## Appendix E1

# SEM/EDS Data for Test #3, Day-30 Aluminum Coupons

## Figures

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This appendix presents SEM/EDS results for metal aluminum coupons described by two different exposure categories: (1) suspended and (2) submerged. *Suspended* refers to the coupons located above the water level of the solution during the ICET tests. Suspended coupons were contacted by the solution only during the 4-hour spray period at initiation of the test. In addition, the surface of the suspended coupons may be affected by moisture in the test chamber vapor space. *Submerged* refers to coupons that were immersed in the solution for the duration of the test.

The coupon samples were collected on May 5, 2005 (the date Test #3 was shut down), and were later examined by SEM/EDS. The aluminum coupon samples were dried in air before coating with Au/Pd for SEM examination. SEM results present the surface condition of the aluminum coupons. In addition, EDS results provide a semi-quantitative elemental analysis of the coupon surface and the corrosion products. Available logbook entries for this laboratory session are included in this appendix as transcribed notes.

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Laboratory session from May 17, 2005. Test #3, Day-30 Metal Coupons



\*\*Coat with Gold

#### **Suspended Al**

Image:	T3D30AlSusp006	100 ×	SEM image	Figure E1-1
	T3D30AlSusp007	1000 ×	SEM image higher magnification	Figure E1-2
	T3D30AlSusp008	1000 ×	Backscattered image	Figure E1-3
EDS:	T3D30SuspAl05		Particles on 007	Figure E1-4
	T3D30SuspAl06		Surface on 007	Figure E1-5

#### Submerged Aluminum

T3D30AlSubm029	$100 \times$	SEM image	Figure E1-6
T3D30A1Subm030	100 ×	Annotated backscatter SEM	Figure E1-7
T3D30SubmA117		Grey surface on 030	Figure E1-8
T3D30SubmA118		Dark spot on 030	Figure E1-9
T3D30AlSubm031	$100 \times$		Figure E1-10
T3D30AlSubm032	500 ×		Figure E1-11
	T3D30AlSubm029 T3D30AlSubm030 T3D30SubmAl17 T3D30SubmAl18 T3D30AlSubm031 T3D30AlSubm032	T3D30AlSubm029 100 ×   T3D30AlSubm030 100 ×   T3D30SubmAl17 100 ×   T3D30SubmAl18 100 ×   T3D30AlSubm031 100 ×   T3D30AlSubm032 500 ×	T3D30AlSubm029100 ×SEM imageT3D30AlSubm030100 ×Annotated backscatter SEMT3D30SubmAl17Grey surface on 030T3D30SubmAl18Dark spot on 030T3D30AlSubm031100 ×T3D30AlSubm032500 ×



Figure E1-1. SEM image magnified 100 times for a Test #3, Day-30 suspended aluminum coupon. (T3D30AlSusp006)



Figure E1-2. SEM image magnified 1000 times for a Test #3, Day-30 suspended aluminum coupon. (T3D30AlSusp007)



Figure E1-3. Backscattered SEM image magnified 1000 times for a Test #3, Day-30 suspended aluminum coupon. (T3D30AlSusp008)







Figure E1-5. EDS counting spectrum for the surface shown in Figure E1-2. (T3D30SuspAl06)



Figure E1-6.

SEM image magnified 100 times for a Test #3, Day-30 submerged aluminum coupon. (T3D30AlSubm029)



Figure E1-7. Annotated backscattered SEM image magnified 100 times for a Test #3, Day-30 submerged aluminum coupon. (T3D30AlSubm030)





The results from the chemical composition analysis for T3D30SubmA117 are given in Table E1-1.

Table E1-1.	Chemical	<b>Compositions</b> for	T3D30SubmAl17,	, Figure E1-8
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May 17 2005 Group : NRC Sample : T3D30 ID# : 20 Comment : Submerged Al grey surface Condition : Full Scale : 20KeV(10eV/ch,2Kch) Live Time : 60.000 sec Aperture # : 2 Acc. Volt : 15.0 KV Probe Current : 7.746E-09 A Stage Point : X=14.568 Y=53.830 Z=11.000 Acq. Date : Tue May 17 15:13:52 2005 Element Mode ROI (KeV) K-ratio(%) +/-Net/Background οк 0.25- 0.77 3531 / Normal 14.6772 0.0027 18 0.5575 0.0008 385 / Na K Normal 0.81- 1.27 26 0.0017 9.9625 9945 / Al K Normal 1.26- 1.78 200 Si K Normal 1.50- 2.05 9.6207 8956 / 486 Ca K Normal 3.40- 4.30 3.5591 0.0080 1610 / 18 Normal 2.34-3.06 0.4826 0.0007 Cl K 308 / 28 Chi square = 38.5655 Element Mass% Atomic\* ZAF Z Α F 0 39.310 53.7531 1.2511 0.9885 1.2656 1.0000 1.3989 1.2317 1.0429 1.1833 0.9980 Na 1.470 Al 22.707 18.4115 1.0647 1.0041 1.0675 0.9934 Si 27.544 21.4549 1.3373 0.9921 1.3483 0.9998 7.761 4.2363 1.0186 1.0000 1.0185 1.0001 Ca Cl 1.208 0.7453 1.1692 1.0450 1.1209 0.9982 Total 100.000 100.0000 Normalization factor = 2.1408re 2.399 1.5375 0.9896 1.0503 1.0060 0.9366 \_\_\_\_\_ Total 100.000 100.0000 Normalization factor = 3.2486



Figure E1-9. EDS counting spectrum for the dark spot (EDS2) shown in Figure E1-7. (T3D30SubmAl18)

The results from the chemical composition analysis for T3D30SubmA118 are given in Table E1-2.

#### Table E1-2. Chemical Compositions for T3D30SubmAl18, Figure E1-9

May 17 2005 Group : NRC Sample : T3D30 ID# : 21 Comment : Submerged Al dark surface Condition : Full Scale : 20KeV(10eV/ch,2Kch) Live Time : 60.000 sec Aperture # : 2 Acc. Volt : 15.0 KV Probe Current : 7,728E-09 A Stage Point : X=14.568 Y=53.830 Z=11.000 Acq. Date : Tue May 17 15:18:41 2005 Element Mode ROI (KeV) K-ratio(%) +/-Net/Background ОК Normal 0.25- 0.77 27.8853 0.0036 6693 / 39 Al K Normal 1.26- 1.78 0.0022 16.9485 16880 / 180 Si K 1.50- 2.05 Normal 6.0037 0.0008 5576 / 814 Ca K Normal 3.40- 4.30 4.1252 0.0085 1862 / 16 0.81- 1.27 0.09- 0.46 Na K Normal 0.2751 0.0008 190 / 40 СК Normal 0 / 0.0000 0.0000 154 ----- $Chi_square = 70.9798$ Element Massf Atomic\* ZAF Z Α F 0 49.003 63.0911 1.0832 0.9889 1.0954 1.0000 Al 29.742 22.7054 1.0817 1.0048 1.0798 0.9970 Si 13.942 10.2255 1.4315 0.9929 1.4419 0.9999 Ca 6.739 3.4632 1.0069 1.0015 1.0053 1.0001 0.5148 1.2875 1.0436 1.2361 0.9980 Na 0.575 С 0.000 0.0000 6.5476 1.0370 6.3144 1.0000 Total 100.000 100.0000 Normalization factor = 1.6223 re 2.399 1.5375 0.9896 1.0503 1.0060 0.9366 Total 100.000 100.0000 Normalization factor = 3.2486



Figure E1-10. SEM image magnified 100 times for a Test #3, Day-30 submerged aluminum coupon. (T3D30AlSubm031)



Figure E1-11. SEM image magnified 500 times for a Test #3, Day-30 submerged aluminum coupon. (T3D30AlSubm032)

## Appendix E2

## SEM/EDS Data for Test #3, Day-30 Copper Coupons

## Figures

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Figure E2-8.	EDS counting spectrum for the white particles (EDS1) shown in	
· · · ·	Figure E2-7. (T3D30SubmCu03)E2	-8
Figure E2-9.	EDS counting spectrum for the dark surface (EDS2) shown in Figure	
	E2-7. (T3D30SubmCu04)E2	-9

This appendix shows the SEM/EDS results for the metal copper coupons under two different catalogs: (1) suspended and (2) submerged. *Suspended* refers to coupons located above the water level of the solution during ICET tests. Suspended coupons were contacted with the solution only during the 4 hour spraying period at the initial date of the test. In addition, the surface of the suspended coupons may also be affected by the moisture in the test chamber gas space during the test. *Submerged* refers to the coupons that were submerged in the solution during the test.

The coupon samples were collected on May 5, 2005 (the date Test #3 was shut down), and subsequently examined by SEM/EDS. The copper coupon samples were dried in air before being coating with Au/Pd for SEM examination. SEM results present the surface condition of the copper coupons. In addition, EDS results provide a semi-quantitative elemental analysis of the coupon surface and the corrosion products. Available logbook entries for this laboratory session are included in this appendix as transcribed notes.

#### Laboratory session from May 17, 2005. Test #3, Day-30 Metal Coupons



#### Suspended Cu

Image:	T3D30CuSusp001	.100 ×	SEM image	Figure E2-1
	T3D30CuSusp002	1000 ×	Annotated SEM image	Figure E2-2
	T3D30CuSusp003	100 ×	Backscattered image	Figure E2-3
EDS:	T3D30SuspCu01		Medium (convex) dark spot on 002	Figure E2-4
	T3D30SuspCu02		Light spot (concave) on 002	Figure E2-5

### Submerged Cu

Image:	T3D30CuSubm004	100 ×	SEM image	Figure E2-6
	T3D30CuSubm005	$1000 \times$	Annotated SEM image	Figure E2-7
EDS:	T3D30SubmCu03		White spot on 005	Figure E2-8
	T3D30SubmCu04		Dark surface on 005	Figure E2-9



Figure E2-1. SEM image magnified 100 times for a Test #3, Day-30 suspended copper coupon. (T3D30CuSusp001)



Figure E2-2. Annotated SEM image magnified 1000 times for a Test #3, Day-30 suspended copper coupon. (T3D30CuSusp002)



Figure E2-3. Backscattered SEM image magnified 100 times for a Test #3, Day-30 suspended copper coupon. (T3D30CuSusp003)



Figure E2-4. EDS counting spectrum for the convex surface (EDS1) shown in Figure E2-32. (T3D30SuspCu01)



Figure E2-5. EDS counting spectrum for the concave surface (EDS2) shown in Figure E2-32. (T3D30SuspCu02)



Figure E2-6. SEM image magnified 100 times for a Test #3, Day-30 submerged copper coupon. (T3D30CuSubm004)



Figure E2-7. Annotated SEM image magnified 1000 times for a Test #3, Day-30 submerged copper coupon. (T3D30CuSubm005)



Figure E2-8. EDS counting spectrum for the white particles (EDS1) shown in Figure E2-7. (T3D30SubmCu03)



Figure E2-9.

EDS counting spectrum for the dark surface (EDS2) shown in Figure E2-7. (T3D30SubmCu04)

## Appendix E3

## SEM/EDS Data for Test #3, Day-30 Galvanized Steel Coupons

### Figures

Figure E3-1.	SEM image magnified 100 times for a Test #3, Day-30 suspended
	galvanized steel coupon. (T3D30GalSteelSusp012)E3-5
Figure E3-2.	SEM image magnified 1000 times for a Test #3, Day-30 suspended
	galvanized steel coupon. (T3D30GalSteelSusp013)E3-5
Figure E3-3.	Backscattered SEM image magnified 250 times for a Test #3, Day-30
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Figure E3-4.	Annotated backscattered SEM image magnified 250 times for a Test
	#3, Day-30 suspended galvanized steel coupon.
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Figure E3-5.	EDS counting spectrum for the white spot (EDS1) shown in Figure
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Figure E3-6.	Annotated SEM image magnified 100 times for a Test #3, Day-30
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Figure E3-7.	EDS counting spectrum for an egg shaped particle (EDS2) shown in
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Figure E3-8.	EDS counting spectrum for the smooth surface (EDS3) shown in
	Figure E3-6. (T3D30SuspGal12)E3-8
Figure E3-9.	EDS counting spectrum for a porous irregular particle (EDS4) shown
	in Figure E3-6. (T3D30SuspGal11)E3-9
Figure E3-10.	SEM image magnified 100 times for a Test #3, Day-30 submerged
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Figure E3-11.	SEM image magnified 1000 times for a Test #3, Day-30 submerged
	galvanized steel coupon. (T3D30GalSteelSubm010)E3-10

Figure E3-12.	Backscattered SEM image magnified 100 times for a Test #3, Day-30
	submerged galvanized steel coupon. (T3D30GalSteelSubm011)E3-10
Figure E3-13.	Annotated SEM image magnified 100 times for a Test #3, Day-30
	submerged galvanized steel coupon. (T3D30GalSteelSubm009)E3-11
Figure E3-14.	EDS counting spectrum for the white surface (EDS5) shown in Figure
	E3-13. (T3D30SubmGal07)E3-11
Figure E3-15.	EDS counting spectrum for the dark surface (EDS6) shown in Figure
	E3-13. (T3D30SubmGal08)

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### Tables

Table E3-1.	Chemical Compositions for T3D30SubmGal07, Figure E3-14E3-12
Table E3-2.	Chemical Compositions for T3D30SubmGal08, Figure E3-15E3-14

This appendix shows the SEM/EDS results for the metal galvanized steel coupons under two categories: (1) suspended and (2) submerged. Suspended refers to coupons located above the water level of the solution during ICET tests. Suspended coupons were contacted with the solution only during the 4-hour spraying period at the initial date of the test. In addition, the surface of the suspended coupons may also be affected by the moisture in the test chamber gas space during the test. Submerged refers to the coupons that were submerged in the solution during the test.

