of a reservoir based on attributes of the resident littoral and bottom-dwelling pelagic fish communities. The average of the electrofishing and gill netting RFAIs was used to describe the overall fish community of a site.

Fish Community Evaluation

Total Score

Rating

 12-21
 Very Poor

 22-31
 Poor

 32-41
 Fair

 42-51
 Good

 52-60
 Excellent

Table 7. Species list and catch per unit effort at the inflow, transition, and forebay during fall electrofishing and gill netting on Chickamauga, 1993 (electrofishing effort = 300 meters of shoreline and gill netting effort = net-nights).

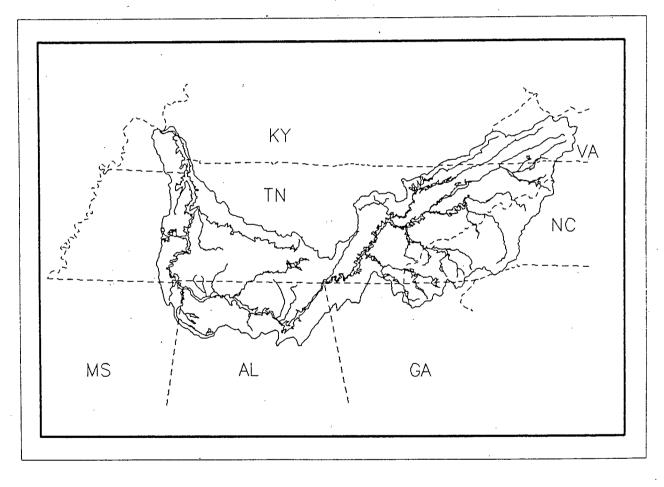
		Electrofishing	g	Gill Netting				
Common name	Inflow	Transition	Forebay	Inflow	Transition	Forebay		
		_						
Spotted gar	.	•	•		0.1			
Longnose gar	0.1	•	•	1.5	•	•		
Skipjack herring	1.7	•		3.5	4.8	3.3		
Gizzard shad	38	26.9	19.3	2.8	4.7	4.4		
Threadfin shad	3559.9	1707.4	810.2		•	•		
Mooneye		•	•		0.3	•		
Central stoneroller		0.1						
Carp	1.5	1.1	0.3	0.3	0.1	•		
Golden shiner		2.8	•		0.1	0.8		
Emerald shiner	59	60.1	50.3					
Spotfin shiner	0.9	0.9	0.1					
Steelcolor shiner	2.6		0.1			•		
Striped shiner	0.1		•			•		
Bluntnose minnow	0.1	•				•		
Quillback carpsucker			•	0.3	•	•		
Northern hog sucker	0.1		•	0.5	•	•		
Smallmouth buffalo		0.2	0.1	0.8	0:1	•		
Spotted sucker	1.1	1	0.1	0.5	0.5	•		
Black redhorse	0.1	•	•	0.5	0.5	•		
Golden redhorse	0.5	•	•	•	•	•		
Blue catfish	0.1	0.1	0.1	0.3	1.8			
Channel catfish	0.7	0.1	0.1	2.5		0.3		
Flathead catfish	0.7	0.1	.0.1		1.3	0.3		
White bass	0.7	0.1	0.1	0.3	0.5	0.2		
Yellow bass	5.3	0.1	0.1	5	9	0.9		
Striped bass	0.5	0.1	•	13	6.2	2.8		
Hybrid striped x white bass	0.5	•	•	1.5	•	•		
Warmouth				0.3	•	•		
Redbreast sunfish	1.1	0.7	0.1	0.3	0.1	•		
Green sunfish	0.3	1	1.8	•	•	•		
Bluegill	0.1		0.2	•	•	•		
-	30.7	30.1	11.7	0.3	0.5	0.3		
Longear sunfish Redear sunfish	0.2	0.5	0.1	•	•	•		
	. 10	4.4	1.8	1.3	2.2	0.7		
Smallmouth bass	2.6	0.4	1.3	•	0.1	0.2		
Spotted bass	5.7	6.2	7.3	1.3	2.5	2.3		
argemouth bass	8.1	6.6	1.3	0.5	0.3	0.2		
White crappie	0.3	0.1		•	0.3	0.5		
Black crappie	1	1.4		0.3	1.2	1.2		
fellow perch	0.2	1	0.1	•	0.8	0.1		
ogperch	0.5	5.3	0.2	•				
auger	•	•	0.1	1	0.1	0.1		
Valleye	•	•		1	•	•		
reshwater drum	0.3	0.4	0.1	1.8	0.7	0.8		
Brook silverside	0.8	8.7	8.5	•		•		
otal	3735.4	1867.5	915.1	39.8	37.9	19.1		
Number of samples	15	15	15	4	12	12		
Sumber collected	56031	28013	13727	159	455	229		
pecies collected	35	26	24	23	24	18		

Table 7b. Scoring results for tweleve metrics and overall Reservoir Fish Assemblage Index (RFAI) for Chickamauga Reservoir.

		Electrofishing						Gill Netting			
Metric	In	Inflow		Transition		Forebay		Transition		Forebay	
	Obs.	Score	Obs.	Score	Obs	Score	Obs.	Score	Obs.	Scor	
A. Species richness and composition											
1. Number of species	35	5	26	5	24	3.	24	5	18	3	
2. Piscivore species	11	5	7	3	6	1	11	5	10	5	
3. Sunfish species	6	5	5	3	6	5	3	5	2	3	
4. Sucker species	4	3	2	3	1	1	2	3	õ	1	
5. Intolerant species	5	5	2	3	1	1	2	3	ĩ	i	
6. Percent tolerant species	56.6%	3	59.7%	3	67.1%	1	12.8%	5	27.1%	3	
7. Dominance (% composition	33.7%	3	39.2%	5	61.4%	1	23.8%	5	23.1%	5	
of most abundant B. Trophic compostion										-	
8. Percent omnivores	23.1%	5	20.0%	5	3.6%	5	21.1%	5	30.1%	5	
9. Percent insectivores	62.0%	5	70.7%	5	90.2%	5	13.2%	.3	9.2%	3	
. Reproductive composition					÷.						
10. Lithophilic spawning species	8	5	4	3	4	3	5	3	3	1	
. Fish abundance and health										:	
11. Average number of individuals	174.9	5	153.3	5	81.9	3	37.8	5	19.1	1	
12. Percent anomalies	2.2%	3	2.5%	3	2.3%	3	0.2%	5	0.0%	5	
RFAI		52 excellent		46 good		32 fair	·· ·	52 excellent	·····	36 fair	

Tennessee Valley Authority Water Resources Division Chattanooga, Tennessee TVA/WR--91/1 August 1991

RESERVOIR MONITORING - 1990 SUMMARY OF VITAL SIGNS AND USE IMPAIRMENT MONITORING ON TENNESSEE VALLEY RESERVOIRS



WATER RESOURCES & ECOLOGICAL MONITORING

<u>Chickamauga Reservoir</u>

Chickamauga Reservoir could be described as an "average" mainstream Tennessee River reservoir. With the dam at TRM 471.0, it has a length of 59 miles, a shoreline of 810 miles, and a surface area of 35,400 acres at full pool. The average annual discharge is 33,099 cfs which provides an average hydraulic retention of ten days.

Vital Signs Monitoring on Chickamauga Reservoir in 1990 identified generally healthy conditions, although some undesirable conditions were found, especially at the inflow site. Overall, the "health" of the aquatic resources in Chickamauga Reservoir were in the midrange compared to the other mainstem reservoirs.

Lack of thermal and DO stratification, presence of an active algal community, water clarity, generally good sediment quality, and healthy/diverse benthic and fish communities at the forebay and transition zone were all desirable characteristics. Relatively few benthic macroinvertebrate taxa and number of organisms per square-meter as well as low number of fish collected in electrofishing efforts were indicative of undesirable conditions at the inflow site.

A high silt content of sediments and relatively high chlorophyll concentrations at the transition zone indicate this site may be too far downstream. Results from 1991 will be reviewed closely to determine if movement of this site upstream is appropriate.

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Synopsis of 1990 conditions:

<u>Water--Maximum</u> water temperature was generally cooler at the forebay (28.0°C) and transition zone (26.5°C) than at comparable sites on other mainstream reservoirs. There were no substantial surface-to-bottom differences in temperature and DO. The minimum DO (at bottom) was 3.1 mg/l in June at the forebay and 4.2 mg/l in July at the transition zone. Supersaturated DO levels and high pH values (>8.5) occurred in April and May at both the forebay and transition zone sites indicating photosynthetic activity. Maximum chlorophyll-a concentrations of 24 ug/l at the forebay and 17 ug/l at the transition zone occurred in June. Relatively low phosphorus concentrations may have been a factor limiting algal growth on some occasions. Bacteriological levels were acceptable except for one sample (440/100 ml) collected at the forebay in January during high reservoir flows.

<u>Sediment</u>--Chemical analyses of sediments revealed no metal or organic analyte to be a concern. Toxicity tests detected a slight decrease in light emitted by the test organism, indicating a potential for toxicity at both the forebay and transition zone. Particle size analysis showed forebay sediments were almost totally silt and clay (98 percent). Likewise sediments at the transition zone were comprised of mostly silt and clay (84 percent silt and clay and 16 percent sand). Results for the transition zone may indicate this site needs to be moved upstream.

Benthic Macroinvertebrates--The benthos at the forebay and transition zone were both abundant and diverse (14 taxa and 614 organisms at the forebay; 12 taxa and 956 organisms at the transition zone) compared to similar sites on the other mainstream reservoirs. At the inflow only six taxa and 191 organisms were collected, which was quite low compared to other mainstream reservoir inflows sites. The hardy tubificid worms were dominant (49 percent) at the inflow, whereas the more typically encountered chironomid <u>Coelotanyopus</u> was the most numerous taxon at the forebay (30 percent) and transition zone (18 percent).

<u>Fish</u>--Fish collections in the open water areas of Chickamauga Reservoir were overwhelmingly dominated by threadfin shad (100, 98, and 98 percent at the forebay, transition zone, and inflow, respectively). Threadfin shad densities may have been enhanced by a considerable decline in aquatic vegetation during 1990 over that observed during the previous five years. This habitat change was caused by increased inflow and resulting turbidity and nutrient loading during late winter and spring 1990. Both number of fish and biomass were similar to or greater than levels observed in the other mainstream reservoirs. Fish collections at near shore areas and offshore bottom areas showed threadfin shad to be the most abundant species at the forebay and transition zone comprising 46 percent of the 2,929 fish per unit effort of electrofishing at the forebay and 64 percent of the 2,493 fish at the transition zone. Threadfin shad were absent from inflow samples with gizzard shad being the most numerous species making up 30 percent of the 313 fish collected. Collection success at the inflow was poor relative to other mainstream inflows. The number of fish species was relatively good at all three site (34, 33, and 32 from downstream to upstream). The FHAI showed the health of largemouth bass to be better than the mainstream average (40) at the inflow (29) and poorer than averages (53 and 51) for the transition zone (69) and forebay (66).