The coupon samples were collected on May 5, 2005 (the date Test #3 was shut down), and subsequently examined by SEM/EDS. The galvanized steel coupon samples were dried in air before coating with Au/Pd for SEM examination. SEM results present the surface condition of the galvanized steel coupons. In addition, EDS results provide a semi-quantitative elemental analysis of the coupon surface and the corrosion products. Available logbook entries for this laboratory session are included in this appendix as transcribed notes.

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Laboratory session from May 17, 2005. Test #3, Day-30 Metal Coupons



#### \*\*Coat with Gold

#### Suspended Gal-Steel

Image:	T3D30GalSteelSusp012	$100 \times$
	T3D30GalSteelSusp013	$1000 \times$
	T3D30GalSteelSusp014	250 ×
	T3D30GalSteelSusp014	250 ×
EDS:	T3D30SuspGal09	
Image:	T3D30GalSteelSusp012	100 ×
EDS:	T3D30SuspGal10	
	- T3D30SuspGal12	
	T3D30SuspGal11	

	Figure E3-1
	Figure E3-2
Backscattered image	Figure E3-3
Annotated backscatter SEM	Figure E3-4
White spot on 014	Figure E3-5
Annotated SEM image	Figure E3-6
Egg shaped particle on 014	Figure E3-7
Smooth surface on 012	Figure E3-8
Porous irregular shape on 014	Figure E3-9

### Submerged Gal-Steel

Image:	T3D30GalSteelSubm009	$100 \times$
	T3D30GalSteelSubm010	$1000 \times$
	T3D30GalSteelSubm011	$100 \times$
	T3D30GalSteelSubm009	$100 \times$
EDS:	T3D30SubmGal07	
	T3D30SubmGal08	

	Figure E3-11
Backscattered image	Figure E3-12
Annotated SEM image	Figure E3-13
White surface shown in 009	Figure E3-14
Dark surface shown in 009	Figure E3-15

Figure E3-10



Figure E3-1. SEM image magnified 100 times for a Test #3, Day-30 suspended galvanized steel coupon. (T3D30GalSteelSusp012)



Figure E3-2. SEM image magnified 1000 times for a Test #3, Day-30 suspended galvanized steel coupon. (T3D30GalSteelSusp013)



Figure E3-3. Backscattered SEM image magnified 250 times for a Test #3, Day-30 suspended galvanized steel coupon. (T3D30GalSteelSusp014)



Figure E3-4. Annotated backscattered SEM image magnified 250 times for a Test #3, Day-30 suspended galvanized steel coupon. (T3D30GalSteelSusp014)



Figure E3-5.

EDS counting spectrum for the white spot (EDS1) shown in Figure E3-4. (T3D30SuspGal09)



Figure E3-6. Annotated SEM image magnified 100 times for a Test #3, Day-30 suspended galvanized steel coupon. (T3D30GalSteelSusp012)



Figure E3-7. EDS counting spectrum for an egg shaped particle (EDS2) shown in Figure E3-6 (T3D30SuspGal10)



Figure E3-8. EDS counting spectrum for the smooth surface (EDS3) shown in Figure E3-6. (T3D30SuspGal12)


Figure E3-9. EDS counting spectrum for a porous irregular particle (EDS4) shown in Figure E3-6. (T3D30SuspGal11)



Figure E3-10. SEM image magnified 100 times for a Test #3, Day-30 submerged galvanized steel coupon. (T3D30GalSteelSubm009)



Figure E3-11. SEM image magnified 1000 times for a Test #3, Day-30 submerged galvanized steel coupon. (T3D30GalSteelSubm010)



Figure E3-12. Backscattered SEM image magnified 100 times for a Test #3, Day-30 submerged galvanized steel coupon. (T3D30GalSteelSubm011)



Figure E3-13. Annotated SEM image magnified 100 times for a Test #3, Day-30 submerged galvanized steel coupon. (T3D30GalSteelSubm009)



Figure E3-14. EDS counting spectrum for the white surface (EDS5) shown in Figure E3-13. (T3D30SubmGal07)

The results from the chemical composition analysis for T3D30SubmGal07 are given in Table E3-1.

Table E3-1.	Chemical Compositions for T3D30SubmGal07, Figure E3-14
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May 17

2005

Group : NRC Sample : Test3 ID# : 10 Comment : Submerged Galsteel Condition : Full Scale : 20KeV(10eV/ch,2Kch) Live Time : 60.000 sec Aperture # : 2 Acc. Volt : 15.0 KV Probe Current : 7.359E-09 A Stage Point : X=47.136 Y=56.832 Z=10.582 Acq. Date : Tue May 17 12:05:14 2005 Element Mode ROI (KeV) K-ratio(%) +/-Net/Background οк 1466 / Normal 0.25- 0.77 6.4123 0.0018 24 Si K 1.50- 2.05 3.40- 4.30 0.0004 1031 / Normal 1.1657 33 1.1657 0.9216 5.9921 0.0000 Ca K 0.0044 Normal 396 / 16 Zn K Normal 0.0061 0.0000 8.22-10.03 396 / 3 Normal 0.09- 0.46 СК 0 / 42  $Chi_square = 7.3085$ Element Mass% Atomic% ZAF Z Α F 39.582 67.4180 0.9868 0.9165 1.0768 1.0000 0 10.788 10.4670 1.4794 0.9181 1.6115 0.9999 Si Ca 5.422 3.6863 0.9405 0.9224 1.0205 0.9991 Zn 44.208 18.4286 1.1794 1.1810 0.9986 1.0000 С 0.000 0.0000 5.1734 0.9614 5.3811 1.0000 ------Total 100.000 100.0000 Normalization factor = 6.2556С 1.633 4.8654 5.2838 0.9099 5.8071 1.0000 Ρ 2.298 2.6548 1.0399 1.0402 0.9998 0.9999 Fe 2.399 1.5375 0.9896 1.0503 1.0060 0.9366 ~~~~~~~~ Total 100.000 100.0000 Normalization factor = 3.2486



Figure E3-15. EDS counting spectrum for the dark surface (EDS6) shown in Figure E3-13. (T3D30SubmGal08)

The results from the chemical composition analysis for T3D30SubmGal08 are given in Table E3-2.

## Table E3-2. Chemical Compositions for T3D30SubmGal08, Figure E3-15

May 17 2005

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Group Sample Comment Condition	: NRC : Test : Subm : Full Live Acc. Stag Acq.	3 ID# : erged Gai Scale Time Volt e Point Date	11 1steel ( 20KeV : 60.0 : 15.0 ) : X=47. : Tue Ma	dark sun (10eV/cl 00 sec KV 136 Y=50 ay 17 12	cface 1,2Kch) Aper Prob 5.832 Z 2:11:13	ture # e Curre: =10.582 2005	: 2 nt : 7.561E	-09 <b>A</b>
Element	Mode	ROT	(KeV)	K-ratio	o(%) +	/- Ne	et/Backgrour	nd
O K	Norma	1 0.25	- 0.77	6.982	28 0.	0019	1640 /	20
AIK	Norma	1,26	- 1.78	0.061	1 0.	0004	60 /	29
Si K	Norma	1 1.50-	- 2.05	0.475	53 0.	0003	432 /	28
Ca K	Norma	1 3.40	- 4.30	0.874	1 0.	0049	386 /	10
Zn K	Norma	1 8.22-	-10.03	18.584	0.	0084	1260 /	2
СК	Norma	1 0.09-	0.46	0.095	51 0.	0003	33 /	45
РК	Norma	1 1.75	- 2.38	0.680	0.	0018	381 /	36
Fe K	Norma	1 6.00-	7.44	0.746	54 0.	0009	134 /	10
*******			Ch	i_square	= 5.3	2540		
Element M	asst	Atomica	ZAF	z	А	F		
0	21.727	48.5961	0.9578	0.8671	1.1047	0.9999		
Al	0.371	0.4925	1.8718	0.8784	2.1318	0.9996		
Si	2.465	3.1407	1.5964	0.8673	1.8413	0.9996		
Ca	2.557	2.2833	0.9006	0.8692	1.0376	0.9986		
Zn	66.549	36.4297	1.1023	1.1023	1.0000	1.0000		
С	1.633	4.8654	5.2838	0.9099	5.8071	1.0000		
P	2.298	2.6548	1.0399	1.0402	0.9998	0.9999		
Fe	2.399	1.5375	0.9896	1.0503	1.0060	0.9366		
Total 1 Normalizat	00.000 tion fa	100.0000 ctor = 3	3.2486					

## Appendix E4

# SEM/EDS Data for Test #3, Day-30 Steel Coupons

## Figures

Figure E4-1.	SEM image magnified 100 times for a Test #3, Day-30 suspended
	carbon steel coupon. (T3D30SteelSusp033)E4-5
Figure E4-2.	Annotated SEM image magnified 100 times for a Test #3, Day-30
	suspended carbon steel coupon. (T3D30SteelSusp033)E4-5
Figure E4-3.	EDS counting spectrum for the convex surface (EDS1) shown in
	Figure E4-2. (T3D30SuspSteel19)E4-6
Figure E4-4.	EDS counting spectrum for the lower surface (EDS2) shown in Figure
	E4-2. (T3D30SuspSteel21)
Figure E4-5.	Backscattered SEM image magnified 100 times for a Test #3, Day-30
	suspended steel coupon. (T3D30SteelSusp034)E4-7
Figure E4-6.	Annotated backscattered SEM image magnified 100 times for a Test
	#3, Day-30 suspended steel coupon. (T3D30SteelSusp034)E4-7
Figure E4-7.	EDS counting spectrum for a dark particle close to the lower edge
	(EDS3) of Figure E4-6. (T3D30SuspSteel20)E4-8
Figure E4-8.	SEM image magnified 1000 times for a Test #3, Day-30 suspended
	carbon steel coupon. (T3D30SteelSusp035)E4-8
Figure E4-9.	SEM image magnified 100 times for a Test #3, Day-30 submerged
	carbon steel coupon. (T3D30SteelSubm015)E4-9
Figure E4-10.	Backscattered SEM image magnified 100 times for a Test #3, Day-30
	submerged carbon steel coupon. (T3D30SteelSubm016)E4-9
Figure E4-11.	Annotated backscattered SEM image magnified 100 times for a Test
	#3, Day-30 submerged carbon steel coupon. (T3D30SteelSubm016).E4-10
Figure E4-12.	EDS counting spectrum for a dark spot (EDS4) shown in Figure
	E4-11. (T3D30SubmSteel13)

Figure E4-13.	EDS counting spectrum for a light spot (EDS5) shown in Figure
	E4-11. (T3D30SubmSteel14)E4-11
Figure E4-14.	SEM image magnified 1000 times for a Test #3, Day-30 submerged
	carbon steel coupon. (T3D30SteelSubm017)E4-11

E4-ii

This appendix shows the SEM/EDS results for the metal steel coupons under two categories: (1) suspended and (2) submerged. Suspended refers to coupons located above the water level of the solution during ICET tests. Suspended coupons were contacted with the solution only during the 4-hour spraying period at the initial date of the test. In addition, the surface of the suspended coupons may also be affected by the moisture in the test chamber gas space during the test. Submerged refers to the coupons that were submerged in the solution during the test.

The coupon samples were collected on the date that Test #3 was shut down, May 5, 2005. The steel coupon samples were dried in air before coating with Au/Pd for SEM examination. SEM results present the surface condition of the steel coupons. In addition, EDS results provide a semi-quantitative elemental analysis of the coupon surface and the corrosion products. Available logbook entries for this laboratory session are included in this appendix as transcribed notes.