The fish assemblage on Chickamauga Reservoir was worse than the average fish community sampled on other Tennessee River mainstream reservoirs. Problems contributing to this below average status include low values for species richness; largemouth bass health; density of piscivores and specialist feeders; number of intolerant species; overall electrofishing catch rate; gill netting catch rates; relative weight of bass; PSDs of selected species; and overall fish abundance. Inspection of fish assemblages within the reservoir revealed that the quality of the forebay environment was slightly better than that found in the average mainstream forebay, the transition zone environment was average and the inflow was worse than the average mainstream reservoir. Comparison of the 1990 results with those obtained in 1989 showed considerable improvement in the fish assemblage of the reservoir between the two years.

Watts Bar Reservoir

Watts Bar Reservoir impounds water from both the Tennessee River and one of the major tributaries to the Tennessee River, the Clinch River. The three dams which bound Watts Bar Reservoir are Watts Bar Dam (located at TRM 529.9), Fort Loudoun Dam (located at TRM 602.3), and Melton Hill Dam located at Clinch River Mile (CRM) 23.1. The total length of Watts Bar Reservoir including the Clinch River arm is 96 miles, the shoreline is 783 miles, and the surface area is 39,000 acres. The average annual discharge from Watts Bar is 27,145 cfs providing as average hydraulic retention time of 19 days.

The confluence of the Clinch and Tennessee Rivers is upstream of the transition zone in Watts Bar, so biological sampling was conducted at the forebay, transition zone, and both the inflow on the Tennessee River and the inflow on the Clinch River. Water entering from the Clinch River arm from Melton Hill Reservoir is quite cool due to hypolimnetic withdrawal from Norris Reservoir (a deep storage impoundment) upstream from Melton Hill. Water entering Watts Bar Reservoir from Fort Loudoun Dam is usually lower in DO during summer months than water entering from Melton Hill Dam.

Vital Signs Monitoring results for Watts Bar Reservoir in 1990 identified several undesirable conditions. At least one monitoring tool (in most cases more than one) identified undesirable conditions at all sample sites. As a result, aquatic environmental resources in Watts Bar would grade below the midrange compared to the other mainstem reservoirs.

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The most significant problems were presence of a strong oxycline with near anoxic conditions during summer at the forebay, presence of relatively high levels of mercury and toxic conditions in the sediments at the transition zone, and relatively poor benthic macroinvertebrate faunas (in terms of abundance and species richness) at both the Tennessee and Clinch River inflow sites as well as few fish collected at the Clinch inflow site.

Synopsis of 1990 conditions:

Water--The maximum water temperature (28.3°C) was observed in July with the greatest surface to bottom temperature differential (6°C) observed during approximately the same time frame. Otherwise, only weak thermal stratification was observed during the monitoring period. A rather strong oxygen gradient (up to 11 mg/1 difference between surface and bottom) existed from June through August. DO concentrations near bottom were below 1 mg/l during much of this period. Upper strata DO levels were high, generally supersaturated, throughout much of this period. These supersaturated DO levels, coupled with high pH values (e.g., 9.2 in June) indicate a high rate of algal photosynthesis. Obviously, nutrients and water clarity were sufficient to support this luxuriant algal growth. Chlorophyll-a concentrations were highest in May, with 20 ug/l at the forebay and 14 ug/l at the transition zone. Although a few of the bacteriological samples were positive, none exceeded a water contact recreation guideline of 200/100 ml.

<u>Sediment</u>--A relatively high concentration of mercury (0.95 ug/g dry weight) and the most toxic response from the sediments tested draw concern for the transition zone on Watts Bar Reservoir. A slight toxic response was also noted in tests on forebay sediments. Particle size analysis showed the substrate at the both the forebay and transition zone to be greater than 99 percent silt and clay.

<u>Benthic Macroinvertebrates</u>--The benthos was average or below in species richness and abundance at all four sample sites. At the forebay, the number of taxa (9) was relatively low, the number of organisms (498 per square-meter) was typical of other forebays, and <u>Chironomus</u> was the most numerous taxon (40 percent of the total). The transition zone was represented by an average to high number of taxa (12) yet a low number of organisms (316) with the mayfly (<u>Hexagenia</u>) the most numerous taxon. The inflow on the Tennessee River had an average to high number of taxa (11) but a very low number of organisms (58) with <u>Corbicula</u> the most numerous organism (31 percent). The inflow on the Clinch River had the poorest benthic macroinvertebrate community of all inflows with only three taxa and 42 organisms, dominated by <u>Corbicula</u> (83 percent).

<u>Fish</u>-Fish data for the open water areas showed Watts Bar to have relatively high density and biomass at all four sample sites. In several instances these were the highest (density or biomass) levels observed at any Vital Signs Monitoring reservoir. Threadfin shad was practically the only species present making up almost 100 percent of the catch at the forebay, transition zone, and Clinch River inflow, and 98 percent at the Tennessee River inflow.

Near shore area and offshore area bottom collections of fish were generally typical of other mainstream reservoirs. A distinct exception was a very low average number of fish (57 per unit effort) collected in electrofishing efforts at the inflow on the Clinch River. Also, number of fish collected at the forebay (757 per unit effort) was low relative to other forebay areas. Number of species (33, 35, 37, and 28 from downstream to upstream on the Tennessee and at the Clinch inflow, respectively) was generally representative of comparable areas on other mainstream reservoirs. Number of fish collected per unit effort of electrofishing at the transition zone (2,178) and Tennessee River inflow (1,783) were typical to high. Threadfin shad was dominant at the forebay (32 percent), the transition zone (52 percent), the Tennessee River inflow (84 percent), but absent at the Clinch River inflow where gizzard shad was most numerous (40 percent of the catch). The FHAI scored the health of largemouth bass to be about average at the forebay (48), less than average at the Tennessee River inflow (53), and better than average at the transition zone (27). Largemouth bass were not captured in sufficient numbers to obtain reliable results in the Clinch inflow.

The fish assemblage on Watts Bar Reservoir (all areas combined) was worse than average for other mainstream Tennessee River reservoirs. Characteristics of the fish community which were below average included species richness, trophic composition, electrofishing catch rates of selected species, overall fish abundance, gill net catch rates of Stizostedion species, mean relative weight of bass, and PSDs of smallmouth bass and channel catfish. Inspection of fish assemblages within reservoir zones revealed that the quality of the Clinch River inflow was worse and the Tennessee River inflow was similar to that found in the average mainstream inflow. The transition zone on Watts Bar maintained a fish community slightly better than that found in the mainstream average and the fish assemblage in the forebay was slightly worse than average. Comparison of 1990 results with those obtained in 1989 revealed a slight decline in the fish assemblage of the reservoir, mainly in species richness, gill net catch rates, and overall fish abundance.

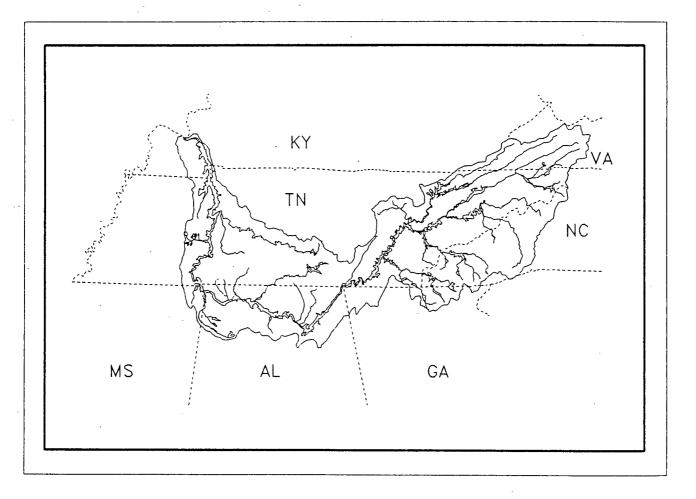
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Tennessee Valley Authority

名言語

Water Resources Division Chattanooga, Tennessee TVA/WR--92/8 July 1992

RESERVOIR VITAL SIGNS MONITORING - 1991 SUMMARY OF VITAL SIGNS AND USE IMPAIRMENT MONITORING ON TENNESSEE VALLEY RESERVOIRS



WATER RESOURCES & ECOLOGICAL MONITORING

4.7 Chickamauga Reservoir

4.7.1 Physical Description

Chickamauga Reservoir can be described as an "average" mainstream Tennessee River reservoir. Chickamauga Dam is located at TRM 471.0. The reservoir is 59 miles long, has 810 miles of shoreline, and has a surface area of 35,400 acres at full pool. The average annual discharge is 34,192 cfs which provides an average hydraulic retention of about ten days.

4.7.2 Reservoir Health

Vital Signs Monitoring on Chickamauga Reservoir in 1991 identified generally healthy conditions, although some undesirable conditions were found at the inflow site. Overall, the "health" of the aquatic resources in Chickamauga Reservoir would rate above average compared to the other mainstem reservoirs.

Lack of thermal and DO suratification, presence of an active (but not overly active) algal community, good water clarity, generally good sediment quality, and healthy/diverse benthic and fish communities at the forebay and transition zone were all desirable characteristics. Undesirable conditions at the inflow site included relatively few benthic macroinvertebrate taxa (dominated by one taxon), low number of fish species, and presence of a large proportion of tolerant fish species in electrofishing collections. These conditions and the overall evaluation of Chickamauga Reservoir were quite similar to those based on 1990 monitoring results.

A question arose from the 1990 results related to relocation of the transition zone sample site. Consideration of results from both 1990 and 1991 resulted in the decision not to change the location of this site.

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4.7.3 <u>Reservoir Use Suitability</u>

Use Suitability Monitoring activities did not identify any impairments on Chickamauga Reservoir. Bacteriological sampling in 1991 was limited to mid-channel collections in association with Vital Signs Monitoring activities. Fecal coliform bacteria were seldom documented, and when present they occurred at very low levels.

There are no fish tissue consumption advisories in effect for Chickamauga Reservoir. Composite fillets from channel catfish collected in autumn 1990 from the forebay, transition zone, and inflow were analyzed for metals, pesticides, and PCBs on the EPA priority pollutant Samples had low or nondetectable levels of most metals (except a list. relatively high concentration of lead in one sample) and pesticides (except slightly elevated levels of chlordane). PCBs were detected, but even the maximum was relatively low. An intensive examination of PCB concentrations in catfish was also conducted in 1991 on Chickamauga Reservoir because of the PCB problems upstream in Watts Bar Reservoir and downstream in Nickajack Reservoir. Ten catfish were collected from five locations and examined individually. Average PCB concentrations were relatively low in all samples, and few samples had a concentration which approached or exceeded 1.0 μ g/g. Many samples had less than detectable concentrations. As a result of these analyses, the TDEC did not include any fish species from Chickamauga Reservoir in their annual update on fish consumption for state waters issued February 27, 1992.

4.7.4 Synopsis of 1991 Conditions

<u>Water</u>--Surface temperatures ranged from 8.3°C in January to 29.9°C in July in the forebay and from 7.1°C to 27.6°C for the same months at the transition zone. Values for DO at the 1.5-meter depth ranged from

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10.4 mg/l in April to 6.0 mg/l in August at the forebay and from 10.1 mg/l in January to 5.5 mg/l in August at the transition zone.

Like many other mainstem Tennessee River reservoirs, Chickamauga is generally well mixed and lacks any strong thermal stratification. In May, a maximum temperature differential of 3.5°C was observed at the forebay. Minimum bottom DOs were measured in June of 3.0 mg/l and 3.4 mg/l, respectively, at the forebay and the transition zone.