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Laboratory session from May 17, 2005. Test #3, Day-30 Metal Coupons



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#### **Suspended Steel**

Image:	T3D30SteelSusp033	$100 \times$	SEM image
	T3D30SteelSusp033	100 ×	Annotated S
EDS:	T3D30SuspSteel19		Convex on 0
	T3D30SuspSteel21		White flat su
Image:	T3D30SteelSusp034	$100 \times$	Backscattere
	T3D30SteelSusp034	100 ×	Annotated b image
EDS:	T3D30SuspSteel20		Dark particle
Image:	T3D30SteelSusp035	$1000 \times$	

SEM image	Figure E4-1
Annotated SEM image	Figure E4-2
Convex on 033	Figure E4-3
White flat surface on 033	Figure E4-4
Backscattered	Figure E4-5
Annotated backscatter image	Figure E4-6
Dark particle on 034	Figure E4-7
	Figure E4-8

### Submerged Steel

Image:	T3D30SteelSubm015	$100 \times$		Figure E4-9
	T3D30SteelSubm016	100 ×	Backscattered image	Figure E4-10
	T3D30SteelSubm016	100 ×	Annotated backscattered image	Figure E4-11
EDS:	T3D30SubmSteel13		Dark spot on 016	Figure E4-12
	T3D30SubmSteel14		White spot on 016	Figure E4-13
Image:	T3D30SteelSubm017	$1000 \times$		Figure E4-14



Figure E4-1. SEM image magnified 100 times for a Test #3, Day-30 suspended carbon steel coupon. (T3D30SteelSusp033)



Figure E4-2. Annotated SEM image magnified 100 times for a Test #3, Day-30 suspended carbon steel coupon. (T3D30SteelSusp033)



Figure E4-3. EDS counting spectrum for the convex surface (EDS1) shown in Figure E4-2. (T3D30SuspSteel19)



Figure E4-4. EDS counting spectrum for the lower surface (EDS2) shown in Figure E4-2. (T3D30SuspSteel21)



Figure E4-5. Backscattered SEM image magnified 100 times for a Test #3, Day-30 suspended steel coupon. (T3D30SteelSusp034)



Figure E4-6. Annotated backscattered SEM image magnified 100 times for a Test #3, Day-30 suspended steel coupon. (T3D30SteelSusp034)



Figure E4-7. EDS counting spectrum for a dark particle close to the lower edge (EDS3) of Figure E4-6. (T3D30SuspSteel20)



Figure E4-8. SEM image magnified 1000 times for a Test #3, Day-30 suspended carbon steel coupon. (T3D30SteelSusp035)



Figure E4-9. SEM image magnified 100 times for a Test #3, Day-30 submerged carbon steel coupon. (T3D30SteelSubm015)



Figure E4-10. Backscattered SEM image magnified 100 times for a Test #3, Day-30 submerged carbon steel coupon. (T3D30SteelSubm016)



Figure E4-11. Annotated backscattered SEM image magnified 100 times for a Test #3, Day-30 submerged carbon steel coupon. (T3D30SteelSubm016)



Figure E4-12. EDS counting spectrum for a dark spot (EDS4) shown in Figure E4-11. (T3D30SubmSteel13)



Figure E4-13. EDS counting spectrum for a light spot (EDS5) shown in Figure E4-11. (T3D30SubmSteel14)



Figure E4-14. SEM image magnified 1000 times for a Test #3, Day-30 submerged carbon steel coupon. (T3D30SteelSubm017)

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# Appendix F

# SEM/EDS Data for Test #3, Day-30 Flow Meter

### Figures

Figure F-1.	SEM image magnified 80 times for Test #3, Day-30 debris within the
	flow meter. (T3D30FlwMetrDebris005)F-4
Figure F-2.	SEM image magnified 600 times for Test #3, Day-30 debris within
	the flow meter. (T3D30FlwMetrDebris006)F-4
Figure F-3.	EDS counting spectrum for the coatings on the fibers shown in Figure
	F-2. (T3D30FlwDebris03)
Figure F-4.	SEM image magnified 200 times for Test #3, Day-30 deposits on the
	inner wall of the flow meter. (T3~Flowmeter)F-7
Figure F-5.	EDS counting spectrum for the large masses of particulate deposits
	shown in Figure F-4. (T3Deposits08)F-7

### Tables

Table F-1.	Chemical Compositions for T3D30FlwDebris03, Figure F-3.	F-6
Table F-2.	Chemical Compositions for T3Deposits08, Figure F-5.	F-8

In ICET Test #3, significant amounts of debris and precipitates were found within the flow meter. SEM/EDS analysis was performed to examine the composition of the debris trapped in the flow meter, as well as the white precipitates deposited on the inner wall of the flow meter. The debris and the precipitates were collected on May 5, 2005, the date Test #3 was shut down. The samples were dried in air before being coated with Au/Pd for SEM examination. Available logbook entries for this laboratory session are included in this appendix as transcribed notes.

### **Transcribed Laboratory Log**

<u>Laboratory session from May 9, 2005.</u> Test #3, Day-30 Flow Meter



#### **Flow Meter Debris**

Image:	T3D30FlwMetrDebris005	80 ×	Figure F-1
	T3D30FlwMetrDebris006	600 ×	Figure F-2
EDS:	T3D30FlwDebris03		Figure F-3

### **Flow Meter Deposits**

Image:	T3~Flow Meter	$200 \times$	Figure F-4
EDS:	T3Deposits08		Figure F-5



Figure F-1. SEM image magnified 80 times for Test #3, Day-30 debris within the flow meter. (T3D30FlwMetrDebris005)



Figure F-2. SEM image magnified 600 times for Test #3, Day-30 debris within the flow meter. (T3D30FlwMetrDebris006)



Figure F-3. EDS counting spectrum for the coatings on the fibers shown in Figure F-2. (T3D30FlwDebris03)

The results from the chemical composition analysis for T3D30FlwDebris03 are given in Table F-1.

### Table F-1.Chemical Compositions for T3D30FlwDebris03, Figure F-3

May 9 2005

Group Sample Comment Condition	: NRC : T3D30 : Flowme : Full S Live T Acc. V Stage 2 Acq. D	ID# : 3 ter Debris cale : 20Ke ime : 60. olt : 15.0 Point : X=77 ate : Mon 1	V(10eV/ch,2K 000 sec A KV P .422 Y=68.99 May 9 12:10	Cch) perture # Probe Curre 2 Z=12.516 1:19 2005	: 1 ent : 1.596E 5	S-09 A
Element	Mode	ROI (KeV)	K-ratio(%)	+/- N	let/Backgrou	ind
ОК	Normal	0.25- 0.77	1.1291	0.0010	422 7	12
Si K	Normal	1.50- 2.05	0.6220	0.0004	899 <sup>'</sup> /	142
ΡK	Normal	1.75- 2.38	4.6152	0.0040	4117 /	58
Ca K	Normal	3.39- 4.30	12.9947	0.0033	9129 /	16
СК	Normal	0.09- 0.46	0.1025	0.0001	57 /	9
Pd L	Normal	2.22- 3.81	0.5748	0.0010	419 /	35
Al K	Normal	1.19- 1.83	0.0834	0.0002	129 /	24
Na K	Normal	0.81-1.27	0.0806	0.0004	86 /	10
Mg K	Normal	0.97- 1.57	0.0536	0.0001	84 /	15
		Cl	hi_square =	8.9413		
Element M	ass* At	tomic & ZAF	Z	A F		
0	15.028 28	8.4942 2.9323	3 0.9460 3.0	996 1.0000		
Si	3.174	3.4286 1.1244	4 0.9475 1.1	960 0.9922	}	
P	17.384 1	7.0260 0.8298	3 1.1366 0.7	330 0.9960	)	
Ca	57.802 43	3.7487 0.9799	9 0.9630-1.0	176 1.0000		
С	1.833 4	4.6300 3.938	5 0, <b>.9925</b> 3.9	689 0.9999	)	
Pd	3.395 (	0.9679 1.301:	1 1.3345 0.9	949 0.9799		
Al	0.487 (	0.5473 1.285	1 0.9696 1.3	304 0.9962		
Na	0.538 (	$0.7099 \ 1.470$	7 0.9973 1.4	750 0.9999		
Mg	0.359 (	0.4474 1.475	1 0.9384 1.5	740 0.9987	1	
Total 1 Normaliza	00.000 100 tion facto	0.0000 or = 4.5392				

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Figure F-4. SEM image magnified 200 times for Test #3, Day-30 deposits on the inner wall of the flow meter. (T3~Flowmeter)



Figure F-5. EDS counting spectrum for the large masses of particulate deposits shown in Figure F-4. (T3Deposits08)

The results from the chemical composition analysis for T3Deposits08 are given in Table F-2.

#### Table F-2.Chemical Compositions for T3Deposits08, Figure F-5

2005 May 9 : NRC Group : T3D30 ID# : 8 Sample : Flowmeter Deposits Comment Condition : Full Scale : 20KeV(10eV/ch,2Kch) Live Time : 60.000 sec Aperture # : 1 Acc. Volt : 15.0 KV Probe Current : 1.607E-09 A Stage Point : X=47.897 Y=71.447 Z=12.516 : Mon May 9 14:45:11 2005 Acq. Date Element Mode ROI (KeV) K-ratio(%) +/-Net/Background 0.0004 СК Normal 0.09- 0.46 0.5235 292 / 100 4085 / ОК Normal 0.25- 0.77 10.8619 0.0030 54 307 / 0.2843 Na K Normal 0.81- 1.27 0.0009 44 Si K Normal 1.50- 2.05 1.0559 0.0005 1537 / 258 7854 / РК Normal 1.75- 2.38 8.7448 0.0054 123 Ca K Normal 3.39- 4.30 17.8554 0.0039 12630 / 21 ------\_\_\_\_\_ \_ \_ \_ \_ \_ \_ \_ \_ \_ \_ \_ Chi square = 35.5886 Element Mass% Atomic\* ZAF Z Α 7.2812 3.8083 1.0178 3.7418 0.9999 C 3.939 60.3090 2.0251 0.9706 2.0865 1.0000 0 43.460 Na 0.842 0.8134 1.4997 1.0240 1.4645 1.0000 Si 2.376 1.8782 1.1389 0.9739 1.1772 0.9933 14.513 10.4028 0.8400 1.1688 0.7203 0.9976 P Ca 34.870 19.3155 0.9884 0.9928 0.9956 1.0000 100.000 100.0000 Total Normalization factor = 1.9758 100.000 100.0000 Total Normalization factor = 2.1120

# Appendix G

# SEM/EDS and ESEM/EDS Data for Test #3, Day-30 Gel

## Figures

Figure G-1. SEM image magnified 100 times for a Test #3, Day-30 white gel-like				
	material on the top of the birdcage. (T3D30GelMaterial003)G-5			
Figure G-2.	SEM image magnified 1000 times for a Test #3, Day-30 white gel-like			
	material on the top of the birdcage. (T3D30GelMaterial004) G-5			
Figure G-3.	EDS counting spectrum for the white gel-like material (whole image)			
	shown in Figure G-2. (T3D30Gel02)G-6			
Figure G-4.	ESEM image magnified 1000 times for a Test #3, Day-30 white gel-like			
	material on the top of the birdcage. (t3Gel08)G-8			
Figure G-5.	EDS counting spectrum for the white gel-like material shown in Figure			
	G-4. (t3GelED4)			
Figure G-6.	Comparison of EDS counting spectra for Figure G-5 (red, the gel-like			
	restaurate above in Figure C.4) and Figure C4.5 (vallage the large deposite			
	materials shown in Figure G-4) and Figure C4-5 (yenow, the large deposits			
	taken from the birdcage exterior shown in Figure C4-4). (t3geled5) G-9			
Figure G-7.	taken from the birdcage exterior shown in Figure C4-4). (t3geled5) G-9 Another EDS counting spectrum for the white gel-like material shown in			
Figure G-7.	taken from the birdcage exterior shown in Figure C4-3. (t3geled5) G-9 Another EDS counting spectrum for the white gel-like material shown in Figure G-4. (t3geled6)			
Figure G-7. Figure G-8.	taken from the birdcage exterior shown in Figure C4-3 (yellow, the large deposits taken from the birdcage exterior shown in Figure C4-4). (t3geled5) G-9 Another EDS counting spectrum for the white gel-like material shown in Figure G-4. (t3geled6)			
Figure G-7. Figure G-8.	taken from the birdcage exterior shown in Figure C4-3 (yellow, the large deposits taken from the birdcage exterior shown in Figure C4-4). (t3geled5) G-9 Another EDS counting spectrum for the white gel-like material shown in Figure G-4. (t3geled6)			
Figure G-7. Figure G-8.	materials shown in Figure G-4) and Figure C4-3 (yellow, the large deposits taken from the birdcage exterior shown in Figure C4-4). (t3geled5) G-9 Another EDS counting spectrum for the white gel-like material shown in Figure G-4. (t3geled6) G-9 Comparison of EDS counting spectra for Figure G-7 (yellow, the gel-like materials shown in Figure G-4) and Figure C4-5 (red, the large deposits taken from the birdcage exterior shown in Figure C4-4). (t3geled7) G-10			

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# Tables

Table G-1.	Chemical Compositions for T3D30Gel02, Figure G-3G-	.7
Table G-2.	Dry Mass Composition of a Test #3, Day 30 White Gel-Like Sample by XRF	
	Analysis	1

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In ICET Test #3, one distinguished phenomenon is the presence of white gel-like precipitates in the testing solution. On the shutdown date of Test #3, deposits of the white gel-like precipitates were observed on the top of the birdcage. These precipitates may increase the containment sump screen head loss during a LOCA; therefore, it is necessary to investigate the morphology and composition of the white gel-like precipitate.

This appendix shows the ESEM/SEM/EDS and XRD/XRF results of the white gel-like precipitates. The precipitates were collected on the date Test #3 was shut down (May 5, 2005). For the SEM examination, the samples were dried in air before being coated with Au/Pd. EDS results provide a semi-quantitative elemental analysis of the sample compositions. Also, XRD results show the crystal structure of the white gel-like precipitates. Based on the XRD results, the composition of the white gel-like precipitates contained crystalline substances of sodium calcium hydrogen carbonate phosphate hydrate [Ca<sub>8</sub>H<sub>2</sub>(PO<sub>4</sub>)<sub>6</sub>·H<sub>2</sub>O·Li<sub>2</sub>CO<sub>3</sub>·H<sub>2</sub>O]. In addition, XRF results indicate the chemical composition of the precipitates. Available logbook entries for this laboratory session are included in this appendix as transcribed notes.