Values of pH ranged from 7.0 to 8.2. Conductivity ranged from 117 to 182 μ mhos/cm, and averaged about 165 μ mhos/cm. Comparison of pH and conductivity at the transition zone with upstream pH and conductivity at Watts Bar Dam forebay shows these are lowered by Hiwassee River inflows to Chickamauga Reservoir about nine miles upstream of the transition zone.

Average total nitrogen concentrations were the lowest measured among Vital Signs Monitoring locations on the Tennessee River in 1991. In addition, both total phosphorus and dissolved ortho phosphorus concentrations were also among the lowest observed at any of the Vital Signs Monitoring locations on the Tennessee River.

The highest chlorophyll-a concentrations were measured in May ranging from 8-10 μ g/l and 11-13 μ g/l, respectively, at the forebay and transition zones. Concentrations of chlorophyll-a averaged about 7 μ g/l at both the forebay and the transition zone in 1991.

Forebay Secchi depth, turbidity, and suspended solids measurements averaged 1.4 meters, 5.7 NTU's, and 4.8 mg/l, respectively. Transition zone Secchi depths, turbidity, and suspended solids averaged 1.4 meters, 4.6 NTUs, and 4.2 mg/l, respectively. In addition, true color values averaged about 10 PCUs at the forebay and transition zones. Together these values indicate the light transparency of Chickamauga Reservoir to be high compared with the other mainstem Tennessee River reservoirs.

<u>Fecal Coliform Bacteria</u>--There were no swimming beaches on Chickamauga Reservoir examined in 1991. Monthly sampling as part of Vital Signs Monitoring did not detect any fecal coliform bacterial colonies in mid-channel at the forebay. Only one sample from the transition zone had a detectable concentration (20 colonies per 100 ml in April).

Sediment--A sediment sample collected from the forebay of Chickamauga Reservoir had a measured concentration of 67 μ g/kg of p,p-DDT. Samples collected in 1990 did not detect (<10 μ g/kg) DDT in sediment from any locations in the Tennessee Valley. Sediment samples collected from Chickamauga Reservoir in 1992 will be used to validate results from the single sample in 1991. There were no other points of concern about sediment quality in Chickamauga Reservoir. All metal and organic analyses were either not detected or found in low concentrations, and toxicity screening tests did not identify any toxic conditions. Particle-size analysis showed sediments were 97 percent silt and clay at the forebay. Transition zone sediments were also mostly silt and clay (83 percent) and sand (17 percent).

Benthic Macroinvertebrates--Collections from the forebay included 797 organisms representing 11 taxa. The chironomid <u>Coelotanyopus</u> (40 percent of the total) and the mayfly <u>Hexagenia</u> (20 percent) were the most numerous taxa collected. The transition zone had an average number of taxa (10) but had the greatest number of organisms collected (1283 per square meter) compared to other mainstream transitions zones. <u>Corbicula</u> accounted for 49 percent of the total, and <u>Hexagenia</u> accounted for 22 percent. The inflow site had relatively few taxa (8) and an average number of organisms (492 per square meter), however, <u>Corbicula</u> comprised 80 percent of the animals collected. Fish Community--Fish information for open-water areas collected with hydroacoustic equipment showed fish densities at the forebay were similar to most mainstream reservoir forebays. Densities at the transition zone were the highest found in the comparable area of any of the mainstream reservoirs. However, these results had very wide confidence intervals indicating the high mean density may have been due to encountering one or more unusually large school(s) of fish. Average fish size was greater at the inflow and lower at the transition zone and forebay when compared to the mean size calculated for equivalent areas of all mainstream reservoirs. Mean values were 4.1, 3.4, and 4.5 cm at the forebay, transition zone, and inflow.

Fish data collected in littoral and profundal zones of the forebay documented emerald shiner was the most abundant species (collected at the rate of 677 fish per electrofishing hour). Overall, emerald shiners accounted for 33 percent of the total number of fish collected. Other dominant species included bluegill (19 percent), gizzard shad (14 percent), YOY threadfin shad (9 percent), redear sunfish (4 percent), largemouth bass (2 percent), spotted bass (2 percent), and yellow perch (1 percent). Total fish abundance was greatest in the forebay due to the large number of emerald shiners, otherwise fish abundance at all three zones was similar.

Electrofishing RIBI analysis showed a fair quality littoral fish community in the inflow zone (RIBI = 33), and good quality communities in the transition zone (RIBI = 39) and forebay (RIBI = 35). The inflow score ranked fifth among other mainstream reservoir inflows, or about average. The transition zone appeared better and ranked second. The forebay ranked seventh (three other mainstream forebays had identical RIBI scores) which would be below average for mainstream forebays. (A below average value can still be ranked good because all the forebays, including those of storage reservoirs, were included in the original trisection of values to determine good, fair, and poor rankings.) All three zones scored good for total fish abundance, number of sunfish species, and percentage of inventories. However, total species diversities (21-25) only scored fair for the three zones, indicating some expected species were absent from the samples. This was especially true for suckers and intolerant species, and also apparent in the numbers of lithophilic broadcast spawning species and migratory spawning species.

Only one species of sucker was found at the inflow and transition zones and none in the forebay. The inflow was rated fair because of the relatively high percentages of tolerant individuals and omnivorous individuals. The other two zones were rated good. The health of largemouth bass was depressed at all three zones, as FHAI values ranged from 75 to 83.

Fish Tissue--There are no fish tissue consumption advisories in effect for Chickamauga Reservoir. Two types of fish tissue studies were conducted on this reservoir in autumn 1990. In one study, fillets from five channel catfish were composited from each site and examined for a broad array of analyses (metals, pesticides, and PCBs on the EPA priority pollutant list). Results from samples collected from the forebay, transition zone, and inflow had low or nondetectable levels of most metals (except lead at 0.80 μ g/g in one sample) and pesticides (except chlordane at 0.10 μ g/g). PCBs were detected but even the maximum was relatively low. In the other study, fillets from ten channel catfish from five locations within the reservoir were examined individually for PCBs. This intensive study was conducted because of the PCB problems upstream in Watts Bar Reservoir and downstream in Nickajack Reservoir. Average PCB concentrations were relatively low in all samples (maximum mean 0.7 μ g/g) near Watts Bar Dam. Many samples had less than detectable concentrations, and few exceeded 1.0 $\mu g/g$. As a result of these analyses, the TDEC did not include any fish species from Chickamauga Reservoir in their annual update on fish consumption advisories for state waters issued February 27, 1992.

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4.8.1 Physical Description

Watts Bar Reservoir impounds water from both the Tennessee River and one of the major tributaries to the Tennessee River, the Clinch River. The three dams which bound Watts Bar Reservoir are Watts Bar Dam (located at TRM 529.9), Fort Loudoun Dam (located at TRM 602.3), and Melton Hill Dam located at Clinch River mile (CRM) 23.1. The total length of Watts Bar Reservoir, including the Clinch River arm is 96 miles, the shoreline is 783 miles, and the surface area is 39,000 acres. The average annual discharge from Watts Bar is 27,849 cfs providing an average hydraulic retention time of about 19 days.

The confluence of the Clinch and Tennessee Rivers is upstream of the transition zone in Watts Bar, so biological sampling was conducted at the forebay, transition zone, and both the inflow on the Tennessee River and the inflow on the Clinch River. Water entering from the Clinch River arm from Melton Hill Reservoir is quite cool due to hypolimnetic withdrawal from Norris Reservoir (a deep storage impoundment) upstream from Melton Hill. Water entering Watts Bar Reservoir from Fort Loudoun Dam is usually warmer and lower in DO during summer months than water entering from Melton Hill Dam.

4.8.2 Reservoir Health

Vital Signs Monitoring results for Watts Bar Reservoir in 1991 identified generally fair conditions, but there was at least one undesirable condition at each sample site. As a result, aquatic environmental resources in Watts Bar ranked below the mid-range compared to the other mainstem reservoirs. The most significant problems were presence of a strong oxycline with near anoxic conditions during summer at the forebay, presence of mercury in the sediments at the forebay and transition zone, and relatively poor benthic macroinvertebrate fauna at the Tennessee River inflow. Within Watts Bar Reservoir the highest quality aquatic resources were at the transition zone.

4.8.3 <u>Reservoir Use Suitability</u>

Use Suitability Monitoring activities did not identify any bacteriological problems on Watts Bar Reservoir in 1991. Bacteriological sampling was limited to mid-channel collections in association with Vital Signs Monitoring activities. Fecal coliform bacteria were below levels of detection in all samples.

As a result of PCB contamination, the TDEC has issued advisories on consumption of several species of fish from Watts Bar. TVA participates on a study team with state agencies and the Oak Ridge National Laboratory to monitor this situation. A variety of species from several locations on the main portion of reservoir (i.e., not in embayments) are examined each year. Results from fish collected in these areas in autumn 1990 showed little differences from those collected the previous year. Details of these results are provided in Bates et al. (1992).

A special embayment study was conducted on Watts Bar Reservoir in autumn 1990 because of the importance of embayments as fishing areas. Channel catfish, largemouth bass, and crappie (black and white mixed) were analyzed for PCBs and chlordane from two places in the Piney River embayment and one place in the Whites Creek embayment (the two largest embayments on the reservoir). All crappie and largemouth bass had either

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nondetectable or only low levels of PCBs and chlordane. Concentrations in catfish from Whites Creek were also nondetectable or quite low. Most catfish from Piney River had detectable concentrations of PCBs which did not differ greatly from those in catfish from the forebay sample site.

4.8.4 Synopsis of 1991 Conditions

Water--Surface water temperatures ranged from 7.2°C in January to 30.2°C in July in the forebay and from 7.7°C to 28.4°C for these same months at the transition zone. Values for DO at the 1.5-meter depth ranged from 12.8 mg/l in April (due to high photosynthetic activity) to 8.1 mg/l in September at the forebay and from 11.2 mg/l in January to 6.6 mg/l in September at the transition zone.

Temperature and dissolved oxygen data show the reservoir to be well mixed early in the year and developing a moderate degree of thermal stratification at the forebay in July and August. A maximum temperature differential (surface to bottom) of 8°C occurred in May. DO versus depth data show a rather strong oxycline to develop in the forebay of Watts Bar Reservoir in June and July. In June and July, about a 10 mg/l decrease (surface to bottom) in DO was measured in Watts Bar forebay; near bottom DO concentrations in the hypolimion were less than 1 mg/l. The transition zone was well mixed. Minimum bottom DO measured at the transition zone was 4.4 mg/l.

Values of pH ranged from 7.0 to 9.2 on Watts Bar Reservoir. In April, May, June, and July, near surface values of pH in the forebay were high, equal to or exceeding 9.0, and DO saturation values were high, ranging from 125-150 percent, indicating high rates of photosynthesis.

The average total phosphorus concentrations observed at the forebay were lower than any of the other Tennessee River Vital Signs Monitoring locations. The average dissolved ortho phosphorus concentrations of 0.008 and 0.009 mg/l, respectively, at the forebay and transition zones were essentially identical to the average concentrations of dissolved ortho phosphorus in Chickamauga Reservoir and were among the lowest observed at any of the Tennessee River Vital Signs Monitoring locations in 1991.

The highest chlorophyll-a concentrations were measured in August at the forebay (19 μ g/l) and in July at the transition zone (13 μ g/l). Surface concentrations of chlorophyll-a averaged about 12 μ g/l at the forebay and about 8 μ g/l at the transition zone in 1991.