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### Transcribed Laboratory Log

#### Laboratory session from May 9, 2005. Test #3, Day-30 Gel Material



#### Gel Material

Image:	T3D30GelMaterial003	100 ×		Figure G-1
	T3D30GelMaterial004	$1000 \times$		Figure G-2
EDS:	T3D30Gel02		Whole screen of image 004	Figure G-3

Laboratory session from May 6, 2005. Test #3, Day-30 Gel Material



### Gel-Like Material on Top of Birdcage

Image:	t3Gel08	$1000 \times$		Figure G-4
EDS:	t3GelED4		White gel has high C and low- Si	Figure G-5
	t3geled5		Comparing t3bcexe2 from Appendix C4 & t3GelED4	Figure G-6
	t3geled6		EDS of gel material	Figure G-7
	t3geled7		Comparing t3bcexe2 from Appendix C4 & EDS6	Figure G-8


Figure G-1. SEM image magnified 100 times for a Test #3, Day-30 white gel-like material on the top of the birdcage. (T3D30GelMaterial003)



Figure G-2. SEM image magnified 1000 times for a Test #3, Day-30 white gel-like material on the top of the birdcage. (T3D30GelMaterial004)



Figure G-3. EDS counting spectrum for the white gel-like material (whole image) shown in Figure G-2. (T3D30Gel02)

The results from the chemical composition analysis for T3D30Gel02 are given in Table G-1.

 Table G-1.
 Chemical Compositions for T3D30Gel02, Figure G-3

May 9 2005 : NRC Group : T3D30 ID# : 2 Sample Comment : GelMaterial Condition : Full Scale : 20KeV(10eV/ch,2Kch) : 60.000 sec Aperture # : 15.0 KV Probe Curre Live Time : 1 Acc. Volt Probe Current : 1.606E-09 A Stage Point : X=79.625 Y=59.260 Z=11.424 Acg. Date : Mon May 9 11:42:11 2005 Element Mode ROI (KeV) K-ratio(%) +/-Net/Background CK Normal 0.09- 0.46 0.6057 0.0005 338 7 119 ОК Normal 0.25- 0.77 12.2043 0.0032 4587 / 68 Na K Normal 0.81- 1.27 0.5675 0.0010 613 / 50 Si K Normal 1.50- 2.05 0.9391 0.0005 1366 / 271 ΡK Normal 1.75- 2.38 8.4975 0.0055 7628 / 107 Ca K Normal 3.39- 4.30 17.1295 0.0038 12109 / 26 ------\_ \_ \_ . Chi square = 42.7915Element Mass% Atomic\* ZAF Z Α С 4.355 7.8616 3.7318 1.0194 3.6611 0.9999 0 45.521 61.6928 1.9361 0.9721 1.9917 1.0000 Na 1.639 1.5456 1.4989 1.0256 1.4614 1.0000 Si 2.072 1.5994 1.1451 0.9756 1.1812 0.9937 P 13.776 9.6435 0.8415 1.1708 0.7203 0.9978 32.638 17.6571 0.9890 0.9947 0.9943 1.0000 Ca Total 100.000 100.0000 Normalization factor = 1.9265 TOLAL T00.000 T00.0000 Normalization factor = 2.1120



Figure G-4. ESEM image magnified 1000 times for a Test #3, Day-30 white gel-like material on the top of the birdcage. (t3Gel08)



Figure G-5. EDS counting spectrum for the white gel-like material shown in Figure G-4. (t3GelED4)



Figure G-6. Comparison of EDS counting spectra for Figure G-5 (red, the gel-like materials shown in Figure G-4) and Figure C4-5 (yellow, the large deposits taken from the birdcage exterior shown in Figure C4-4). (t3geled5)



Figure G-7. Another EDS counting spectrum for the white gel-like material shown in Figure G-4. (t3geled6)



Figure G-8. Comparison of EDS counting spectra for Figure G-7 (yellow, the gel-like materials shown in Figure G-4) and Figure C4-5 (red, the large deposits taken from the birdcage exterior shown in Figure C4-4). (t3geled7)



Figure G-9.

XRD results for Test #3, Day-30 white gel-like material.

Sample ID	SiO <sub>2</sub>	TiO₂	Al <sub>2</sub> O <sub>3</sub>	Fe <sub>2</sub> O <sub>3</sub>	FeO	MnO	MgO	CaO	Na <sub>2</sub> O	K <sub>2</sub> O	H20(- )	H <sub>2</sub> O(+) CO <sub>2</sub>	P <sub>2</sub> O <sub>5</sub>	Total	H2O(+)CO2 /DF (10) & Cover. To %
Test #3,			ĺ												
Day 30				1											
White															
Gel	5.26	0.02	0.63	0.07	0.00	0.00	0.25	35.01	2.39	0.06	4.75	19.24	27.09	94.77	1.0196

Table G-2.Dry Mass Composition of a Test #3, Day 30 White Gel-Like Sample<br/>by XRF Analysis

# Appendix H

# SEM/EDS and ESEM/EDS Data for Test #3, Day-30 Cal-Sil

## Figures

Figure H-1.	SEM image magnified 100 times for a raw cal-sil sample.
	(T3~RawCal~Sil007)H-6
Figure H-2.	SEM image magnified 1000 times for a raw cal-sil sample.
;	(T3~RawCal~Sil008)H-6
Figure H-3.	Annotated SEM image magnified 1000 times for a raw cal-sil sample.
	(T3~RawCal~Sil008)H-7
Figure H-4.	EDS counting spectrum for the particles (EDS1) shown in Figure H-3.
	(T3Prtcle04)H-7
Figure H-5.	EDS counting spectrum for the fiber (EDS2) shown in Figure H-3.
	(T3fiber05)H-9
Figure H-6.	EDS counting spectrum for the whole image shown in Figure H-3.
	(EDS06)
Figure H-7.	SEM image magnified 100 times for a raw cal-sil sample.
	(T3~RawCal~Sil009)
Figure H-8.	SEM image magnified 200 times for a baked cal-sil sample
	(T3~BakedCal~Sil010)H-13
Figure H-9.	EDS counting spectrum for the baked cal-sil sample (whole image)
	shown in Figure H-7. (EDS07) H-14
Figure H-10.	SEM image magnified 1000 times for a baked cal-sil sample.
	(T3~BakedCal~Sil011)H-16
Figure H-11.	ESEM image magnified 100 times for the exterior of a Test#3, Day-
	30 raw cal-sil sample submerged in the birdcage. (t3calx21) H-16
Figure H-12.	ESEM image magnified 500 times for the exterior of a Test#3, Day-
	30 raw cal-sil sample submerged in the birdcage. (T3Calx22)

Figure H-13.	Annotated ESEM image magnified 500 times for the exterior of a
	Test#3, Day-30 raw cal-sil sample submerged in the birdcage.
	(T3Calx22)
Figure H-14.	EDS counting spectrum for the particles shown in Figure H-13.
	(t3calx23)
Figure H-15.	ESEM image magnified 100 times for the interior of a Test#3, Day-30
	raw cal-sil sample submerged in the birdcage. (T3CalI24) H-18
Figure H-16.	ESEM image magnified 500 times for the interior of a Test#3, Day-30
	raw cal-sil sample submerged in the birdcage. (T3Call25)
Figure H-17.	Annotated ESEM image magnified 500 times for the interior of a
	Test#3, Day-30 raw cal-sil sample submerged in the birdcage.
	(T3CalI25)
Figure H-18.	EDS counting spectrum for the particles shown in Figure H-17.
	(T3cali26)
Figure H-19.	ESEM image magnified 500 times for the interior of a Test#3, Day-30
	raw cal-sil sample submerged in the birdcage. (T3Call27) H-20
Figure H-20.	ESEM image magnified 100 times for the exterior of a Test#3, Day-
	30 submerged high-flow baked cal-sil sample. (T3bcal39)
Figure H-21.	ESEM image magnified 1000 times for the exterior of a Test#3, Day-
	30 submerged high-flow baked cal-sil sample. (t3bcal40) H-21
Figure H-22.	Annotated ESEM image magnified 1000 times for the exterior of a
	Test#3, Day-30 submerged high-flow baked cal-sil sample.
·	(t3bcal40)
Figure H-23.	EDS counting spectrum for the light particles (EDS1) shown in Figure
	H-22. (T3bcal41)
Figure H-24.	EDS counting spectrum for the dark particles (EDS2) shown in Figure
	H-22. (t3bcal42)
Figure H-25.	ESEM image magnified 100 times for the interior of a Test#3, Day-30
	submerged high-flow baked cal-sil sample. (T3Bcal43) H-23
Figure H-26.	ESEM image magnified 1000 times for the interior of a Test#3, Day-
	30 submerged high-flow baked cal-sil sample. (t3bcal44)

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Figure H-27.	ESEM image magnified 1000-times for the interior of a Test#3, Day-			
	30 submerged high-flow baked cal-sil sample. (t3bcal45) H-2	4		
Figure H-28.	EDS counting spectrum for the whole image shown in Figure H-27.			
	(t3bcal46)H-2	5		
Figure H-29.	XRD results for the unused raw cal-sil sample	6		
Figure H-30.	XRD results for the unused baked cal-sil sample	7		

# Tables

Table H-1.	Chemical Compositions for T3Prtcle04, Figure H-4	H-8
Table H-2.	Chemical Compositions for T3fiber05, Figure H-5 H	-10
Table H-3.	Chemical Compositions for EDS06, Figure H-6H	-12
Table H-4.	Chemical Compositions for EDS07, Figure H-9 H	-15
Table H-5.	Dry Mass Composition of Unused Cal Sil Samples by XRF Analysis H	-27

ICET #3 is the first ICET that incorporates calcium silicate (cal-sil) to represent insulation materials, along with fiberglass, in a power plant. This appendix presents the ESEM/SEM/EDS and XRD/XRF results of different cal-sil samples, including unused raw and unused baked cal-sil samples. The Test #3, Day-30 raw cal-sil samples were submerged in the birdcage, whereas the Test #3, Day-30 baked cal-sil sample were submerged in the high-flow zone.

The submerged cal-sil samples were collected on the date Test #3 was shut down (May 5, 2005). For the SEM examination, the samples were dried in air before being coated with Au/Pd. EDS results provide a semi-quantitative elemental analysis of the sample compositions.

In addition, XRD/XRF results show the crystal structure and the chemical composition of the unused raw and unused baked cal-sil samples. Based on XRD results, both unused raw and unused baked cal-sil samples contained crystalline substances of tobermorite  $[Ca_{2.25}(Si_3O_{7.5}(OH)_{1.5})(H_2O)]$  and calcite (CaCO<sub>3</sub>). Available logbook entries for this laboratory session are included in this appendix as transcribed notes.

#### Transcribed Laboratory Log

<u>Laboratory session from May 9, 2005.</u> Test #3, Day-30 Cal-Sil



#### **Raw Cal-Sil**

Image:	T3~RawCal-Sil007	$100 \times$		Figure H-1
	T3~RawCal-Sil008	1000 ×		Figure H-2
EDS:	T3Prtcle04		On particles of image 008	Figure H-3
	T3fiber05		On fiber of image 008	Figure H-4
	EDS06		Whole image of image 008	Figure H-5
Image:	T3~RawCal-Sil009	100 ×		Figure H-6

#### **Baked Cal-Sil**

Image:	T3~BakedCal-Sil010	$200 \times$		Figure H-7
EDS:	EDS07		EDS of image 010	Figure H-8
Image:	T3~BakedCal-Sil011	1000 ×		Figure H-9

### **Transcribed Laboratory Log**

Laboratory session from May 10, 2005. Test #3, Day-30 ESEM Birdcage.



#### Cal-Sil Exterior in Birdcage

Image:	t3calx21	500 ×		Figure H-10
	T3Calx22	500 ×		Figure H-11
EDS:	t3calx23		EDS on T3Calx22	Figure H-12

### Cal-Sil Interior in Birdcage

Image:	T3Call24	100 ×		Figure H-13
	T3Call25	500 ×		Figure H-14
EDS:	T3cali26		EDS on T3Call25	Figure H-15
Image:	t3cali27	500 ×		Figure H-16

### Transcribed Laboratory Log

Laboratory session from May 11, 2005. Test #3, Day-30 Baked Cal-Sil



#### Submerged Baked Cal-Sil Exterior

Image:	T3bcal39	100 ×		Figure H-17
	t3bcal40	1000 ×	Particles	Figure H-18
EDS	t3bcal41		EDS on white particles shown in t3bcal40	Figure H-19
	t3bcal42		EDS on dark particles shown in t3bcal40	Figure H-20
Image:	T3Bcal43	100 ×		Figure H-21
	t3bcal44	1000 ×		Figure H-22
	t3bcal45	1000 ×		Figure H-23
EDS:	t3bcal46		EDS on whole screen of image 45	Figure H-24



Figure H-1. SEM image magnified 100 times for a raw cal-sil sample. (T3~RawCal~Sil007)



Figure H-2. SEM image magnified 1000 times for a raw cal-sil sample. (T3~RawCal~Sil008)



Figure H-3. Annotated SEM image magnified 1000 times for a raw cal-sil sample. (T3~RawCal~Sil008)



Figure H-4. EDS counting spectrum for the particles (EDS1) shown in Figure H-3. (T3Prtcle04)

The results from the chemical composition analysis for T3Prtcle04 are given in Table H-1.

 Table H-1.
 Chemical Compositions for T3Prtcle04, Figure H-4

2005 May 9 Group : NRC Sample : T3D30 ID# : 4 : Raw Cal-Sil Particle Comment Condition : Full Scale : 20KeV(10eV/ch,2Kch) : 60.000 вес Aperture # : 1 Live Time : 15.0 KV Acc. Volt Probe Current : 1.604E-09 A Stage Point : X=58.349 Y=61.668 Z=12.516 Acq. Date : Mon May 9 14:05:35 2005 Element Mode ROI (KeV) K-ratio(%) +/-Net/Background CK Normal 0.09- 0.46 0.9637 0.0005 537 / 119 10.7009 0.0031 OK Normal 0.25- 0.77 4017 / 65 1.1322 Na K Normal 0.81- 1.27 0.0012 1221 / 43 Mg K Normal 0.97- 1.57 0.1062 0.0003 166 / 80 AĨ K Normal 1.19- 1.83 0.0004 0.9046 1409 / 80 Si K Normal 1.50- 2.05 6.6718 0.0010 9692 / 107 Ca K Normal 3.39- 4.30 13.6411 0.0036 9631 / 25 2.22- 3.81 1.1981 Pd L Normal 0.0013 878 / 68 Fe K Normal 6.00-7.44 0.8951 0.0012 257 / 18 -----------------Chi square = 21.7880Element Mass\* Atomic\* ZAF Z Α 13.1026 3.7872 1.0155 3.7297 0.9999 7.204 С 38.993 53.2408 1.8460 0.9683 1.9064 1.0000 0 3.1581 1.4872 1.0216 1.4562 0.9997 3.324 Na 0.2898 1.5385 0.9616 1.6029 0.9982 0.323 Mg 2.373 1.9210 1.3288 0.9940 1.3428 0.9956 Al Si 15.630 12.1570 1.1868 0.9716 1.2226 0.9991 Ca 26.877 14.6492 0.9982 0.9904 1.0081 0.9997 Pd 0.6437 1.3257 1.3709 0.9776 0.9891 3.135 Fe 2.142 0.8379 1.2123 1.1945 1.0148 1.0000 \_\_\_\_\_ 100.000 100.0000 Total

Normalization factor = 1.9740



Figure H-5. EDS counting spectrum for the fiber (EDS2) shown in Figure H-3. (T3fiber05)

The results from the chemical composition analysis for T3fiber05 are given in Table H-2.

May 9	2005					
Group Sample Comment Conditio	: NRC : T3D30 : Raw Cal n : Full Sc Live Ti Acc. Vo Stage P Acq. Da	ID# : 5 -Sil Fiber ale : 20KeV me : 60.0 lt : 15.0 1 oint : X=58.3 te : Mon Ma	(10eV/ch,2K 00 sec A KV P 273 Y=61.61 ay 9 13:56	ch) perture # robe Curre 7 Z=12.516 :08 2005	: 1 ent : 1.603E	-09 A
Element C K O K Na K Al K Si K Ca K	Mode Normal Normal Normal Normal Normal	ROI(KeV) 0.09- 0.46 0.25- 0.77 0.81- 1.27 1.19- 1.83 1.50- 2.05 3.39- 4.30	K-ratio(%) 33.8965 10.3921 3.1096 0.1520 1.1022 2.0189	+/- N 0.0016 0.0032 0.0017 0.0003 0.0005 0.0016	Met/Backgrou 18866 / 3899 / 3352 / 237 / 1600 / 1425 /	nd 99 1192 52 64 74 17
		Ch	i_square =	50.8203		
Element ! C O Na Al Si Ca	Mass% At 65.848 74 25.742 21 4.419 2 0.219 0 1.434 0 2.338 0	omic% ZAF .0755 1.6099 .7397 2.0528 .5969 1.1775 .1098 1.1954 .6899 1.0783 .7882 0.9598	Z 1.0143 1.5 0.9679 2.1 1.0228 1.1 0.9966 1.2 0.9750 1.1 0.9997 0.9	A F 873 1.0000 208 1.0000 510 1.0002 002 0.9994 061 0.9999 601 1.0000		
Total : Normaliza	100.000 100 ation facto:	.0000 r = 1.2067	· · · · · · · · · · · · · · · · · · ·			

#### Chemical Compositions for T3fiber05, Figure H-5 Table H-2.

May



Figure H-6. EDS counting spectrum for the whole image shown in Figure H-3. (EDS06)

The results from the chemical composition analysis for EDS06 are given in Table H-3.

Group Sample Comment Condition	: NRC : T3D30 : Raw Cal h : Full So Live Ti Acc. Vo Stage F Acq. Da	ID# : 6 1-Sil cale : 20KeV ime : 60.0 olt : 15.0 Point : X=57 ate : Mon M	/(10eV/ch,2H )00 sec / KV F .876 Y=61.13 May 9 14:28	Kch) Aperture # Probe Curr 38 Z=12.51 3:12 2005	: 1 ent : 1.606E 6	-09 A
Floment	Nodo	POT (KoW)	K-ratio (8)	/	Net /Packgrou	nd
C K FIEWEUC	Normal	0.09 - 0.46	1 1389		635 /	112
O K	Normal	0 25- 0 77	11 5668	0.0033	4348 /	73
Na K	Normal	0.81- 1.27	1.5352	0.0013	1658 /	56
Mg K	Normal	0.97-1.57	0.1356	0.0003	213 /	79
AĬ K	Normal	1.19- 1.83	1.1066	0.0005	1726 /	98
Si K	Normal	1.50- 2.05	7.1883	0.0010	10455 /	146
Ca K	Normal	3.39- 4.30	12.0212	0.0034	8498 /	24
Fe K	Normal	6.00- 7.44	0.9544	0.0012	274 /	11
	*******	Ch	i_square =	26.8019		·
Element M	lasst At	omict ZAF	Z	A F		
С	9.300 16	.0597 4.1007	1.0233 4.0	075 0.9999	Ð	
0	39.825 51	.6305 1.7291	0.9759 1.7	719 1.0000	)	
Na	4.382 3	.9538 1.4336	1.0297 1.3	927 0.9996	5	
Mg	0.411 0	.3503 1.5204	0.9694 1.5	717 0.9979	9	
Al	2.909 2	.2365 1.3203	1.0021 1.3	240 0.9952	2	•
Si	17.041 12	.5848 1.1905	0.9797 1.2	160 0.9994		
Ca	23.814 12	.3235 0.9948	0.9993 0.9	959 0.9996	5	
Fe	2.318 0	.8608 1.2196	1.2068 1.0	106 1.0000	J 	
Total 1 Normaliza	.00.000 100 tion facto	0.0000 or = 1.9913			. –	

### Table H-3. Chemical Compositions for EDS06, Figure H-6

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Figure H-7. SEM image magnified 100 times for a raw cal-sil sample. (T3~RawCal~Sil009)



Figure H-8. SEM image magnified 200 times for a baked cal-sil sample (T3~BakedCal~Sil010)



Figure H-9. EDS counting spectrum for the baked cal-sil sample (whole image) shown in Figure H-7. (EDS07)

The results from the chemical composition analysis for EDS07 are given in Table H-4.

Group : NRC Sample : T3D30 ID# : 7 Comment : Baked Cal-Sil Condition : Full Scale : 20KeV(10eV/ch,2Kch) Live Time : 60.000 sec Aperture # : 1 Acc. Volt : 15.0 KV Probe Current : 1.604E-09 A Stage Point : X=49.364 Y=57.751 Z=12.516 Acq. Date : Mon May 9 14:20:15 2005						
Flowert	Mode	POT (KoV)	K-ratio (8)		Int /Packaroun	4
C K FIGWEIIC	Normal	0.09 - 0.46	N-14010(%)	, -,	516 /	้าธา
O K	Normal	0.25 - 0.77	14.9326	0.0037	5606 /	77
Na K	Normal	0.81- 1.27	1.0025	0.0012	1081 /	65
Mar K	Normal	0.97-1.57	0.1609	0.0003	252 /	79
AI K	Normal	1.19- 1.83	1.4816	0.0005	2308 /	124
Si K	Normal	1.50- 2.05	9.5017	0.0012	13802 /	172
Ca K	Normal	3.39- 4.30	15.8398	0.0038	11184 /	30
Fe K	Normal	6.00- 7.44	0.7928	0.0013	227 /	10
Chi_square = 33.5587						
Element Ma	.ss% At	omic% ZAF	Z	A F		
С	6.304 11	.1172 4.1881	L 1.0251 4.0	0858 0.9999	)	
04	2.250 55	.9339 1.7404	0.9776 1.7	7803 1.0000	)	
Na	2.361 2	.1752 1.448	7 1.0315 1.4	4051 0.9996	5	
Mg	0.391 0	.3407 1.4950	0.9710 1.9	5432 0.9977	1	
Al	3.143 2	.4673 1.3049	9 1.0037 1.3	3069 0.9947	1	
Si 1	8.294 13	.7966 1.1843	3 0.9812 1.2	2077 0.9994	<b>k</b>	
Ca 2	5.681 13	.5714 0.9973	3 1.0007 0.9	9968 0.9998	\$	
re	1.576 0	.5977 1.2226	5 1.2082 1.0	0119 1.0000	)	
Total 100.000 100.0000						
Normalization factor = 1.6257						

### Table H-4. Chemical Compositions for EDS07, Figure H-9

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Figure H-10. SEM image magnified 1000 times for a baked cal-sil sample. (T3~BakedCal~Sil011)



Figure H-11. ESEM image magnified 100 times for the exterior of a Test#3, Day-30 raw cal-sil sample submerged in the birdcage. (t3calx21)



Figure H-12. ESEM image magnified 500 times for the exterior of a Test#3, Day-30 raw cal-sil sample submerged in the birdcage. (T3Calx22)



Figure H-13. Annotated ESEM image magnified 500 times for the exterior of a Test#3, Day-30 raw cal-sil sample submerged in the birdcage. (T3Calx22)



Figure H-14. EDS counting spectrum for the particles shown in Figure H-13. (t3calx23)



Figure H-15. ESEM image magnified 100 times for the interior of a Test#3, Day-30 raw cal-sil sample submerged in the birdcage. (T3Call24)



Figure H-16. ESEM image magnified 500 times for the interior of a Test#3, Day-30 raw cal-sil sample submerged in the birdcage. (T3Call25)



Figure H-17. Annotated ESEM image magnified 500 times for the interior of a Test#3, Day-30 raw cal-sil sample submerged in the birdcage. (T3Call25)



Figure H-18. EDS counting spectrum for the particles shown in Figure H-17. (T3cali26)



Figure H-19. ESEM image magnified 500 times for the interior of a Test#3, Day-30 raw cal-sil sample submerged in the birdcage. (T3Call27)



Figure H-20. ESEM image magnified 100 times for the exterior of a Test#3, Day-30 submerged high-flow baked cal-sil sample. (T3bcal39)



Figure H-21. ESEM image magnified 1000 times for the exterior of a Test#3, Day-30 submerged high-flow baked cal-sil sample. (t3bcal40)



Figure H-22. Annotated ESEM image magnified 1000 times for the exterior of a Test#3, Day-30 submerged high-flow baked cal-sil sample. (t3bcal40)



Figure H-23. EDS counting spectrum for the light particles (EDS1) shown in Figure H-22. (T3bcal41)



Figure H-24. EDS counting spectrum for the dark particles (EDS2) shown in Figure H-22. (t3bcal42)



Figure H-25. ESEM image magnified 100 times for the interior of a Test#3, Day-30 submerged high-flow baked cal-sil sample. (T3Bcal43)



Figure H-26. ESEM image magnified 1000 times for the interior of a Test#3, Day-30 submerged high-flow baked cal-sil sample. (t3bcal44)



Figure H-27. ESEM image magnified 1000 times for the interior of a Test#3, Day-30 submerged high-flow baked cal-sil sample. (t3bcal45)






Figure H-29. XRD results for the unused raw cal-sil sample.



Figure H-30. XRD results for the unused baked cal-sil sample.