Forebay Secchi depth, turbidity, and suspended solids measurements averaged 1.5 meters, 4.4 NTU's, and 5.7 mg/l, respectively. These values indicate the light transparency of Watts Bar Reservoir forebay to be among the highest of the mainstem Tennessee River reservoirs in 1991. <u>Fecal Coliform Bacteria</u>--These were no swimming beaches on Watts Bar Reservoir examined as part of this monitoring program in 1991. Monthly samples collected in mid-channel at the forebay and transition zone as part of Vital Signs Monitoring activities had less than detectable concentrations in all samples.

<u>Sediment</u>--Elevated concentrations of mercury were again detected in the sediment of Watts Bar reservoir in 1991. Concentrations of 0.51 and 0.69 μ g/kg were measured in the forebay and transition zone, respectively. The most likely source of this contamination is past operations at Oak Ridge National Laboratory where major environmental cleanup activities are now underway. Although Microtox provided an indication of toxicity in transition zone pore water in 1990, there was no toxicity in either Microtox or Rototox tests in 1991. Sediments were almost entirely silt and clay (99 percent) at both the forebay and transition.

<u>Benthic Macroinvertebrates--An average number of taxa (11) were</u> collected in the forebay; however, there were relatively few organisms compared to other mainstream forebays (455 per square meter) and 43 percent of the total was <u>Chironomus</u>. The transition zone site and both inflow sites had a relatively high number of organisms. The transition site had 750 organisms per square meter (12 taxa) with the most numerous taxa being <u>Hexagenia</u> (20 percent) and <u>Coelotanyopus</u> (19 percent). The Tennessee River inflow site had 12 taxa and 513 organisms per square meter. The Clinch River had the most taxa (21) found in the mainstream inflow sites and 545 organisms per square meter. <u>Corbicula</u> was the dominant taxon in both inflow sites comprising 66 percent in the Tennessee River and 73 percent in the Clinch River samples.

<u>Aquatic Macrophytes</u>--An estimated 10 acres of aquatic plants were on Watts Bar Reservoir in 1991. Only 80 acres were present in 1990. In the late 1980s, populations were at about 600 to 700 acres and were dominated by Eurasian watermilfoil and spinyleaf naiad.

<u>Fish Community</u>-Fish information from open-water areas based on hydroacoustic equipment showed unusually high numbers of fish and extremely wide confidence interval when compared to equivalent areas on other mainstream reservoirs. This reflects a dense school of fish in the area at the time of the survey. There was also a reduced number of transects in this area resulting in a small volume of water being sampled acoustically. Both of these factors resulted in an estimate of fish density higher than what might be expected for this area. Fish densities at the remaining three Tennessee River sample areas were about average. Average fish size was the smallest at the Clinch River inflow and largest at the transition zone. Each of the other areas had values less than the mean size calculated for equivalent areas in all mainstream reservoirs. Values for the forebay, transition zone, Tennessee River inflow, and Clinch River inflow were 3.7, 4.7, 3.1 and 1.8 cm, respectively.

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Shoreline electrofishing and offshore/deep gill netting sampled a total of 4432 fish represented by 43 species. Three species made up the majority of the overall sample: gizzard shad (20 percent), bluegill (18 percent), and brook silverside (17 percent). Other subdominant species included threadfin shad (7 percent), skipjack herring (5 percent), emerald shiner (5 percent), spotfin shiner (4 percent), carp (2 percent), and largemouth bass (2 percent). Fish were most abundant in the transition zone (2021), followed by the forebay (1339), and the Tennessee River inflow zone (723) and were least abundant in the Clinch River inflow (349). Each of the four zones sampled yielded 30-31 species. YOY threadfin shad were found only in the forebay. FHAI analysis found largemouth bass health to be fair in the Tennessee inflow (52) and the transition (65), and poor in the forebay (73). No FHAI was possible in the Clinch inflow due to low numbers of largemouth bass collected.

RIBI analysis of shoreline electrofishing data indicated fair littoral fish communities in the two inflow zones (Clinch Arm RIBI = 35, Tennessee Arm RIBI = 31) and the forebay (RIBI = 33). The transition zone (RIBI = 37) was designated good. Compared to other mainstream reservoirs, the Clinch inflow ranked fourth, while the Tennessee inflow ranked seventh. The transition zone ranked fourth, slightly better than average, and the the forebay ranked tenth, next to the worst. In spite of the lower total numbers of fish sampled in the inflow stations, more diversity was found in sucker species, intolerant species, migratory spawning species, and lithophilic spawning species there than either the transition zone or the forebay. Sunfish diversity was rated good in the transition designation were percentages of tolerant individuals, omnivores, and inventories, and overall number of fish sampled.

Fish Tissue--Fish from Watts Bar Reservoir have been under intensive investigation for several years because of PCB contamination. TDEC has issued an advisory warning to the public not to eat certain species and to limit consumption of other species. Two of these species (channel catfish and striped bass, including striped bass X white bass hybrids) were reexamined in autumn 1990 as part of the continuing study to remain abreast of conditions in this reservoir. These fish were examined individually for PCBs and pesticides. Results showed maximum PCB concentrations were generally higher in 1990 than in 1989; however, mean concentrations were generally similar between the two years. The maximum concentration in an individual channel catfish in 1990 was 5.8 μ g/g, and the greatest average concentration in channel catfish at the site was 1.6 μ g/g. Parallel concentrations in striped bass were maximum individual 4.7 μ g/g and maximum average 1.3 μ g/g. Concentrations tended to be greater in upstream reservoir areas, especially toward the Tennessee River inflow, than in lower reservoir areas near the forebay. Overall, PCB concentrations were lower in 1989 and 1990 compared to those in 1988. Most pesticides were not detected in any of the 1990 samples. Only chlordane was routinely detected with concentrations in most samples <0.10 μ g/g. However, a few samples

exceeded this level and one channel catfish (concentration of 0.34 μ g/g) exceeded the FDA action limit of 0.30 μ g/g for chlordane. The maximum average chlordane concentration at each sample site was 0.11 μ g/g for channel catfish and 0.13 μ g/g for striped bass.

Channel catfish composites from selected sites were analyzed for metals. All 12 metals included in the analyses were relatively low. Even mercury, which was found in sediments at the forebay and transition zone, was low with a maximum of 0.2 μ g/g.

A special embayment study was conducted on Watts Bar Reservoir in autumn 1990. Fish for the continuing study referenced above are collected from the main river portion of the reservoir and are not collected from embayments. Ten individuals each of channel catfish, largemouth bass, and crappie (black and white mixed) were analyzed for PCBs and chlordane from two places in the Piney River embayment and one place in the Whites Creek embayment. There were no PCBs or chlordane detected in any of the 30 crappie examined. Only a few (7 of 30) of the largemouth bass had detectable concentrations and most of the seven had concentrations at the level of detection (0.1 μ g/g). The maximum found in a largemouth bass was 0.5 μ g/g. Chlordane was detected in only one largemouth bass at a concentration of 0.05 μ g/g. As expected, catfish tended to have higher concentrations of both PCBs and chlordane than crappie or largemouth bass. Averages at the two locations in Piney River embayment were 0.6 μ g/g and 0.4 μ g/g with a maximum of 1.1 μ g/g at both sites. Chlordane concentrations averaged 0.05 and 0.02 μ g/g at the two Piney River sites. PCB concentrations in catfish from Piney River were generally similar to those observed out in the main portion of the reservoir near the forebay. Catfish from Whites Creek had lower PCB concentrations with detectable levels in only two of the ten examined and as average of 0.1 μ g/g.

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Tennessee Valley Authority

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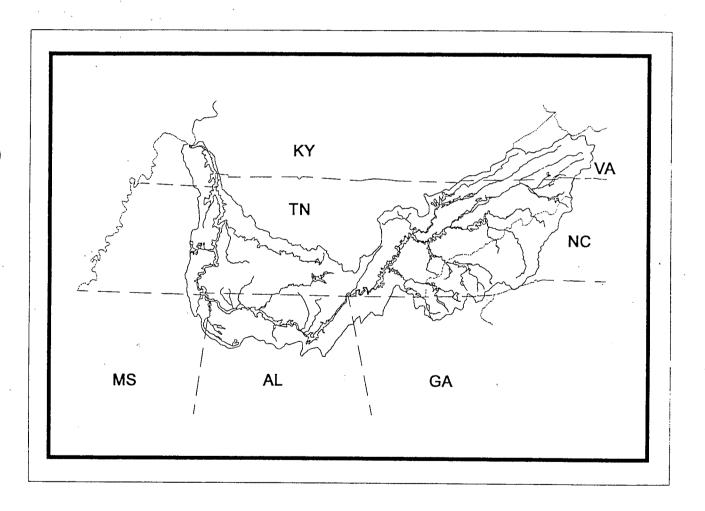
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Water Management Chattanooga, Tennessee August 1993

RESERVOIR MONITORING - 1992

SUMMARY OF VITAL SIGNS AND USE SUITABILITY MONITORING ON TENNESSEE VALLEY RESERVOIRS





4.7 Chickamauga Reservoir

4.7.1 <u>Physical Description</u>

Chickamauga Reservoir can be described as an "average" mainstream Tennessee River reservoir. Chickamauga Dam is located at TRM 471.0. The reservoir is 59 miles long, has 810 miles of shoreline, and has a surface area of 35,400 acres at full pool. The average annual discharge is approximately 34,000 cfs which provides an average hydraulic retention of nine to ten days (table 3.1).

A major tributary to the Tennessee River, the Hiwassee River, flows into the middle portion of Chickamauga Reservoir at about TRM 499. The flow from the entire Hiwassee River watershed contributes approximately 16.5 percent of the flow through Chickamauga Reservoir. The Hiwassee River just below Appalachia Dam (which does not include any flow from the Ocoee River or any other downstream tributaries) contributes about 6.5 percent of the flow of the Tennessee River through Chickamauga Reservoir.

4.7.2 <u>Reservoir Health</u>

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The overall ecological health rating for Chickamauga Reservoir was good in 1992, although only marginally so. Several health indicators scored lower in 1992 than in 1991. Sediment quality ratings changed from good in 1991 to poor in 1992 at the forebay and transition zone. Both of the tests used to evaluate sediment quality: chemical examination for heavy metals; and survival of test organisms in water extracted from the sediments (Microtox[®] and Rotox[®]), indicated poor conditions at both locations tested in 1992. Elevated concentrations of copper and zinc were found in the sediment in Chickamauga Reservoir. Both Microtox[®] and Rotox[®] tests showed low survival for the test organisms at the forebay, indicating potential sediment toxicity. Rotox[•] tests also indicated potential sediment toxicity at the transition zone.

DO was rated fair at the transition zone because DO was measured less than the state standard of 5 mg/L at the five-foot depth in September. DO was rated fair at the inflow to Chickamauga Reservoir due to the release of water with DOs less than 5 mg/L from Watts Bar Dam in July 1992. A poor benthic community was also found in the inflow, with a small number of benthic macroinvertebrate taxa. A representative fish community sample could not be collected at the transition zone in 1992 because of particularly adverse weather conditions during the field survey.

Aquatic macrophytes on Chickamauga Reservoir covered 387 acres in 1992 compared to 680 acres in 1991. Aquatic macrophytes peaked at about 7500 acres in 1988 and have continuously declined since then.

The ecological health of the fixed station monitoring site on the Hiwassee River was good in 1992. All ecological health indicators (nutrients, sediment quality, benthic community, and fish community) rated either good or fair.

4.7.3 <u>Reservoir Use Suitability</u>

There are no fish consumption advisories for Chickamauga Reservoir. Fillets from Chickamauga Reservoir catfish have been examined for several years as part of a variety of studies. Study results have indicated no consistent or reservoir-wide Results from most of these studies have usually found problems. higher concentrations of PCBs in catfish from the inflow area than from other sites in the reservoir. An intensive study was conducted in autumn 1990 because of the PCB problems upstream in Watts Bar Reservoir and downstream in Nickajack Reservoir. Ten catfish were collected from five locations and examined individually. Average PCB concentrations were relatively low in all samples with many samples having less than detectable concentrations. Channel catfish were collected for screening purposes in autumn 1991 and autumn 1992 from the inflow, transition zone, and forebay. In 1991 concentrations of all

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analytes from all locations were low, except PCBs at the inflow $(1.2 \ \mu g/g)$. This was also the case in 1992, except even the PCB concentration at the inflow was low (0.7 μ g/g) relative to most previous studies. No bacteriological studies were conducted at swimming beaches on Chickamauga Reservoir in 1992. However, the most recent data show that swimming areas previously tested on Chickamauga Reservoir fully support water contact recreation. The public use area near the dam and Lake Junior were surveyed in 1989 and seven swimming areas were surveyed in 1990, and all were found safe for water contact recreation at that time. Monthly Vital Signs sampling in 1992, at the forebay and transition zone in the open water portion of Chickamauga Reservoir, found all samples at or below the detection limit.

4.7.4 Synopsis of 1992 Conditions

<u>Water</u>--Surface temperatures ranged from 6.8°C in January to 28.0°C in July in the forebay and from 6.1°C to 26.1°C for the same months at the transition zone. Values for DO at the 1.5-meter depth ranged from 11.4 mg/L in January to 5.4 mg/L in September at the forebay and from 11.5 mg/L to 4.6 mg/L for these same months at the transition zone. The 4.6 mg/L concentration of DO at the 1.5-meter depth is the lowest in-reservoir DO measured at the 1.5-meter depth on any of the Vital Signs reservoirs in 1992, and is less than the state of Tennessee minimum water quality criteria for fish and aquatic life of 5.0 mg/L. The lowest measured DO in Chickamauga Reservoir in 1992 was 2.8 mg/L, found at the bottom of the forebay in July.

Like many other mainstem Tennessee River reservoirs, Chickamauga is generally well mixed and lacks any strong thermal stratification. However, the low flows of the Tennessee River system in April and early May facilitated the development of a weak thermocline and oxycline in these months at both the forebay and transition zone sampling locations, in 1992. Maximum temperature differentials (surface to bottom) of 4.5°C and 3.0°C were observed at the forebay, in April and May, respectively. At the transition zone, in April and May, maximum temperature differentials of 2.4°C and 3.0°C, respectively, were measured. During these same two months, oxygen differentials of 3.2 mg/L and 5.8 mg/L, respectively, were measured at the forebay; and, 3.3 mg/L and 4.7 mg/L, respectively, were measured at the transition zone. (The larger oxygen differentials measured in May were a result of high DOs at the water surface during a period of high photosynthetic activity.) Minimum DOs measured in

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hickamauga Reservoir in 1992 were 2.8 mg/L and 3.5 mg/L, at the bottom of the forebay and the transition zone, respectively, in July.

Values of pH ranged from 7.0 to 8.6. Conductivity ranged from about 155 to 195 μ mhos/cm, and averaged about 170 μ mhos/cm. Comparison of pH and conductivity at the transition zone with upstream pH and conductivity at Watts Bar Dam forebay indicates these are lowered by the soft water inflows of the Hiwassee River to Chickamauga Reservoir, about nine miles upstream of the transition zone.

Average total nitrogen concentrations in Chickamauga Reservoir were among the lowest measured at Vital Signs Monitoring locations on the Tennessee River in 1992. In addition, both total phosphorus and dissolved orthophosphorus concentrations were also among the lowest observed at any of the Vital Signs Monitoring locations on the Tennessee River.

The highest chlorophyll-a concentrations were measured in May, 12 μ g/L and 7 μ g/L, respectively, at the forebay and transition zones. Concentrations of chlorophyll-a averaged 6-7 μ g/L at the forebay and 4-5 μ g/L at the transition zone in 1992.

Fecal Coliform Bacteria--No bacteriological studies were conducted at swimming beaches on Chickamauga Reservoir in 1992. Monthly Vital Signs sampling at the forebay and transition zone found the June bacteria samples at the detection limit at each station. All other samples were below the detection limit.

<u>Sediment</u>--As in 1991, sediment samples collected in Chickamauga Reservoir in 1992 had slightly elevated concentrations of copper and zinc. In 1991, screening tests did not identify any potential toxic conditions. However in 1992, sediment collected from the forebay showed toxic effects on test organisms for both the Microtox[®] test (EC_{10} =13 percent) and the Rotox[®] test (rotifer survival=56 percent); and sediment collected from the transition zone also showed toxic effects on test organisms for the Rotox[®] test (rotifer survival=87 percent).

Particle size analysis showed sediments were 97 percent silt and clay at the forebay; and transition zone sediments were mostly silt and clay (80 percent) and sand (20 percent).

<u>Benthic Macroinvertebrates</u>--The forebay and transition zone benthic macroinvertebrate communities rated good, while the inflow communities rated poor. The forebay had 13 taxa and 900 organisms/m². The most numerous taxa collected were the mayfly <u>Hexagenia</u> sp (36 percent of the total), the chironomid <u>Coelotanyopus</u> sp (23 percent), and the asiatic clam <u>Corbicula</u> sp

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(17 percent). The transition zone which had an average number of taxa (14) and had the greatest number of organisms collected (1312 per square meter) among all the Vital Signs transition zones sampled in 1992. <u>Hexagenia</u> sp accounted for 32 percent of the total while Sphaeriidae and Corbicula sp accounted for an additional 19 and 18 percent, respectively. The Chickamauga Reservoir inflow site had the fewest taxa (12) from among the Vital Signs inflow sites sampled in 1992; however, this is an increase at this site over the 1990 and 1991 collections. number of organisms (933 per square meter) also increased The substantially from previous years with 80 percent of the species collected being Corbicula sp. The forebay and transition zone densities slightly increased while the number of taxa remained

Fish Community--Fish data collected in littoral (20 electrofishing transects) and offshore zones (15 net-nights) of Chickamauga forebay resulted in the collection of 37 species (1737 individuals). Emerald shiner was the most abundant species (collected at the rate of 43 per 300 meter transect), accounting for 25 percent of the total number of fish collected. comprised 23 percent of the samples, gizzard shad (17 percent), Bluegill largemouth bass (7 percent), smallmouth bass (6 percent) and spotted bass (4 percent). Due to large numbers of emerald shiners, fish abundance was twice as great in the forebay as the inflow. If emerald shiners are disregarded, the forebay still contained one-third more individuals than the inflow. representative sample could not be collected in the transition zone of Chickamauga Reservoir in fall 1992 due to adverse weather conditions on the day of the survey. Electrofishing RFAI analysis showed a fair quality littoral fish community in both the inflow zone (RFAI=35) and forebay (RFAI=35). Both areas of Chickamauga Reservoir ranked in the middle 30 percentile when compared to mainstream reservoirs. Metrics receiving good rankings for both included percent omnivores, invertivores, and tolerant individuals. Few number of sucker species were present at either location resulting in poor scores for that metric. health of largemouth bass was fair at both the inflow zone The (FHAI=52) and forebay (FHAI=54).

<u>Fish Tissue</u>--There are no fish tissue consumption advisories in effect for Chickamauga Reservoir. Samples for screening studies were conducted in autumn 1991 and 1992. Fillets from five channel catfish were collected from the inflow, transition zone, and forebay, composited by site, and examined for a broad array of analyses (selected metals, pesticides, and PCBs on the EPA Priority Pollutant List). Results from samples collected from all locations in 1991 had low or nondetectable levels of metals and pesticides. PCB concentrations were 0.4, 0.7, and 1.2 μ g/g at the forebay, transition zone, and inflow, respectively. This general trend had been documented in several previous studies but not always as pronounced as in the 1991

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esults. Such was the case for 1992 results - PCB concentrations were 0.6, 0.7, and 0.7 μ g/g at the forebay, transition zone, and forebay, respectively. All other analytes were not detected or found in low concentrations in the 1992 fish samples.

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4.8 <u>Watts Bar Reservoir</u>

4.8.1 Physical Description

Watts Bar Reservoir impounds water from both the Tennessee River and one of the major tributaries to the Tennessee River, the Clinch River. The three dams which bound Watts Bar Reservoir are Watts Bar Dam (located at TRM 529.9), Fort Loudoun Dam (located at TRM 602.3), and Melton Hill Dam located at Clinch River mile (CRM) 23.1. The total length of Watts Bar Reservoir, including the Clinch River arm is 96 miles, the shoreline length is 783 miles, and the surface area is 39,000 acres. The average annual discharge from Watts Bar is approximately 27,800 cfs providing an average hydraulic retention time of about 18 days.

The confluence of the Clinch and Tennessee Rivers is upstream of the transition zone sampling location in Watts Bar, so biological sampling was conducted at the forebay, transition zone, and both the inflow on the Tennessee River and the inflow on the clinch River. Water entering from the Clinch River arm from Melton Hill Reservoir is quite cool due to the hypolimnetic withdrawal from Norris Reservoir (a deep storage impoundment) upstream from Melton Hill. Water entering Watts Bar Reservoir from Fort Loudoun Dam is usually warmer and lower in DO during summer months than water entering from Melton Hill Dam.

A major tributary to the Clinch River arm of Watts Bar Reservoir is the Emory River which supplies on the average about 5 percent of the total flow through Watts Bar Reservoir. The Tennessee and Little Tennessee Rivers (i.e., discharge from Fort Loudoun Dam) account for about 75 percent of the flow and the Clinch River (i.e., discharge from Melton Hill Dam) accounts for about 15 percent through Watts Bar Reservoir.

4.8.2 <u>Reservoir Health</u>

The ecological health of Watts Bar was fair in 1992, same as in 1991. During both years this fair rating was only slightly elow the level considered good. Algae was rated good at both

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The forebay and transition zone locations sampled in 1992. The sediment quality testing at the forebay and the transition zone in 1992 found low survival of test organisms and high concentrations of either ammonia or zinc. In August, concentrations of dissolved oxygen were less than 5 mg/L in the Tennessee River inflow to Watts Bar Reservoir due to the release of water with low DOs from Fort Loudoun Dam. Bottom-dwelling animals rated poor in both 1992 and 1991 at the Tennessee River inflow to Watts Bar Reservoir, possibly related to the low DOs from Fort Loudoun Dam.

Aquatic plants have declined from about 700 acres in the late 1980s to about 10 acres in 1992.

The overall ecological health of the Emory River at the fixed station monitoring site was fair in 1992. The primary problem was with poor sediment quality, evidenced by poor survival of test organisms, suggesting that toxicity may be emanating from active and abandoned coal mines in the watershed.