 Table H-5.
 Dry Mass Composition of Unused Cal Sil Samples by XRF Analysis

Sample ID	SiO <sub>2</sub>	TiO <sub>2</sub>	Al <sub>2</sub> O <sub>3</sub>	Fe <sub>2</sub> O <sub>3</sub>	FeO	MnO	MgO	CaO	Na <sub>2</sub> O	K <sub>2</sub> O	H <sub>2</sub> 0(-	H <sub>2</sub> O(+) CO <sub>2</sub>	P <sub>2</sub> O <sub>5</sub>	Total	H2O(+)CO2 /DF (10) & Cover. To %
Baked Cal Sil	38.34	0.18	5.02	2.54	0.00	0.06	0.79	34.76	2.32	0.42	0.35	18.75	0.15	103.67	1.0191
Raw Cal Sil	33.87	0.36	4.27	2.07	0.00	0.05	1.35	34.66	2.27	0.35	0.56	1.59	0.12	81.50	1.0016

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# Appendix I

# ESEM/EDS Data for Test #3, Day-30 Sediment

## Figures

Figure I-1.	ESEM image magnified 1000 times for Test #3, Day-30 pink
	sediment. (t3PNKP29)I-4
Figure I-2.	Annotated ESEM image magnified 1000 times for Test #3, Day-30
	pink sediment. (t3PNKP29)I-4
Figure I-3.	EDS counting spectrum for the particles shown in Figure I-2.
	(t3pnkp30)I-5
Figure I-4.	ESEM image magnified 100 times for Test #3, Day-30 pink sediment.
	(t3pnkp31)I-5
Figure I-5.	ESEM image magnified 100 times for Test #3, Day-30 pink sediment.
	(t3pnkp32)l-6
Figure I-6.	ESEM image magnified 100 times for Test #3, Day-30 yellow
	sediment. (t3ylwp34)I-6
Figure 1-7.	Annotated ESEM image magnified 100 times for Test #3, Day-30
	yellow sediment. (t3ylwp34)I-7
Figure I-8.	EDS counting spectrum for the particles shown in Figure I-7.
	(t3ylwp35)I-7
Figure I-9.	Comparison of EDS counting spectra for pink sediment (yellow,
	t3pnkp30) and yellow sediment (red, t3ylwp35). (t3ylwp36)I-8
Figure I-10.	ESEM image magnified 100 times for Test #3, Day-30 yellow
	sediment. (t3ylwp37)I-8
Figure I-11.	XRD results forTest #3, Day-30 sedimentI-9

### Tables

Table I-1.	Dry Mass Composition of a Test #3, Day-30 Sediment by XRF	
	AnalysisI	-9

I-i

Particulate sediments at the bottom of the tank directly relate to the corrosion products and debris generated during ICET. This appendix lists the ESEM/EDS and XRD/XRF results for the sediment samples collected from the bottom of the tank on the date Test #3 was shut down (May 5, 2005). The sediment samples were identified and classified by their pink or yellow color. The purpose of these analyses is to provide information on the morphology and composition of these sediments.

ESEM was used to examine the sediment samples without any coating. EDS results provide an elemental composition of the sediment. Based on XRD results, the sediment sample contained crystalline substances of tobermorite  $[Ca_{2.25}(Si_3O_{7.5}(OH)_{1.5})(H_2O)]$  and calcite (CaCO<sub>3</sub>), the same as unused raw or unused baked cal-sil samples. XRF results show the chemical composition of the sediment. Available logbook entries for this laboratory session are included in this appendix as transcribed notes.

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Laboratory session from May 6, 2005. Test #3, Day-30 Sediment.



#### **Pink Sediment**

Image:	t3PNKP29	$1000 \times$	ESEM image	Figure I-1
	t3PNKP29	$1000 \times$	Annotated ESEM image	Figure I-2
EDS:	t3pnkp30		EDS on image 29	Figure I-3
Image:	t3pnkp31	100 ×	ESEM image	Figure I-4
	t3Pnkp32	100 ×	ESEM image	Figure I-5

#### **Yellow Sediment**

Image	t3ylwp34	$100 \times$	ESEM image	Figure I-6
	t3ylwp34	100 ×	Annotated ESEM image	Figure I-7
EDS:	t3ylwp35		EDS on image 34	Figure I-8
	t3ylwp36		Comparing EDS image t3ylwp35 with pink sediment EDS image t3pnkp30	Figure I-9
Image:	t3ylwp37	100 ×		Figure I-10



Figure I-1. ESEM image magnified 1000 times for Test #3, Day-30 pink sediment. (t3PNKP29)



Figure I-2.

Annotated ESEM image magnified 1000 times for Test #3, Day-30 pink sediment. (t3PNKP29)







Figure I-4. ESEM image magnified 100 times for Test #3, Day-30 pink sediment. (t3pnkp31)



Figure 1-5. ESEM image magnified 100 times for Test #3, Day-30 pink sediment. (t3pnkp32)









Annotated ESEM image magnified 100 times for Test #3, Day-30 yellow sediment. (t3ylwp34)







Figure I-9. Comparison of EDS counting spectra for pink sediment (yellow, t3pnkp30) and yellow sediment (red, t3ylwp35). (t3ylwp36)



Figure I-10. ESEM image magnified 100 times for Test #3, Day-30 yellow sediment. (t3ylwp37)



Figure I-11. XRD results forTest #3, Day-30 sediment.

Table I-1.	Dry	Mass	Composition	of	a	Test	#3,	Day-30	Sediment	by	XRF
	Anal	lysis									

Sample ID	SiO <sub>2</sub>	TiO <sub>2</sub>	Al <sub>2</sub> O <sub>3</sub>	Fe <sub>2</sub> O <sub>3</sub>	FeO	MnO	MgO	CaO	Na <sub>2</sub> O	K <sub>2</sub> O	H <sub>2</sub> 0(-	H <sub>2</sub> O(+) CO <sub>2</sub>	P <sub>2</sub> O <sub>5</sub>	Total	H2O(+)CO2 /DF (10) & Cover. To %
Test #3, Day 30 Sedi- ment	36.20	0.20	4 92	2 24	0.00	0.06	0.62	27.16	2 19	0.45	0.58	20.65	3.05	98.42	1 0211

1-9

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# Appendix J

# **TEM Data for Test #3 Solution Samples**

## Figures

Figure J-1.	Electron micrograph magnified 2000 times for one Test #3, Day-4
	filtered sample location. (T3D4F1)J-4
Figure J-2.	TEM energy-dispersive x-ray spectrum for the Day-4 filtered test
	sample as shown in Figure J-1. (T3D4FEDS) J-4
Figure J-3.	TEM image for Test #3, Day-4 filtered sample solution (T3D4F2) J-5
Figure J-4.	Electron micrograph magnified 2000 times for one Test #3, Day-4
	unfiltered sample location. (T3D4UF2)J-5
Figure J-5.	TEM energy-dispersive x-ray spectrum for the Day-4 unfiltered test
	sample as shown in Figure J-4. (T3D4UFEDS) J-6
Figure J-6.	TEM image for Test #3, Day-4 unfiltered sample solution (T3D4UF) J-6
Figure J-7.	Electron micrograph magnified 4000 times for one Test #3, Day-15
	unfiltered sample location. (T3D15U-4k-01) J-7
Figure J-8.	TEM energy-dispersive x-ray spectrum for Test #3, Day-15 unfiltered
	test sample as shown in Figure J-7. (T3D15UFEDS) J-7
Figure J-9.	TEM image magnified 2000 times for a Test #3, Day-15 unfiltered
	sample solution (T3D15U-SAD20cm-01) J-8
Figure J-10.	Electron micrograph magnified 4000 times for one Test #3, Day-30
	unfiltered sample location. (T3D30U-4k-01) J-8
Figure J-11.	TEM energy-dispersive x-ray spectrum for Test #3, Day-30 unfiltered
	test sample as shown in Figure J-10. (T3D30UFEDS) J-9
Figure J-12.	TEM image magnified 2000 times for a Test #3, Day-30 unfiltered
	sample solution (T3D30U-SAD) J-9

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This appendix presents TEM images, EDS, and diffraction patterns for Test #3, Day-4, Day-15, and Day-30 filtered and unfiltered solution samples. The filtered solution samples were passed through a 0.7-µm fiberglass filter at 60°C. The unfiltered solution samples were extracted from the tank directly. Based on the results, no significant diffraction pattern was observed because of the amorphous nature of the samples. Available logbook entries for this laboratory session are included in this appendix as transcribed notes.

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#### **Transcribed Laboratory Log**

#### Laboratory session from May 6, 2005.

TEM Test #3, Day-4 solution samplesTest #3, Day-15 filtered and unfiltered solutionTest #3, Day 30 unfiltered solution

#### **TEM Filtered Day-4 Solution Samples**

Image:	T3D4F1	2000 ×		Figure J-1
EDS:	T3D4FEDS		Spectrum for image T3D4F1	Figure J-2
Image:	T3D4F2	2000 ×		Figure J-3

#### **TEM Unfiltered Day-4 Solution Samples**

Image:	T3D4UF2	2000 ×		Figure J-4
EDS:	T3D4UFEDS		Spectrum on image T3D4UF2	Figure J-5
Image:	T3D4UF	2000 ×		Figure J-6

#### **TEM Unfiltered Day-15 Solution Samples**

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Image:	T3D15U-4k-01	$4000 \times$		Figure J-7
EDS:	T3D15UFEDS	· .	Spectrum on image T3D15U- 4k-01	Figure J-8
Image:	T3D15U-SAD20cm- 01	2000 ×		Figure J-9

#### TEM Unfiltered Day-30 Solution Samples

Image:	T3D30U-4k- 01	4000 ×		Figure J-10
EDS:	T3D30UFEDS		Spectrum on image T3D30U-4k-01	Figure J-11
Image:	T3D30U- SAD_	2000 ×		Figure J-12



Figure J-1. Electron micrograph magnified 2000 times for one Test #3, Day-4 filtered sample location. (T3D4F1)



Figure J-2. TEM energy-dispersive x-ray spectrum for the Day-4 filtered test sample as shown in Figure J-1. (T3D4FEDS)



Figure J-3. TEM image for Test #3, Day-4 filtered sample solution (T3D4F2)



Figure J-4. Electron micrograph magnified 2000 times for one Test #3, Day-4 unfiltered sample location. (T3D4UF2)



Figure J-5. TEM energy-dispersive x-ray spectrum for the Day-4 unfiltered test sample as shown in Figure J-4. (T3D4UFEDS)



Figure J-6. TEM image for Test #3, Day-4 unfiltered sample solution (T3D4UF)



Figure J-7. Electron micrograph magnified 4000 times for one Test #3, Day-15 unfiltered sample location. (T3D15U-4k-01)



Figure J-8. TEM energy-dispersive x-ray spectrum for Test #3, Day-15 unfiltered test sample as shown in Figure J-7. (T3D15UFEDS)



Figure J-9. TEM image magnified 2000 times for a Test #3, Day-15 unfiltered sample solution (T3D15U-SAD20cm-01)



Figure J-10. Electron micrograph magnified 4000 times for one Test #3, Day-30 unfiltered sample location. (T3D30U-4k-01)



Figure J-11. TEM energy-dispersive x-ray spectrum for Test #3, Day-30 unfiltered test sample as shown in Figure J-10. (T3D30UFEDS)



Figure J-12. TEM image magnified 2000 times for a Test #3, Day-30 unfiltered sample solution. (T3D30U-SAD)

## Appendix K

# UV Absorbance Spectrum—Day-30 Solution Samples

### Figures

Figure K-1. UV absorbance spectrum for Test #3, Day-30 solution samples...... K-3

#### Tables

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This appendix presents the ultraviolet (UV) absorbance result of the Test #3, Day-30 solution sample. The purpose of this analysis was to find any distinguishing absorbance peaks to identify the organics present in the solution. The solution sample at 60°C was collected through a 0.7-µm fiberglass filter to remove particulate impurities, followed by being scanned over the wavelength ranging from 200 to 800 nm by a UV-visible spectrophotometer. The spectrum of deionized water was used as background subtraction. From the result, the test solution did not exhibit any distinguished absorbance peaks. No logbook entries are available for this laboratory session, so, no transcribed notes are provided in this appendix.

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Figure K-1. UV absorbance spectrum for Test #3, Day-30 solution samples.

Test #3, Day-30 Samples	
Collection Time:	5/5/2005 11:33:48 AM
Operator Name:	1 
Scan Software Version:	3.00(182)
Parameter List :	
Instrument:	Cary 50
Instrument Version:	3.00
Start (nm):	800.0
Stop (nm):	200.0
X Mode:	Nanometers
Y Mode:	Abs
UV-Vis Scan Rate (nm/min):	600.00
UV-Vis Data Interval (nm):	1.00
UV-Vis Ave. Time (s):	0.1000
Beam Mode:	Dual beam
Baseline Correction:	On
Baseline Type:	Baseline correction
Baseline File Name:	· · · · · · · · · · · · · · · · · · ·
Baseline Std Ref File Name:	
Cycle Mode:	Off
Comments:	
Method Log:	
Method Name:	Default
Date/Time Stamp:	5/5/2005 11:28:54 AM
Method Modifications:	
Cell Changer $6 \times 6$ Changed:	5/5/2005 11:28:58 AM / Old:1 / New:0
UVVIS SAT Changed:	5/5/2005 11:29:22 AM / Old:0.0125 / New:0.1000
NIR SAT Changed:	5/5/2005 11:29:22 AM / Old:0.0125 / New:0.1000
Common SAT Changed:	5/5/2005 11:29:22 AM / Old:0.0125 / New:0.1000
Baseline Correction Changed:	5/5/2005 11:29:22 AM / Old:0 / New:1
Temp Controller Changed:	5/5/2005 11:29:26 AM / Old:0 / New:2

 Table K-1.
 Test #3, Day-30 Solution Sample Laboratory Settings

Sipper Type Changed:	5/5/2005 11:30:11 AM / Old:Internal RSA / New:External sipper
End Method Modifications	
<current wavelength=""></current>	200.1

K-5

and the second sec
# Appendix L

# ICET Test #3: Pre-Test, Test, and Post-Test Project Instructions

L-i

The ICET series is conducted under the guidance of PIs, which identify the steps to follow for certain activities. These PIs are revised or rewritten as needed for each test. For Test #3, a new PI was written to address test operations. The PIs that address pre-test and post-test operations were revised for Test #3. These three PIs are included in this appendix to describe more completely the test apparatus and chemical solution preparations, the test startup and daily sampling, and the steps followed after test shutdown.

L-l

# **1.0 INTRODUCTION**

#### **1.1 PURPOSE**

The purpose of this instruction is to ensure that all data acquisition, test samples, testing supplies, chemicals, and related materials are ready and accounted for prior to testing. In addition, this instruction provides instructions on preparing the chemical test apparatus for testing.