.8.3 Reservoir Use Suitability

Use Suitability Monitoring activities have not identified any bacteriological problems on Watts Bar Reservoir. The swimming areas at Roane County Park and Riley Creek campground fully support recreation. The informal recreation area near the upper end of Caney Creek embayment partially supports recreation. These evaluations are based on 1990 survey results. Bacteriological sampling in 1992 on Watts Bar Reservoir was limited to midchannel collections in association with Vital Signs Monitoring activities. Fecal coliform bacteria were below levels of detection in all samples.

As a result of PCB contamination, the Tennessee Department of Environment and Conservation (TDEC) has issued advisories on consumption of several fish species from Watts Bar Reservoir. In the Tennessee River portion catfish, striped bass, and striped bass/white bass hybrids should not be eaten. Also a precautionary advisory (children and pregnant or lactating women do not eat ish; all others limit fish consumption to 1.2 pounds per month)

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is in effect for largemouth bass, white bass, sauger, carp and smallmouth buffalo. In the Clinch River arm striped bass should not be eaten and a precautionary advisory is in effect for catfish and sauger.

Also, TDEC has issued a do not eat advisory for fish taken from the East Fork of Popular Creek due to mercury, metals, and organic chemical contamination.

4.8.4 Synopsis of 1992 Conditions

Water--Surface water temperatures ranged from 6.0°C in January to 27.3°C in July in the forebay and from 6.2°C to 26.3°C for these same months at the transition zone. Values for DO at the 1.5-meter depth ranged from 11.6 mg/L in January (as well as 11.6 mg/L in April due to high photosynthetic activity) to 6.3 mg/L in September at the forebay; and, from 11.4 mg/L in January to 5.8 mg/L in September at the transition zone. The minimum observed DO concentration in Watts Bar Reservoir in 1992 was 0.6 mg/L at the bottom of the forebay in July.

Temperature and dissolved oxygen data show that during the summer of 1992, Watts Bar Reservoir developed a moderate degree of both thermal and oxygen stratification in the forebay. Surface to bottom temperature differentials (ΔTs) were 7.0°C in April (during the period of low flows) and exceeded 6°C in May and June. DO versus depth data showed a rather strong oxycline to develop in the forebay of Watts Bar Reservoir from May through August. During these four months surface to bottom differences in DO were consistently greater than 7.0 mg/L, and near bottom DO concentrations in the hypolimnion were less than 1 mg/L in July. The transition zone was much more well mixed during the summer of Maximum ΔTs were 4.1°C (in April) and the minimum bottom DO 1992. measured was 5.5 mg/L (in September).

Values of pH ranged from 6.7 to 9.1 on Watts Bar Reservoir. Throughout the summer (April-August) near surface values of pH in the forebay were often high, exceeding 8.5, with DO saturation values commonly exceeding 100 percent, indicating high rates of photosynthesis.

The average total phosphorus concentrations observed in Watts Bar Reservoir (0.029 mg/L at the forebay and 0.033 mg/L at the transition zone) were among the lowest of the Tennessee River Vital Signs Monitoring locations. In addition, the average dissolved orthophosphorus concentrations of 0.008 mg/L and 0.010 mg/L, respectively, at the forebay and transition zones were also among the lowest observed at any of the Tennessee River Vital Signs Monitoring locations in 1992. The highest chlorophyll-a concentrations were measured in June at the forebay (14 μ g/L) and in May at the transition zone (14 μ g/L). Surface concentrations of chlorophyll-a averaged about 7 μ g/L at the forebay and about 8 μ g/L at the transition zone in 1992. The high TN/TP ratios observed at the transition zone indicate the possibility of phosphorus limitation on primary productivity.

Forebay Secchi depth and suspended solids measurements averaged 1.4 meters and 4.9 mg/L, respectively. These values indicate the light transparency of Watts Bar Reservoir forebay to be relatively high compared with other mainstem Tennessee River reservoirs in 1992.

<u>Fecal Coliform Bacteria</u>--These were no swimming beaches on Watts Bar Reservoir examined as part of this monitoring program in 1992. (The swimming areas at Roane County Park and Riley Creek campground were sampled in 1990, at which time they fully supported water contact recreation. The informal recreation area near the upper end of Caney Creek embayment partially supports recreation, based on 1990 survey results.) Monthly samples collected in midchannel of Watts Bar Reservoir, at the forebay and transition zone as part of the 1992 Vital Signs Monitoring activities, all had concentrations at or less than the detection limit (10 fecal coliform colonies per 100 mL).

ALC: NO.

<u>Sediment</u>--Slightly elevated concentrations of mercury were detected in the sediment of Watts Bar Reservoir in 1992. Concentrations of 0.50 and 0.60 mg/kg were measured in the forebay and transition zone, respectively. The most likely source of this contamination is past operations at Oak Ridge National Laboratory where major environmental cleanup activities are now underway. In addition, elevated sediment zinc concentrations (220 mg/kg) were found in the transition zone, and high concentrations of un-ionized ammonia (470 μ g NH3/L) in sediment pore water were found in the forebay of Watts Bar Reservoir. Sediments were almost entirely silt and clay (97-98 percent) at both the forebay and transition zone.

The toxicological screening of sediment using rotifers (Rotox[®]) and light emitting bacteria (Microtox[®]) in Watts Bar Reservoir in 1992 found indications of toxicity at both locations. Low survival of rotifers (50 percent survival) was found using sediment pore water collected in the forebay of Watts Bar Reservoir, and Microtox[®] tests provided an indication of toxicity in sediment pore water collected at the transition zone.

Benthic Macroinvertebrates--In 1992, the forebay area of Watts Bar Reservoir and the Clinch River inflow had fair benthic communities. The transition zone had a good benthic community, while the Tennessee River inflow had a poor benthic community. he forebay had 19 taxa and 693 organisms/m² which is an increase from 1991. Tubificidae comprised 41 percent of the organisms collected and <u>Chironomus</u> sp 27 percent. The transition zone density (868 organisms/m²) and number of taxa (16) were similar to 1991 with the most numerous taxa being <u>Musculium</u> sp (34 percent) and <u>Hexagenia</u> sp (27 percent). The Tennessee River inflow location had 23 taxa and 547 organisms/m², which was an increase in number of taxa compared to 1991, but similar densities. The dominant taxon was <u>Corbicula</u> sp (62 percent). The Clinch River had 20 taxa and 335 organisms/m² dominated by <u>Corbicula</u> sp (43 percent) and the chironomid <u>Dicrotendipes</u> sp (28 percent).

Aquatic Macrophytes--Aquatic plants have declined from about 700 acres in the late 1980s to an estimated 10 acres in 1992. Eurasian watermilfoil and spinyleaf naiad were the dominant species prior to the recent decline.

<u>Fish Community</u>--Shoreline electrofishing (40 transects) and offshore gill netting (46 net-nights) sampled a total of 4081 fish represented by 41 species. Two species made up the majority of the overall sample: gizzard shad (54 percent) and bluegill (13 percent). These species were followed in abundance by emerald shiners (4 percent), brook silversides (2 percent), and largemouth bass (1 percent). Fish were most abundant in the Clinch River inflow zone (1565) followed by the Tennessee River inflow zone 316), transition zone (769), and forebay (521). Number of taxa resent ranged from 23 in the Clinch River inflow zone to 38 in the Tennessee River inflow zone. FHAI analysis found largemouth bass health to be fair in the forebay (FHAI=53) and transition zone (FHAI=67) and poor in the Tennessee River inflow zone (FHAI=73). No FHAI was possible in the Clinch River inflow zone due to low numbers of largemouth bass collected.

RFAI analysis of shoreline electrofishing data indicated fair littoral fish communities in the two inflow zones (Clinch River Arm RFAI=37, Tennessee River Arm RFAI=37) and the transition (RFAI=31). The forebay fish community was poor (RFAI=27). Compared to respective zones of other mainstream reservoirs, both inflow zones ranked in the upper third, while the forebay and transition zone ranked in the middle 30 percentile. Conditions exhibited in the two inflow stations indicated more species and more diversity in sucker, intolerant, migratory spawning, and lithophilic spawning species than either the transition zone or forebay. Sunfish diversity was rated good in the transition zone, forebay, and Tennessee River inflow. Metrics contributing to the poor forebay designation were a high percentage of tolerant individuals, low fish abundance, and low numbers of sucker, migratory spawning, intolerant, and lithophilic spawning species.

<u>Fish Tissue</u>--Fish from Watts Bar Reservoir have been under ensive investigation for several years because of PCB

TDEC has issued an advisory warning the public not contamination. to eat certain species and to limit consumption of other species. Four of these species (channel catfish, striped bass including striped bass/white bass hybrids, sauger, and largemouth bass) were reexamined in autumn 1991 as part of the continuing study to remain abreast of conditions in this reservoir. These fish were examined individually for PCBs. Average PCB concentrations among sample sites ranged 1.1 to 2.6 μ g/g for channel catfish (eight locations), 0.6 to 2.4 μ g/g for striped bass (three locations), 0.1 to 0.8 $\mu g/g$ for sauger (three locations), and 0.3 to 0.5 $\mu g/g$ for largemouth bass (four locations). (Note: some of the above channel catfish data and all largemouth bass data are part of a Department of Energy study on Watts Bar Reservoir and are still In 1992 three of the above four species considered preliminary.) were reexamined. White bass were examined in 1992, and largemouth bass were not. Average PCB concentrations among sample sites were 0.4 to 1.9 μ g/g for channel catfish (five sites), 1.0 to 1.1 μ g/g for striped bass (two sites), 0.2 to 0.6 μ g/g for sauger (three sites), and the average for white bass at the single location was 0.7 μ g/g. Additional data for channel catfish and striped bass collected in autumn 1992 will be available in the future from the above referenced DOE study.

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Tennessee

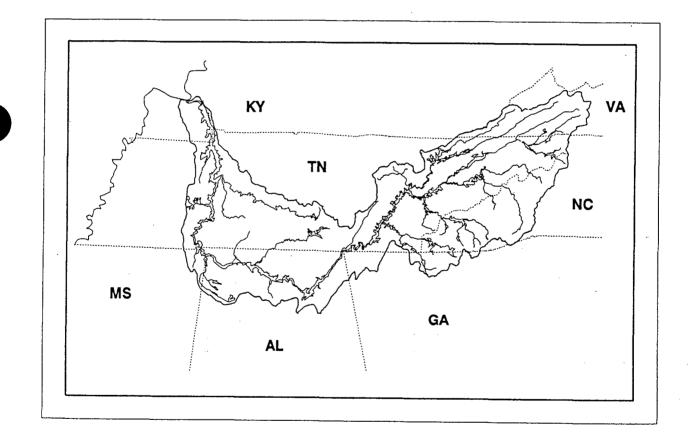
Valley

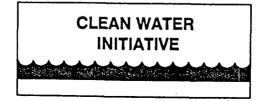
Authority

Water Management Chattanooga, Tennessee May 1994

TENNESSEE VALLEY RESERVOIR AND STREAM QUALITY - 1993 SUMMARY OF VITAL SIGNS AND USE SUITABILITY MONITORING

VOLUME II





Chickamauga Reservoir

Summary of 1993 Conditions - Ecological Health

Water—During the April-September 1993 monitoring period, coolest surface water temperatures in Chickamauga Reservoir were in April and the warmest in July. Surface temperatures ranged from a minimum of 17.0°C to a maximum of 31.7°C at the forebay; from 16.2°C to 30.1°C at the transition zone; and from 19.1°C to 28.8°C in the Hiwassee River embayment. Tennessee's maximum water temperature criteria for the protection of fish and aquatic life is 30.5°C.