#### 1.2 SCOPE

The pre-test operations preparation will ensure that successful initiation of the testing activity is achieved.

## **1.3 REFERENCES**

- Test Plan: Characterization of Chemical and Corrosion Effects Potentially Occurring Inside a PWR Containment Following a LOCA, Revision 12.c, March 30, 2005
- Test 2 Chemical Additive Analysis ICET-CALC-011
- Laboratory Safety Guidelines
- ASTM A 380 99, Standard Practice for Cleaning, Descaling, and Passivation of Stainless Steel Parts, Equipment, and Systems
- Material Safety Data Sheets (MSDS) for all chemicals involved

## 2.0 **PREREQUISITES**

The data acquisition setup and inspection; instrument calibration; and the coupon receipt, preparation, inspection, and storage tasks must be completed in full prior to the completion of this activity. Fiberglass and calcium silicate (cal-sil) samples must be weighed and their planned locations in the tank identified. That data must be recorded.

#### 2.1 Training Requirements

The following personnel training is required for this task:

- 1) LabVIEW and computer data acquisition training
- 2) Chemical handling training, specifically for ethyl alcohol, ammonium hydroxide, and lithium hydroxide.
- 3) Safe lift execution training

# 2.2 Equipment Requirements

The following equipment is required to perform this activity: computer with installed LabVIEW software, data acquisition system, and fully assembled and calibrated ICET test apparatus.

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Safety equipment must be available: goggles, gloves, lab coats, eye wash station.

## **3.0 DOCUMENTATION REQUIRED**

MSDSs must be available for all chemicals used.

A lab notebook must be maintained throughout the pre-test operations instruction. Contained within the lab notebook will be the date, times, description of activities, and quantities of chemicals added, number of cleanings, and physical observations of the tank cleaning and preparation procedures.

## 4.0 HAZARDS

The hazards associated with this activity include potential injuries associated with chemical handling.

# 5.0 **INSTRUCTIONS**

- Ensure that all testing materials and supplies are ready and on-site. See checklist at the end of this document. Verify that eye wash station is operational. Note: The following solutions are not used in this instruction, but are to be prepared in advance of entering ICET-PI-014, "Test Operations, Test #3 (cal-sil and fiberglass with TSP)." After preparation, clearly label the containers with the solutions and place in an area restricted for ICET Project test use.
- 2. Prepare 21.2 g of concrete dust and 63.7 g of latent debris.
- 3. Prepare LiOH solution: dissolve 0.663 g of lithium hydroxide (LiOH) into about 100 mL water in a 250-mL sample container.
- 4. Prepare TSP Solution Batch 1.
  - a. Heat about 1.5 gallons of demineralized water, add 300 g of boric acid (H<sub>3</sub>BO<sub>3</sub>), and stir until the boric acid is dissolved. Pour the solution into a 5-gallon plastic container. Dissolve the boric acid in multiple batches if necessary.
  - b: Add additional demineralized water to the 5-gallon plastic container until it contains about 4 gallons.
  - c. Dissolve 1893 g of TSP (Na<sub>3</sub>PO<sub>4</sub>-12H<sub>2</sub>O) into the water in the container.
  - d. Dilute with additional demineralized water until the volume is 5 gallons.
  - e. Label container as "TSP Solution Batch 1."
- 5. Prepare TSP Solution Batch 2.
  - a. Heat about 1.5 gallons of demineralized water, add 300 g of boric acid (H<sub>3</sub>BO<sub>3</sub>), and stir until the boric acid is dissolved. Pour the solution into a 5-gallon plastic container. Dissolve the boric acid in multiple batches if necessary.

- b. Add additional demineralized water to the 5-gallon plastic container until it contains about 4 gallons.
- c. Dissolve 1893 g of TSP (Na<sub>3</sub>PO<sub>4</sub>-12H<sub>2</sub>O) into the water in the container.
- d. Add 211 mL of 12.29 N hydrochloric acid (HCl) to the water in the container.
- e. Dilute with additional demineralized water until the volume is 5 gallons.
- f. Label container as "TSP Solution Batch 2."
- 6. Prepare laboratory control sample (LCS). See ICET-PI-005, "Chemical Sampling and Analysis," for details on the laboratory control sample.
- 7. Start the data acquisition system. Verify that the data acquisition system is monitoring flow rate, pump speed, temperature, and pH.
- 8. Clean the tank and piping.
  - a. Cleaning should commence as soon after a test is completed as possible, to prevent material from hardening in the tank or piping and to maximize the time available for cleaning.
  - b. Cleaning chemicals may consist of weak acids (e.g., acetic acid, citric acid, or dilute mineral acids), weak bases (e.g., ammonium hydroxide), week organic solvents (e.g., ethanol), or detergents/surfactants (e.g., trisodium phosphate, sodium dodecyl sulfate), as necessary. Cleaning solutions can be heated if necessary. Note that the discharge limit to the sanitary sewer is a maximum temperature of 140 °F and pH between 5.0 and 11.5. Cleaning solutions that are not within this range should be neutralized before discharge.
  - c. During cleaning, the pump should be run and water directed through both recirculation lines (through the spray nozzles and lower headers)
  - d. The sample line should be removed from the piping, physically cleaned, and carefully inspected. If the sample line cannot be adequately cleaned, it should be replaced.
  - e. After each cleaning step, the tank and piping should be thoroughly rinsed with tap water or demineralized water.
  - f. After each cleaning step, a segment of pipe should be removed, and the interior of the pipe visually inspected.
  - g. Cleanliness criteria: When the tank visually appears to be satisfactorily cleaned, the tank and piping should be thoroughly rinsed with demineralized water. The interior surfaces of the tank and piping shall be free of any deposits that can be removed by vigorous scrubbing. Demineralized water drained from the tank should have turbidity less than 0.3 NTU and conductivity less than 50 uS/cm.
- 9. Tank is now ready for testing. Proceed immediately to Instruction No. ICET-PI-014, "Test Operations, Test #3 (cal-sil and fiberglass with TSP).

# 6.0 **ATTACHMENTS**

No forms are attached to this document.

# 7.0 Materials Checklist

- \_\_\_\_\_ lithium hydroxide, 0.663 g
- \_\_\_\_\_ TSP, 3.785 kg
- \_\_\_\_\_ 211 mL of 12.29 N HCl
- \_\_\_\_\_ Boric acid, 600 g
- \_\_\_\_\_ tap water supply
- \_\_\_\_\_ demineralized water production system
- chemical handling safety equipment (lab coat, goggles, rubber gloves)
- \_\_\_\_\_ analytical balance
- \_\_\_\_\_ top loading balance
- \_\_\_\_\_ chemical spatula
- \_\_\_\_\_ chemical scoop
- \_\_\_\_\_ weigh boats
- \_\_\_\_\_ two 5-gallon plastic containers
- \_\_\_\_\_ 250 mL graduated cylinder
- \_\_\_\_\_ 250-mL HDPE or PP bottle
- 2.5 gallons ethanol
- 2.5 gallons ammonium hydroxide
- \_\_\_\_\_ turbidimeter and associated equipment
- \_\_\_\_\_ conductivity meter and associated equipment

# **1.0 INTRODUCTION**

### **1.1 PURPOSE**

The intent of the instruction is to outline the steps that are to be followed during testing.

# 1.2 SCOPE

This activity forms the core of the entire Chemical Effects Testing project. All activities involved in this project affect and are affected by this activity.

# **1.3 REFERENCES**

- Test Plan: Characterization of Chemical and Corrosion Effects Potentially Occurring Inside a PWR Containment Following a LOCA, Revision 12.c, March 30, 2005
- ASTM Standard G 4-01
- ASTM Standard D 3370-95a
- ASTM Standard G 31-72
- Material Safety Data Sheets (MSDS) for all chemicals involved
- LabVIEW operation manual
- Laboratory Safety Guidelines
- Test 2 Chemical Additive Analysis ICET-CALC-011
- John Gisclon email to Bhagwat Jain, Cal-sil Information Used in Test #3, March 31, 2005

# 2.0 PREREQUISITES

All sample coupons must be placed in their corresponding racks. Also, the preoperation test preparation activity must be completed in full.

## 2.1 Training Requirements

The following personnel training is required for this task:

- 1) LabVIEW and computer data acquisition training.
- 2) Chemical handling training for all chemicals involved.

#### 2.2 Equipment Requirements

The following equipment is required to perform this activity: computer with installed LabVIEW software, data acquisition system, and fully assembled and calibrated ICET test apparatus.

Safety equipment must be available: goggles, gloves, lab coats, hard hats, steel-toed shoes, eye wash station, hydrogen detector and hydrogen removal system.

# **3.0 DOCUMENTATION REQUIRED**

A lab notebook must be maintained throughout the testing procedure. In addition, a binder will be maintained that includes pertinent test instructions and the completed daily log sheets (see Attachment A). The daily log sheet contains the date, times, physical description, and quantity of fiberglass and water samples obtained each day. In addition, the daily log sheet contains information from the data acquisition system (DAS), the water samples taken, and other test information.

The electronic data that are acquired are backed up daily and stored in a separate location each testing day. Refer to ICET-PI-001, Data Acquisition Setup and Inspection.

## 4.0 HAZARDS

The hazards associated with this activity include tipping of the chemical tank assembly, ingestion and/or respiration of any chemicals involved, and scalding and/or burning hazards involved in daily tank venting, and possible hydrogen gas generation from corrosion reactions. Appropriate measures to control hydrogen gas must be in place before operations commence.

Lifting hazards associated with the tank lid and coupon racks are also associated with this activity.

## 5.0 INSTRUCTIONS

- Because of the time required for heating the tank contents and dissolving chemicals, this sequence should be started at least 48 hours before the scheduled time t = 0. Pre-Test Operations preparation should be complete before proceeding with this sequence.
- 2. Ensure that all testing materials and supplies are ready and on-site (see checklist at end of this instruction).
- 3. Add 240 gallons of RO water to the tank by pumping water from the RO skid through the totalizing flow meter. Record flow to the nearest 0.5 gallon.
- 4. Verify valves are positioned as follows:

Test Operations, Test #3 (cal-sil and fiberglass, with TSP) ICET-

ICET-PI-014, Rev 0 04/05/05 Page 3 of 7

Valve	Description	Position
V-1	tank drain	closed
V-2	pump isolation	open
V-3	instrument loop supply	open
V-4	instrument loop discharge	open
V-5	instrument loop bypass	closed
V-6	in-line filter isolation	open
V-7	tank spray supply	closed
V-8	recirculation supply	open
V-9	sample line	closed
V-10	loop drain	closed
V-11	recirculation line injection	closed

- 5. Start pump and adjust to flow rate of approximately 25 gpm.
- 6. Start computer, start LabVIEW, verify that flow rate, pump speed, temperature, and pH are being recorded properly.
- 7. Turn on heater and allow water in tank to heat to  $60 \degree C \pm 2 \degree C$ . (This may take up to 20 hours.)
- 8. Add the pre-mixed LiOH solution.
- 9. Add 14.54 kg of boric acid (H<sub>3</sub>BO<sub>3</sub>), weighing in approx. 2 kg increments, recording the weight of each increment to the nearest 10 g.
- 10. Allow the water to circulate until the solution is visibly clear, indicating that the boric acid is completely dissolved.
- 11. Allow water in tank to heat to 65 °C  $\pm$  2 °C.
- 12. Take grab water sample for analysis for the parameters identified in steps a h below. Also record physical appearance of the sample (clarity, presence of gelatinous material, etc). All Day 1 and subsequent samples will be analyzed by Assaigai Analytical Laboratory. In addition, periodic test samples and laboratory control samples (LCSs) will also be analyzed by the UNM laboratory.
  - a. pH
  - b. temperature
  - c. turbidity
  - d. viscosity
  - e. total suspended solids (TSS)
  - f. dissolved oxygen (DO)
  - g. chloride
  - h. metals (Al, B, Ca, Cu, Fe, Pb, Li, Mg, Ni, K, Si, Na, and Zn), total and dissolved
- 13. Add 21.2 g of concrete dust and 63.7 g of latent debris samples (prepared earlier), wait 10 minutes, take 100 mL water sample for particulate size distribution, density, and TSS.
- 14. Stop pump.
- 15. Add the pre-determined amount of cal-sil dust. This will be approximately 43.5 lb.