Dissolved oxygen (DO) concentrations at the 1.5m depth ranged from a low of 6.9 mg/l in September to a high of 11.4 mg/l in April at the forebay; from 5.7 mg/l in September to 10.3 mg/l in April at the transition zone; and from 7.3 mg/l in August to 9.9 mg/l in April at the sampling location in the Hiwassee River embayment. At the inflow sampling site (i.e., the tailrace of Watts Bar dam) a minimum DO of 3.7 mg/l was recorded in August. Tennessee's minimum dissolved oxygen criteria for the protection of fish and aquatic life is 5.0 mg/l, measured at the 1.5 meter depth.

Temperature data depict seasonal warming and weak thermal stratification in Chickamauga Reservoir from May through July. The maximum observed surface to bottom temperature differentials $(\Delta T's)$, occurred in July. $\Delta T's$ were 5.5°C at the forebay, 3.2°C at the transition zone, and 4.1°C in the Hiwassee River embayment. There was also an oxycline at the forebay and transition zone in June and July when differences between surface and bottom DO's (DO's) were about 6 to 9 mg/l at the forebay and transition zone. In July 1993, a minimum DO of less than 0.1 mg/l was measured on the bottom at the forebay and a minimum of 1.6 mg/l was measured on the bottom at the transition zone. Better DO conditions were observed in the Hiwassee River embayment portion of Chickamauga Reservoir, where maximum DO's were only 1.7 mg/l and near bottom DO's only slightly below 6 mg/l.

DO ratings used in the overall reservoir ecological health evaluation for Chickamauga Reservoir were good at the forebay; good to excellent at the transition zone; excellent in Hiwassee River embayment; and fair at the inflow. The forebay would have rated higher had it not been for the low near bottom oxygen concentrations which existed in July. The fair rating at the inflow sampling site on Chickamauga Reservoir was a result of oxygen levels being measured about 1.5 mg/l below the Tennessee criteria (5 mg/l, at the 1.5 meter depth) in the releases from Watts Bar dam.

Values of pH ranged from 6.8 to 8.8 on Chickamauga Reservoir, in 1993. Near surface pH values exceeding 8.5 (and DO saturation values exceeding 100 percent) were observed on only two occasions (April and July), both at the forebay. Both of these periods of high pH and high oxygen saturations were also coincident with high chlorophyll *a* concentrations, indicative of periods of high photosynthetic activity. Tennessee's maximum pH criteria for the protection of fish and aquatic life is 8.5.

Total nitrogen (TN), total phosphorus (TP), and dissolved ortho phosphorus (DOP) were low in the Tennessee River portion of Chickamauga Reservoir in 1993. TN averaged only 0.37 mg/l at the forebay, the lowest TN concentration measured at any of the Tennessee River sampling sites in 1993. At both the forebay and the transition zone, TP and DOP concentrations averaged only about 0.026 mg/l and 0.005 mg/l, respectively, and were among the lowest TP and DOP concentrations measured at any of the fennessee River sampling sites in 1993. Because of these low concentrations (and because TN/TP ratios often exceeded 20), periods of phosphorus limitation on algal productivity were likely to have occurred.

In 1993, Chickamauga Reservoir chlorophyll a concentrations averaged 8.5 µg/l, 7.8 µg/l, and 5.5 µg/l, respectively, at the forebay, transition zone, and Hiwassee River embayment. Consequently, the chlorophyll a ratings used in the 1993 ecological health evaluation for Chickamauga Reservoir were good (i.e., falling in the 3 to 10 μ g/l range) at all three locations.

Sediment Quality-As in 1990, 1991, and 1992, chemical analyses of sediments from Chickamauga Reservoir in 1993 found high levels of copper (64 mg/kg) and zinc (320 mg/kg) in the forebay. High levels of copper (50 mg/kg) were also found in the Hiwassee River embayment, which was sampled for the first time in 1993. Chlordane was also detected in the forebay (16 μ g/g) and the transition zone (15 µg/g). Toxicity tests indicated no acute toxicity to either species from the three sites tested, but survival of rotifers (75 percent survival) was reduced in the transition zone. Toxicity to rotifers was detected in both forebay and transition zone samples in 1992. Particle size analysis showed sediments from the forebay were 97 percent silt and clay; from the transition zone were 86 percent silt and clay, 14 percent sand; and from the Hiwassee River embayment were 63 percent silt and clay, 37 percent sand.

Sediment quality ratings used in the overall Chickamauga Reservoir ecological health evaluation for 1993 were fair at the forebay (presence of copper, zinc and chlordane); fair at the transition zone (presence of chlordane and reduced survival of rotifers); and, good in the Hiwassee River embayment (presence of copper).

Benthic Macroinvertebrates-The forebay and transition zone sites had excellent benthic communities, and the inflow site was fair. The Hiwassee embayment, a major component of Chickamauga Reservoir, was also included in the ecological health rating. It was shown to support a good benthic community. The forebay site had 19 taxa and 847 organisms/m². The most numerous taxa collected were the chironomid <u>Coelotanypus</u> sp (29 percent), the mayfly <u>Hexagenia limbata</u> (20 percent), the asiatic clam Corbicula fluminea (19 percent) and Tubificidae (17 percent). The transition zone was represented by 25 taxa and 897 organisms/m² with <u>Hexagenia limbata</u> comprising 26 percent of the total organisms and Tubificidae comprising 18 percent of the total organisms. The inflow had 21 taxa and 845 organisms/m². Gammarus fasciatus, an amphipod, was the dominant species present comprising 36 percent of the total organisms. The Hiwassee embayment had the greatest diversity and abundance of organisms than any other site on Chickamauga Reservoir. It had 2312 organisms/m² representing 49 species; Tubificidae were the dominant taxa collected (36 percent) followed by the snail Musculium transversum (17 percent).

The forebay on Chickamauga supported an excellent benthic community, however, the overall benthic score was lowered due to an elevated chironomid community and lowered EPT community. The transition zone also received an excellent rating but fell short of perfect because of an elevated chironomid community and lowered numbers of long-lived taxa. The inflow site rated fair primarily because of an absence of long-lived organisms such as Corbicula sp and Hexagenia sp, and because of reduced diversity and EPT taxa present. The Hiwassee embayment supported a good benthic community in 1993 because of an excellent EPT representation, diversity, low numbers of Chironomids, and evenness of the dominant species. An abundance of tubificids and a lack of long-lived species contributed to this site receiving a good



Aquatic Macrophytes—Coverage of aquatic macrophytes increased from 387 acres in 1992 to 1,185 acres in 1993. Most macrophytes were in Dallas Bay embayment and in small embayments and overbank habitat upstream of TRM 499. Aquatic macrophytes on Chickamauga Reservoir peaked at about 7,500 acres in 1988 and continuously declined until 1993 when coverage increased. Spinyleaf and southern naiad were the dominant species in 1993 although small colonies of Eurasian watermilfoil, American pondweed, and American lotus also were present.

Fish Assemblage—Fish data collected in littoral (45 electrofishing transects) and offshore zones (28 net-nights) of the forebay resulted in the collection of 44 species (6,994 individuals). Emerald shiner was the most abundant species (collected at the rate of 56 per 300 meter electrofishing transect), accounting for 36 percent of the total number of fish collected. Gizzard shad comprised 16 percent of the sample, followed closely by bluegill at 14 percent. Electrofishing results showed approximately twice as many individuals in the inflow (2,624) and transition (2,300) zones as the forebay (1,229), due to numbers of gizzard shad and bluegill in the sample. Numbers of YOY threadfin shad followed a similar pattern with high catch rates in the forebay (CPUE=810 per 300m transect) and transition (CPUE=1,707 per 300m transect) and very high catch rates in the inflow zone (CPUE=3,559 per 300m transect). Gill netting fish abundance was higher in the transition (454) than the forebay (229); although abundance at the inflow zone (158) was lower because of reduced effort, catch rate was similar to the transition zone.

The Reservoir Fish Assemblage Index (RFAI) rated the littoral fish community (based on results of electrofishing samples) fair in the forebay (RFAI=32), good in the transition (RFAI=46), and excellent in the inflow (RFAI=52) zones of Chickamauga Reservoir. The inflow index of 52 was the highest score observed for run-of-the-river reservoir inflows and received maximum scores for all metrics except number of sucker and tolerant species, dominance by a single specie, and percent anomalies. In 1992 the inflow rated only fair (RFAI=34).

The gill netting RFAI rated the transition zone excellent (RFAI=52) and the forebay fair (RFAI=36). The excellent score of 52 in the transition zone was the second highest ever observed for runof-the-river reservoirs and resulted from maximum scores for all metrics except number of sucker, intolerant, and lithophilic spawning species, and percent insectivores. Gill netting RFAI values were not calculated for inflow zones of run-of-the-river reservoirs due to low numbers of replicate samples.

The combined electrofishing and gill netting RFAI score for the transition (RFAI=49) and forebay (RFAI=34) were rated good and fair, respectively. The electrofishing RFAI for the inflow (RFAI=52) zone received an excellent rating, which was one of the highest scores for all inflows sampled in 1993.

Combined fish samples in shoreline electrofishing (15 transects) and offshore gill netting (12 netnights) produced a total of 2263 individuals including 31 species in the Hiwassee River embayment of Chickamauga Reservoir. The three most abundant species were redear sunfish (29 percent), gizzard shad (19 percent), and bluegill (16 percent). There were six times as many fish collected by electrofishing as gill netting, largely attributed to high numbers of sunfishes inhabiting shoreline areas.

The electrofishing RFAI score of 36 rated the embayment community as fair and gill netting results indicated good (RFAI=50) fish community conditions. Combining RFAI scores (RFAI=43) rated the Hiwassee River embayment good (scoring criteria for run-of-the-river transition was used to obtain RFAI ratings). Metrics

for both electrofishing and gill netting that influenced the high scoring included low percent dominance by a single species, low percent omnivores, and high numbers of lithophilic spawning species.

Summary of 1993 Conditions - Use Suitability

Fecal Coliform Bacteria—No bacteriological studies were conducted at recreation sites in Chickamauga Reservoir in 1993. Fecal coliform bacteria concentrations at the monthly Vital Signs locations, the forebay, transition zone, and Hiwassee River Embayment, were all 10/100 ml or less except for one sample. The April sample in the Hiwassee River Embayment had a concentration of 300/100 ml.

Fish Tissue—There are no fish tissue consumption advisories in effect for Chickamauga Reservoir. Samples for screening studies were conducted in autumn 1991 and 1992. Fillets from five channel catfish were collected from the inflow, transition zone, and forebay, composited by site, and examined for a broad array of analyses (selected metals, pesticides, and PCBs on the EPA priority pollutant list). Results from samples collected from all locations in 1991 had low or nondetectable levels of metals and pesticides. PCB concentrations were 0.4, 0.7, and 1.2 μ g/g at the forebay, transition zone, and inflow, respectively. This general trend had been documented in several previous studies but not always as pronounced as in the 1991 results. Such was the case for 1992 results - PCB concentrations were 0.6, 0.7, and 0.7 μ g/g at the forebay, transition zone, and forebay, respectively. All other analytes were not detected or found in low concentrations in the 1992 fish samples.

Watts Bar Reservoir

Summary of 1993 Conditions - Ecological Health

Water—During the April-September 1993 monitoring period, surface water temperatures ranged from a minimum of 18.3° C in April to a maximum of 30.2° C in July in the forebay; and from 16.7° C to 29.8°C (for the same months) at the transition zone. The State of Tennessee's maximum water temperature criteria for the protection of fish and aquatic life is 30.5° C.

Values for DO at the 1.5m depth ranged from a low of 6.5 mg/l in September to a high of 12.6 mg/l in April at the forebay, and from 7.1 mg/l to 11.3 mg/l (for the same months) at the transition zone. At the inflow sampling site on the Tennessee River arm of Watts Bar Reservoir (i.e. the tailrace of Fort Loudoun dam) a minimum DO of 3.9 mg/l was recorded in September. At the inflow sampling site on the Clinch River arm of Watts Bar Reservoir (i.e., the tailrace of Melton Hill dam) a minimum DO of 6.3 mg/l was recorded in March. Tennessee's minimum dissolved oxygen criteria for the protection of fish and aquatic life is 5.0 mg/l, measured at the 1.5 meter depth.

Temperature and dissolved oxygen data show that Watts Bar Reservoir developed a moderate degree of both thermal and oxygen stratification throughout most of the summer of 1993. For the period April through August, monthly surface to bottom temperature differentials (Δ T's) were: 5.2°C, 5.5°C, 7.4° C, 7.3°C, and 4.0°C at the forebay; and 2.3°C, 2.6°C, 3.9°C, 6.2°C, and 2.2°C at the transition zone.

DO versus depth data show that a rather strong oxycline also developed in Watts Bar Reservoir, particularly from June through August. During these three months, surface to bottom differences in DO were: 9.2 mg/l, 9.2 mg/l, and 5.8 mg/l at the forebay; and 7.2 mg/l, 5.8 mg/l, and 3.1 mg/l at the transition zone. At the forebay, near bottom DO concentrations in the hypolimnion were less than 2 mg/l in June and July. In addition, the proportion of the hypolimnion with low DO's (i.e. less than 2 mg/l) averaged about 13 percent of the total cross sectional area, higher than in any other Tennessee River reservoir. The minimum observed DO concentration in Watts Bar Reservoir in 1993 was 0.6 mg/l at the bottom of the forebay in July, but DO's were never less than 4 mg/l at the transition zone.

DO ratings used in the overall reservoir ecological health evaluation for Watts Bar Reservoir were poor at the forebay; excellent at the transition zone and at the inflow sampling site on the Clinch River; and fair at the inflow site on the Tennessee River. The low forebay rating was due to the large proportion of the forebay hypolimnion with low DO concentrations (i.e., less than 2 mg/l). The fair rating at the inflow sampling site on the Tennessee River arm of Watts Bar Reservoir was a result of oxygen levels being measured about 1 mg/l, below the Tennessee criteria (5 mg/l, at the 1.5 meter depth) in the releases from Fort Loudoun dam.

Historically, the pH's of water in Watts Bar Reservoir has been higher than other Tennessee River sampling site. This is due to the addition of the cool, clear, well oxygenated, nitrate rich, and hard water of the Clinch River which combines with the Tennessee River (and Watts Bar Reservoir) at TRM 567.9, about seven miles upstream from the transition zone sampling site. In the summer of 1993, values of pH ranged from 6.8 to 9.0 on Watts Bar Reservoir. During much of the April-September sample period, near surface values of pH frequently exceeded 8.5 at both the forebay and the transition zone, with DO saturation values commonly exceeding 100 percent, indicating high rates of photosynthesis. Tennessee's maximum pH criteria for the protection of fish and aquatic life is 8.5. The average total phosphorus concentrations observed in Watts Bar Reservoir (0.029 mg/l at the forebay and 0.035 mg/l at the transition zone) were among the lowest of the Tennessee River Vital Signs Monitoring locations in 1993. In addition, the average dissolved ortho phosphorus concentrations of 0.007 mg/l and 0.004 mg/l, respectively, at the forebay and transition zones were also among the lowest observed at any of the Tennessee River Vital Signs Monitoring locations in 1993. TN/TP ratios on Watts Bar Reservoir are higher than on any other Tennessee River reservoir. The low phosphorus concentrations in combination with the relatively high nitrogen concentrations (supplied by both the Clinch and Tennessee River inflows) results in the high TN/TP ratios in Watts Bar (particularly at the transition zone) and suggest periods of phosphorus limitation on primary productivity.

The highest chlorophyll *a* concentrations were measured in August at the forebay (10 μ g/l) and in May at the transition zone (11 μ g/l). Surface concentrations of chlorophyll *a* averaged about 7 μ g/l at the forebay and about 8 μ g/l at the transition zone in 1993. Consequently, the chlorophyll *a* ratings used in the 1993 ecological health evaluation for Watts Bar Reservoir were good (i.e., falling in the 3 to 10 μ g/l range) at both locations.

Forebay Secchi depth and suspended solids measurements averaged 1.5 m and 6.3 mg/l, respectively. These values indicate the light transparency of Watts Bar Reservoir forebay to be relatively high compared with other mainstem Tennessee River reservoirs in 1993.

Sediment—Chemical analyses of sediments in Watts Bar Reservoir in 1993 indicated elevated levels of un-ionized ammonia (240 μ g/l) in the forebay, and the presence of chlordane (18 μ g/kg) in the transition zone. Mercury was also detected at the transition zone at a slightly elevated level (0.72 mg/kg), but at a level below sediment quality guidelines for mercury (i.e. 1.0 mg/kg). Toxicity tests detected acute toxicity to daphnids and rotifers (40 percent survival each) in the forebay. The forebay was also toxic to rotifers in 1992. Particle size analysis showed sediments from the forebay were near 100 percent silt and clay; and 98 percent silt and clay from the transition zone.

Sediment quality ratings used in the overall Watts Bar Reservoir ecological health evaluation for 1993 were "poor" at the forebay (acute toxicity to test animals and presence of ammonia); and "good" at the transition zone (presence of chlordane).

Benthic Macroinvertebrates—The forebay site had a good benthic macroinvertebrate community, the transition zone fair, and both the Tennessee River and Clinch River inflow sites had poor benthic communities. The forebay on Watts Bar had 805 organisms/m² representing 18 taxa; the dominant species were the chironomids <u>Chironomus</u> sp (32 percent) and <u>Coelotanypus tricolor</u> (16 percent). The transition zone had 14 taxa and 1,280 organisms/2 with the snail <u>Musculium transversum</u> (34 percent), the mayfly <u>Hexagenia limbata</u> (27 percent) and the chironomid <u>Chironomus</u> sp (17 percent) as the dominant species present. The Tennessee River inflow site had 314 organisms/m² representing 20 taxa; <u>Corbicula fluminea</u> was the dominant species comprising 71 percent of the total organisms. The Clinch River inflow site had 145 organisms/m² made up of 16 taxa; <u>Corbicula fluminea</u> (49 percent), <u>Pseudochironomus</u> sp (18 percent) and Tubificidae (18 percent), were the dominant taxa.

The Watts Bar forebay scored well on all metrics except for the paucity of EPT taxa and the preponderance of chironomids. Those two factors kept this site from obtaining an excellent rating. The

transition zone exhibited a fair community. Reduced diversity, minimal numbers of long-lived species, above average numbers of chironomids, and unevenness associated with the dominant species all contributed to the fair rating this site received. The Tennessee River and Clinch River inflow sites both had a poor benthic communities because of the lack of diversity, EPT taxa, and long-lived species. The unevenness of dominant taxa also negatively impacted these benthic communities. Interestingly, the percent of the total organisms comprised of tubificids and chironomids, normally considered tolerant organisms, was relatively low at both inflows.

Aquatic Macrophytes—Aquatic plants have declined from about 700 acres in the late 1980's to an estimated 10 acres in 1993. Eurasian watermilfoil and spinyleaf naiad were the dominant species prior to the recent decline.

Fish Community—Shoreline electrofishing (60 transects) and offshore gill netting (39 net-nights) sampled a total of 5,174 fish represented by 50 species. Three species made up the majority of the overall sample: gizzard shad (37 percent), bluegill (13 percent), and emerald shiners (12 percent). Electrofishing results showed catch rates to be similar in the Clinch River inflow (CPUE=51 per 300m transect), Tennessee River inflow (CPUE=53 per 300m transect), and forebay (CPUE=56 per 300m transect) but much higher at the transition zone (CPUE=129 per 300m transect). The higher catch rate in the transition was attributed mainly to abundance of emerald shiners and bluegill. Threadfin shad YOY catch rates were moderate in all sample zones except the Tennessee River inflow which was considered high. Gill netting catch rates were much the same in all four sample areas.

The Reservoir Fish Assemblage Index (RFAI) rated the littoral fish community (based on results of electrofishing samples) good in the transition (RFAI=48), fair in the forebay (RFAI=34) and Tennessee River inflow (RFAI=34), and poor in the Clinch River inflow (RFAI=30). The lower Clinch River inflow rating (compared to the Tennessee River inflow) resulted from slightly fewer numbers of sunfish and intolerant species. The gill netting RFAI rated both the transition zone (RFAI=38) and forebay (RFAI=32) fair. Gill netting RFAI values were not calculated for inflow zones of run-of-the-river reservoirs due to low numbers of replicate samples.

Combined electrofishing and gill netting RFAI scores for the forebay (RFAI=33) received a fair rating, followed by the transition (RFAI=43) zones which was rated good. Electrofishing RFAI scores for the Tennessee (RFAI=34) and Clinch River (RFAI=30) inflow zones were rated fair and poor, respectively.

Summary of 1993 Conditions - Use Suitability

Fecal Coliform Bacteria—Fourteen swimming areas were tested for fecal coliform bacteria 12 times each in 1993. Only one sample at each site was collected within 48 hours of a rainfall of at least one-half inch. Bacteria concentrations were generally higher after rainfall. If the one rainfall sample is excluded, all sites met Tennessee's water quality criteria for geometric mean concentration. However, four sites had one or more concentrations to exceed 1000/100 ml, Tennessee's maximum concentration for one sample. Only three of the fourteen areas had very low geometric mean concentrations for all samples (<20/100 ml), a much lower ratio than the other Tennessee River Reservoirs. All monthly fecal coliform bacteria samples taken at the two Vital Signs locations were <10/100 ml.

Fish Tissue—Fish from Watts Bar Reservoir have been under intensive investigation for several years because of PCB contamination. TDEC has issued an advisory warning the public to avoid eating certain species and to limit consumption of other species. Four of these species (channel catfish, striped bass including striped bass/white bass hybrids, sauger, and white bass) were reexamined in autumn in 1992. Average PCB concentrations among sample sites ranged from 0.4 to 1.9 μ g/g for channel catfish (five sites), 1.0 to 1.1 μ g/g for striped bass (two sites), 0.2 to 0.6 μ g/g for sauger (three sites), and the average for white bass at the single location was 0.7 μ g/g. Additional data for channel catfish and striped bass collected in autumn 1992 will be available in the future from studies conducted for DOE study. This is also true for additional fish collected for TVA studies in autumn 1993.

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EROSION/STORM WATER POLLUTION PREVENTION PLAN

ECM Chapter 4 Revision 2 Page 25 of 26

FIGURE 1 Page 1 of 1

STORM WATER DRAINAGE MAP

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