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- 16. Place coupon racks, fiberglass holders, and cal-sil holders into tank. This is done in accordance with previously determined quantities, size distributions, and locations. (Details of the cal-sil preparation and size distributions are given in the referenced email.)
- 17. Verify locations of coupon racks, fiberglass holders, and cal-sil holders.
- 18. Verify the tank temperature is 62 °C. (Because the tank lid will be off, the test will be started with the water temperature at its upper limit.)
- 19. Start pump and adjust pump speed to 25 gpm.
- Open valve V-7 (tank spray supply) to direct water to nozzles and adjust valves V-7 (tank spray supply) and V-8 (recirculation supply) until nozzle flowmeter is reading 3.5 gpm. Verify total flow is still 25 gpm and adjust variable frequency drive (VFD) if necessary.
- 21. Record date and time at which nozzle flow started. This is time t = 0 for the test.
- 22. The spray phase will begin with the tank lid off. The objective is to be able to carefully monitor possible nozzle blockage and take immediate action to prevent it. At the first sign that a nozzle may be starting to plug, the spray flow rate should be increased rapidly to 5-10 gpm for approximately 5 s. (As long as the nozzle spray pattern is not affected however, the spray flow should remain at 3.5 gpm.) If a nozzle should block in spite of the increased flow rate, a stainless steel wire should be used to clear the nozzle exit.
- 23. If the spray through the nozzles is not affected by the cal-sil and shows no signs of blocking for a period of 15 minutes, the tank lid should be put in place. Flow through the nozzles should continue to be monitored every 5 minutes by looking through the tank view windows. In addition, every 15 minutes, the spray flow should be bumped to 5-10 gpm for approximately 5 s.
- 24. At 30 minutes, open valve V-11 (recirculation line injection) and start chemical metering pump from TSP Solution Batch 1 at a rate of 0.0476 gpm (180 mL/min). The objective here and in step #25 is to add a total of 10 gallons of TSP solution in 3.5 hours.
- 25. After 2 hours, switch the chemical metering pump to TSP Solution Batch 2.
- 26. Take a measurement of hydrogen concentration. At 2-hour increments, repeat the hydrogen concentration measurement. If the concentration reaches 10% of the flammability limit, purge the tank atmosphere. This needs to be repeated until the hydrogen concentration has been determined to be below 10% of the flammability limit, and then the frequency of hydrogen concentration measurements is to be re-evaluated.
- 27. At t = 4 hours, stop the chemical metering pump and close valves V-7 and V-11.
- 28. Immediately after closing valves V-7 and V-11 (at t = 4 hours), take water grab sample for analysis for the parameters listed below. Record the time of sample collection.
  - a. pH
  - b. temperature
  - c. turbidity

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- d. viscosity
- e. chloride
- f. total suspended solids (TSS)
- g. dissolved oxygen (DO)
- h. metals (Al, B, Ca, Cu, Fe, Pb, Li, Mg, Ni, K, Si, Na, and Zn), total and dissolved
- 29. At t = 24 hours, and daily thereafter, take water grab sample for analysis for the parameters listed below. (The LANL PI will propose a different sampling frequency to the project sponsors if test data support it.) Record the time of sample collection.
  - a. pH
  - b. turbidity
  - c. viscosity
  - d. temperature
  - e. total suspended solids (TSS)
  - f. metals (Al, B, Ca, Cu, Fe, Pb, Li, Mg, Ni, K, Si, Na, and Zn), total and dissolved. An exception is that B, Li, K, Pb, and chloride analyses will be performed only at t = days 15 and 30. Also, dissolved oxygen will be measured at day 30.
- 30. During each daily water sample collection, look inside tank (through windows) and record observations. If the tank water level indicates that the water volume is 245 gallons or less, add RO water to bring the volume up to 250 gallons and record the amount added.
- 31. At t = 24 hours, weekly thereafter, and at the end of the test, collect 100 mL water sample for particulate size distribution and density analysis, to be performed at AALI. The particulate size ranges to be used will be as close as possible to those called out in the test plan: (in microns), 1-10, 11-25, 26-50, 51-75, 76-100, and > 100 microns.
- 32. At t = 24 hours, weekly thereafter, and at the end of the test, collect water samples for strain rate viscosity measurements (see PI-010 for sample details.)
- 33. At 3 days  $\leq t \leq 5$  days, 14 days  $\leq t \leq 16$  days and at the end of the test, collect a sacrificial fiberglass sample to be inspected and examined with SEM.
- 34. At 24 hours, at 14 days ≤ t ≤ 16 days and at the end of the test, run 1L of water through a nucleopore filter. The filter will be taken for SEM analysis as specified in ICET-PI-007. (Note that depending on the solution, some filter material will not work well for this operation. If possible, use a nucleopore filter for SEM analysis, and then collect a second sample on nitrocellulose filter for later digestion and ICP analysis.)
- 35. Shut down pump
- 36. Indicate end of test on the data acquisition system and shut down the data acquisition software.
- 37. Proceed directly to PI-008 Post-Test Operations.

## 6.0 ATTACHMENTS

Attachment A. Daily Log Sheet.

# 7.0 MATERIAL CHECKLIST

- \_\_\_\_\_ boric acid, 14.54 kg
- \_\_\_\_\_ pre-mixed lithium hydroxide solution
- \_\_\_\_\_ concrete dust, 21.2 g
- \_\_\_\_\_ latent debris, 63.7 g
- \_\_\_\_\_ Nucleopore filter
- TSP, 3786 g evenly mixed in two 5-gallon containers
- \_\_\_\_\_ chemical handling safety equipment (lab coat, goggles, rubber gloves)
- \_\_\_\_\_ top-loading balance
- \_\_\_\_\_ weigh pan for 2 kg aliquots of boric acid
- \_\_\_\_\_ stainless steel filter paper holder
- \_\_\_\_\_ 500 mL graduated cylinder (for TSS)
- \_\_\_\_\_ totalizing flow meter
- \_\_\_\_\_\_ sample containers (see Chemical Sampling Instruction)
- \_\_\_\_\_ analytical equipment (see Chemical Sampling Instruction)
- pre-assembled coupon racks
- pre-assembled fiberglass baskets, total of 2.2 lb of fiberglass
- pre-assembled cal-sil baskets, total of 26.7 lb of cal-sil
- pre-measured cal-sil dust, 43.5 lb
- coupon handling safety equipment (hard hat, leather gloves, boots)
- \_\_\_\_\_ computer disks for backup of Labview data
- \_\_\_\_\_ Masterflex peristaltic pump and tubing
- \_\_\_\_\_ demineralized water production system

Daily I Integrated Chemica	Log Sheet I Effects Test (Test # 2) lection:
Sample taking and data reduction by	and
Sample bottle identification: Assaigai (total): Assaigai (filtered): UNM (total): UNM (filtered):	
Control system readings: Temperature: Flow:	pH:
Analyses:   Volume filtered for TSS:   Temperature:   Turbidity (at 60 °C):   Viscosity, unfiltered (60 °C):   Viscosity, filtered (60 °C):   Water Level:   Hydrogen:   Fiberglass or other samples taken:   TSS filter #:	pH: Dissolved oxygen: (at 23 °C; and 10 min.) (at 23 °C) (at 23 °C) (at 23 °C) Water Added: Other: TSS (mg/L):
Comments:	

Observations written in lab notebook by \_\_\_\_\_

# **1.0 INTRODUCTION**

# 1.1 PURPOSE

The intent of this instruction is to ensure that the experimental samples are removed from the test apparatus, the test apparatus is rinsed and inspected, and the test apparatus is made ready for subsequent pre-test operations.

# 1.2 SCOPE

This activity marks the end of one chemical effects test run. Experimental sample removals and inspections, test apparatus rinsing, and preparations for cleaning and subsequent tests are addressed here.

# **1.3 REFERENCES**

- Test Plan: Characterization of Chemical and Corrosion Effects Potentially Occurring Inside a PWR Containment Following a LOCA, Revision 12.c, March 30, 2005
- ASTM Standard G 4-01
- ASTM Standard G 31-72
- ICET-PI-002, Coupon Receipt, Preparation, Inspection, and Storage, November 19, 2004
- ICET-PI-014, Rev. 0, Test Operations, Test #3 (cal-sil and fiberglass, with TSP, April 5, 2005
- ICET-PI-005, Rev. 1, Chemical Sampling and Analysis, February 3, 2005
- Laboratory safety guidelines
- ICET Project Safety Plan

## 2.0 **PREREQUISITES**

All test operation PI criteria must be completed prior to conducting this task.

## 2.1 Training Requirements

- Laboratory Safety Guidelines
- ICET Project Safety Plan

# 2.2 Equipment Requirements

A city tap water supply outlet is required for this activity and chemical handling and lifting safety equipment. A reverse osmosis unit is required for the final flush.

# **3.0 DOCUMENTATION REQUIRED**

Documentation related to test parameters, chemical water analyses, coupon and fiberglass examinations, and daily test operations are outlined elsewhere. In this instruction, the steps required to remove samples from the test apparatus and to make it ready for the next test are outlined. In addition, observations as to the test apparatus' condition are obtained and recorded here.

## 4.0 HAZARDS

The hazards associated with this activity include ingestion/respiration and/or dermal and eye contact with residual chemicals. Lifting hazards associated with the tank lid and coupon racks are also associated with this activity.

# 5.0 INSTRUCTIONS

- 1) On the last day of testing, collect water samples and perform analyses as outlined in ICET-PI-014 and ICET-PI-005.
- 2) Remove 10L of water from the test apparatus and store at test temperature, for future analyses
- 3) Shut off the recirculation pump.
- 4) Remove the small fiberglass samples for SEM examination.
- 5) Leave one heater on and continue to monitor tank water temperature.
- 6) Isolate and drain the test apparatus piping.
- 7) Remove the tank lid.
- 8) Before removing coupon racks or insulation samples, examine and take photographs and notes of the inside of the tank, the coupons and racks, and the insulation samples.
- 9) Remove the six non-submerged coupon racks to a staging area for drying and post-test examinations (refer to ICET-PI-002).
- 10) Take additional photographs of the inside of the tank.
- 11) Drain the tank slowly, down to the level that uncovers the submerged rack, but keeping the water level above the heater.
- 12) Remove the submerged coupon rack to the staging area.
- 13) Repeat step # 10.
- 14) Turn off the heater.
- 15) Completely drain the tank, taking precautions so that the sediment on the bottom of the tank is not disturbed any more than necessary.
- 16) Store water that was drained from the test apparatus until it is cleared for disposal or shipment. (This step was just moved from later in the PI – the old step #26.)
- 17) When the tank is drained, repeat step # 10. Note especially the locations and orientations of the remaining samples.
- 18) Remove the remaining insulation samples to the staging area to dry.

- 19) Ensure that all samples removed from the tank are clearly marked as to their location and orientation within the tank.
- 20) After all samples have been removed, repeat step # 10.
- 21) Inspect the interior of the tank, noting any observations.
- 22) Note the presence of any sediment. Carefully remove as much sediment as possible, noting any unique aspects of it, such as location. Place the sediment in plastic containers with lids, marking the location of the sediment in the tank.
- 23) Remove the tank drain screen and remove the insulation sample for future analysis.
- 24) Remove the flow meter from the loop and take pictures of the flow meter interior.
- 25) Remove any deposits within the flow meter and place the deposits in plastic containers with lids. This is to keep the samples hydrated.
- 26) Remove a section of pipe, take pictures of the pipe interior, and remove and store any deposits there.
- 27) Replace the flow meter and piping section.
- 28) Rinse the tank with tap water and drain the water.
- 29) Fill the system with 250 gallons of tap water and circulate water through the spray nozzles and recirculation headers for at least 60 minutes. Repeat with de-mineralized water.
- 30) If any signs of deterioration are observed on the inside of the test apparatus tank, remove selected insulation on the tank. Inspect the stainless steel tank for any abnormalities.

## 6.0 ATTACHMENTS

No forms are attached to this document.

NRCMD 3.7	U.S. NUCLEAR REGULATORY COMMISSION	1. REPORI NUMBER (Assigned by NRC, Add Vol., Supp., Rev., and Addendum Numbers, if any.)		
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Los Alamos National Laboratory PO Box 1663 Los Alamos, NM 87545	University of New Mexico Omicron Safety Department of Civil Engineering 2500 Louisiana Albuquerque, NM 87110 Albuquerque, N	ety & Risk Technologies, Inc. ana Blvd., Suite 410 e, NM 87199-3065		
Washington, DC 20555-0001 10. SUPPLEMENTARY NOTES B. P. Jain, NRC Project Manager; p	repared in cooperation with Electric Power Research Institu	te		
A 30-day test was conducted in the chemical environment present insid initial chemical environment contain hydrochloric acid (211 mL), and add the test. The test was conducted at included representative amounts of insulation samples (80% calcium si	Integrated Chemical Effects Test (ICET) project test appara le a pressurized water reactor containment water pool after a ned 14.54 kg of boric acid and 0.663 g of lithium hydroxide. T ditional boric acid (600 g) were added beginning at 30 minute a constant temperature of 60°C (140°F). The materials teste submerged and unsubmerged aluminum, copper, concrete, licate and 20% fiberglass). Representative amounts of concr	tus. The test simulat a loss-of-coolant-acc frisodium phosphate es and lasting until 4 ed within this environ zinc, carbon steel, a ete dust and latent a	ed the ident. The (3.786 kg), hours into ment ind	
also added to the test solution. The than 1 NTU after 24 hours. During t observed through the submerged o complete, and the white flocculence indicated similar behavior of the sol the water after Day 1, and no precip amounts of white deposits of varyin zinc coated steel coupons. The bott solution remained clearly Newtonian	test solution reached a pH of 7.9 by Day 3, and the test solution introduction of trisodium phosphate at the beginning of the beservation window. Turbidity and TSS initially rose, but drop was no longer visible in the water after the first day. Observation at both room temperature and test temperature. No chronitation occurred as samples cooled from test temperature to g size were observed on the submerged galvanized steel, al com of the tank was filled with sediment that had a pinkish-w n for the entire test. Aluminum was detectable in the solution	the test, a white floccu oped after chemical a vations of the test so emical byproducts w o room temperature. luminum, copper, an hite deposit on top. T h, but only in trace an	lebris were sed to less addition was lution ere visible in Large d inorganic The test nounts.	
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